

Provided by the author(s) and University of Galway in accordance with publisher policies. Please cite the published version when available.

Title	Sterile post-traumatic immunosuppression
Author(s)	Islam, Md Nahidul; Bradley, Benjamin A.; Ceredig, Rhodri
Publication Date	2016-04-29
Publication Information	Islam, Md Nahidul, Bradley, Benjamin A., & Ceredig, Rhodri. (2016). Sterile post-traumatic immunosuppression. Clin Trans Immunol, 5, e77. doi: 10.1038/cti.2016.13
Publisher	Nature Publishing Group
Link to publisher's version	http://dx.doi.org/10.1038/cti.2016.13
Item record	http://hdl.handle.net/10379/6289
DOI	http://dx.doi.org/10.1038/cti.2016.13

Downloaded 2024-04-29T04:01:46Z

Some rights reserved. For more information, please see the item record link above.



# STERILE POST-TRAUMATIC IMMUNOSUPPRESSION

Md Nahidul Islam, M.Sc., Ph.D.<sup>1</sup>, Benjamin A Bradley, MBChB, Ph.D.<sup>1,2</sup>, Rhodri Ceredig, MB. ChB., Ph.D., FRCPath<sup>1</sup>

<sup>1</sup> Regenerative Medicine Institute, National Centre of Biomedical Engineering Science and School of Medicine, Nursing and Health Sciences, National University of Ireland, Galway, Galway, Ireland.

<sup>2</sup> Orthopaedic Research Unit, Learning and Research Building, Southmead Hospital, University of Bristol, Bristol, United Kingdom.

# ABSTRACT

After major trauma, the human immune system initiates a series of inflammatory events at the injury site that is later followed by suppression of local inflammation favouring the repair and remodelling of the damaged tissues. This local immune response involves complex interactions between resident cells such as macrophages and dendritic cells, soluble mediators such as cytokines and chemokines, and recruited cells such as neutrophils, monocytes and mesenchymal stromal cells. If of sufficient magnitude, these initial immune responses nevertheless have systemic consequences resulting in a state called post-traumatic immunosuppression (PTI). However, controversy exists regarding the exact immunological changes occurring in systemic compartments triggered by these local immune responses. PTI is one of the leading causes of post-surgical mortality and makes patients vulnerable to hospital acquired infections, multiple organ failure, and many other complications. Additionally, haemorrhage, blood transfusion, immune-senescence and immunosuppressant drugs aggravate PTI. PTI has been intensively studied, but published results are frequently cloudy. The purpose of this review is to focus on the contributions made by different responsive modalities to immunosuppression following sterile trauma and to try to integrate these into an overall scheme of PTI.

#### STERILE POST-TRAUMATIC IMMUNOSUPPRESSION

An evolutionarily sophisticated and balanced immune system exists in our body whose equilibrium can be altered by different physical, environmental, or psychological stresses. Trauma, including major surgery and accidental injury, leads to post-traumatic immunosuppression (PTI) that increases a patient's vulnerability to hospital-acquired infections. However, the underlying mechanisms of PTI are poorly defined and as yet, there are no universally accepted treatments.

In 1856 Florence Nightingale drew attention to the 'utter insignificance' of risk of from battle wounds dving acquired during the *Crimean War* compared to the risk of dying from subsequent zymotic (infectious) diseases acquired within the Scutari Hospital.<sup>2</sup> continuous Nightingale's improvements in hospital hygiene gradually reduced deaths from infectious diseases in patients with combat injuries. However more and more new infections are still threatening the battle wound patients.<sup>3, 4</sup> An important, but poorly highlighted, question remains namely "why do wounded systemic patients acquire infections even in a hygienic environment?" Research over the last two decades suggests that an "imbalanced immunity in patients following trauma" is the most important factor in increasing patient's а vulnerability to acquire infections. Therefore, although sanitation has been improved in the hospitals, an equivalent phenomenon of post-traumatic deaths from systemic infections persists to this day. Recent in

#### **DEFINITIONS.**

**Immune status** is the level of appropriately targeted resistance to internal, opportunistic and external pathogenic microorganisms, where resistance is attributable to multiple innate or acquired mechanisms located throughout the organism. There is no single measure of immune status, but rather multiple biom.arkers that make up a profile describing broad aspects of innate and adaptive immunity.

**Post-Traumatic Immunosuppression (PTI)**, for the purpose of this review, is the condition of suppressed immune status that follows sterile trauma. It ranges from mild to severe, where mild is exemplified by strenuous exercise and severe by immunosuppression induced by multiple extensive trauma or major open surgery.

**Sterile trauma** refers to tissue damage devoid of primary wound infection, and is best exemplified by elective open surgery, for example in knee joint arthroplasty. Inflammation following sterile trauma without any exposure to microbial pathogens is termed *'sterile inflammation'*.<sup>1</sup> Immunosuppression followed by this initial sterile inflammation termed as *'sterile immunosuppression'*.

**Non-sterile trauma** refers to tissue damage due to sepsis or any injury/surgery in the presence of microbial infection. Inflammation following any trauma with exposure to microbial pathogens is termed 'non-sterile inflammation'. Immunosuppression followed by initial non-sterile inflammation can be termed as non-sterile immunosuppression.

#### Inflammation has different definitions.

*Clinical Concept* – John Hunter's outstanding surgical and experimental observations on "inflammation" suggested four clinical signs namely redness, heat, swelling, and pain.<sup>5</sup> However, Hunter's definition of "Inflammation" was not based on understanding the immunological responses.

*Immunological Concept* – It is only relatively recently that inflammation has become an important term in immunology. The Pro-/Anti-inflammatory paradigm is the basic concept of inflammation reflected in a balance between two opposing cytokine networks that activate or suppress immunity.

*vivo* studies confirmed the trauma-associated translocation of endogenous bacteria from the gut following sterile head injury,<sup>6,7</sup> indicating another source of PTI.

One attempt to conceptualise the phenomenon of PTI was the so-called SIRS-CARS paradigm, where trauma is described as a 'Systemic Inflammatory Response Syndrome', followed by a 'Compensatory Anti-inflammatory Response Syndrome'. In 1991 Dr Roger Bone introduced the term 'Systemic Inflammatory Response Syndrome (SIRS)" to describe physiological changes common to all cases of burn, trauma or sepsis. This paradigm included four physiological changes namely, increased body temperature, elevated heart rate, tachypnea or hyperventilation, and leukocytosis or lekocytopenia.<sup>8</sup> Despite its obvious limitations, until now SIRS has been the dominant paradigm adopted by clinicians worldwide. For example, *body* temperature can increase following sepsis and sterile trauma, but also after myocardial infarction, pulmonary embolism and strenuous exercise. Elevated heart rate and tachypnea can occur following sepsis and sterile trauma, but other physiological complications such as cardiac and respiratory failures, hypovolemic shock and erythropenia can also affect these parameters. On the other hand *alteration* in white blood cell count can also happen in different disease conditions such as heart failure, pancreatitis and burns. In 1996, Bone modified his SIRS paradigm by adding a sequel named "Compensatory Anti-inflammatory Response Syndrome (CARS)". CARS was characterized by a decrease in antigen presentation, macrophage paralysis, decrease in T-cell proliferation, increase in T-cell and dendritic cell apoptosis and shift in the T-cell subsets from Th1 to Th2 phenotype.<sup>9</sup> However in 2002, these parameters were further modified with many additional criteria included, such as increases in C-Reactive protein (CRP), creatinine, bilirubin or lactate, or hyperglycemia in the absence of diabetes,<sup>10</sup> but in general their addition has resulted in more confusion in the field. Injured tissues release soluble factors that act on the endocrine, lymphoid and haematopoietic organs. Moreover, the SIRS-CARS criteria as initially proposed were poorly associated with immuno-functional parameters and recent observations failed to fit with this original paradigm.<sup>11-16</sup> Therefore, the SIRS-CARS concept may not be the best way of describing PTI.

Micro- versus macro-surgery (minor- versus major- injury) also plays an important role in post-traumatic immunology and a patient's clinical outcome. Minimally invasive surgeries reduce complications compared with conventional operations. Considering the advantages of minimally invasive surgical procedures such as the requirement of smaller incisions, less analgesics, less injury scores and reduced stay in hospital, this form of surgery has become common practice. The severity of trauma is reflected in the amount of cytokine production<sup>17, 18</sup> with little cytokine release following minimally invasive surgeries, such as laparoscopic Following major surgeries, procedures. such as arthroplasty or other colorectal/vascular procedures, higher levels of cytokines are released.

Investigations after major trauma, where venous blood samples are monitored, assume that the results obtained reflect systemic immunity, although in reality, much

of the immune response is confined to locally injured tissues. Following the initial local inflammatory events that tend to increase tissue damage, there is activation of an anti-inflammatory cascade that helps local wound healing. This anti-inflammatory cascade is therefore a very important physiological event to maintain immune-homeostasis following sterile trauma. Rock *et al*<sup>19</sup> emphasize current limitations in our knowledge about the immunological consequences of sterile inflammation particularly the role of dead cells and their released sterile irritant particles in IL-1 dependent inflammasome activation. Thus their review provided a concise account of the effects of different released particles from dying and necrotic cells on inflammation. Although sterile PTI is a complex event, little information is available on changes in the levels of different immune- and non-immune- soluble mediators and cell types in this process.

Both sterile- and non-sterile trauma can trigger partly similar physiological responses. This review is therefore purposely restricted to discuss only the immunological events following sterile trauma.

# IMMUNOLOGICAL CHANGES FOLLOWING STERILE TRAUMA

# 1. Changes at Local Site

#### - Skin, the primary Barrier

Human skin acts as both a physical and by local production of antimicrobial peptides (AMP) a chemical barrier against foreign microbial infections. Keratinocytes and resident cells of the innate immune system reduce the epidermal microbial load by triggering the release of AMPs.<sup>20</sup> Injury to the skin results in immediate haemostasis, followed by infiltration of neutrophils and macrophages and simultaneous production of cytokines such as IL-6 and IL-8.<sup>21</sup> IL-6 plays a role in fibroblast proliferation, collagen deposition and angiogenesis. Interestingly, a strong correlation exists between local IL-6 concentrations and the speed of wound healing.<sup>22</sup>

#### - Ischemia/Reperfusion related changes

Major orthopaedic surgery is frequently carried out under tourniquet, creating tissue hypoxia that, following tourniquet release, gives rise to ischemia-reperfusion injury to tissues. A key component of a cell's response to hypoxia is up-regulation of hypoxia-induced-transcription-factor (HIF) expression<sup>23</sup> In hypoxic conditions, increasing HIF and Toll like receptor (TLR) signalling synergistically activate the NFkB pathway thereby triggering recruitment of monocytes, phagocytosis and release of IL-18, TNF- $\alpha$  and DAMPs.<sup>24</sup> HIF also inhibits apoptosis of recruited neutrophils at the site of injury thereby prolonging their lifespan.<sup>25</sup> However, HIF up-regulation in T-cells stimulates IL-10 production, inducing a shift from inflammatory Th1 phenotype to anti-inflammatory Th2 phenotype.<sup>25</sup> Hypoxia increases adenosine levels that regulate innate immunity and controls inflammation.<sup>26</sup> Following tourniquet

release, reperfusion injury may also enhance the expression of inflammatory cytokines such as IL-17 facilitating further recruitment of leukocytes to the site of injury<sup>27</sup>; however DAMP driven immunological responses may reduce inflammatory events to facilitate healing.

# - Local Changes in Soluble Mediators

Reviewing the published literature, considerable variation has been reported in the protein levels of different soluble mediators at the surgical wound site, summarized in **Table-1** and detailed in Supplementary Table S1. There is general agreement that increased levels of IL-1 $\beta$ , IL-4, IL-6, IL-8, IL-10, TNF- $\alpha$ , PGE<sub>2</sub> and complement proteins- C3 and C5 were seen post-operatively.

Table-1: Lists of studies investigating the changes in biomarker levels at the surgical wound site.

Biomarker	Biomarker	No. of Investigations				
Туре	Name	Changes in wound site biomarker levels				Total No.
		relative to pre-operative baseline levels				of Studies
		Decrease	No Change	Increase	N.D.	
	IL-1β	-	2	10	1	13
	IL-2	-	1	-	1	2
Pro-	IL-6	-	-	24	-	24
inflammatory	IL-12	-	1	-	-	1
Cytokines	IL-17	-	1	-	-	1
	IFN-γ	-	1	-	-	1
	TNF-α	-	4	6	-	10
	IL-4	-	1	2	0	3
Anti-	IL-5	-	1	-	-	1
inflammatory	IL-10	1	1	3	-	5
Cytokines	IL-13	-	1	-	-	1
	IL-1RA	-	-	2	-	2
Chemokines	IL-8	-	-	12	-	12
	MCP-1	-	-	1	-	1
	MIP-1a	-	1	-	-	1
Others	PGE2	-	-	2	-	2
	sIL-6R	1	-	-	-	1
	C3	-	-	7	-	7
	C5	-	-	4	-	4
	Platelet	4	-	4	-	8
Here: N.D. = Not detectable						

### 2. Changes in hypothalamic-pituitary-Adrenal axis

The response to sterile trauma involves activation of the Hypothalamus-Pituitary-Adrenal (HPA) axis and subsequent changes in hormonal levels. There are complex functional inter-relationships between HPE and the immune system. Secretion of inflammatory cytokines such as IL-1, IL-6 and TNF- $\alpha$  has impacts on the HPA-axis.<sup>28</sup> Following trauma, release of these mediators triggers the secretion of corticotrophin releasing factor (CRF), a hypothalamic releasing substance by the anterior pituitary gland, that immediately stimulates the release of adrenocorticotropic hormone (ACTH).<sup>13</sup> ACTH rapidly stimulates production of the glucocorticoid cortisol. The production of ACTH following trauma overrides cortisol-dependent ACTH downregulation, ultimately resulting in continuous increases in both ACTH and cortisol.<sup>13</sup> Cortisol triggers the release of anti-inflammatory mediators such as IL-6, IL-1RA and soluble TNF-receptors that play an important role in controlling inflammatory events, thereby leading to systemic immunosuppression. Post-traumatic stress disorder (PTSD), leading to PTI, alters the homeostatic balance between the HPA axis and immune system due to imbalanced secretions of both non-immune- and immunemolecules that are involved in these physiological mechanisms.<sup>29</sup> Although the HPA axis and immune system are hugely inter-connected, little is known about the balance between these two axes in the clinical settings of PTSD induced PTI. Thus, mild traumatic brain injury (mTBI), another form of sterile trauma, may also lead to immunosuppression.<sup>30</sup>

### 3. Changes in Immune Cells

Increases in the number of neutrophils, monocytes, and MSCs in blood reflect recruitment of these cells from extra-vascular compartments (spleen and bone marrow) after sterile injury.<sup>31-33</sup> In contrast, depending on the degree of injury, reduction in the levels of red blood cells (RBC) and lymphocytes are seen.<sup>34</sup> The effects of trauma on individual immune cell types are briefly discussed below:

### - Neutrophils

During sterile trauma, neutrophils egress from the extra vascular spaces into the circulation and are recruited to the site of injury. This neutrophil egress is facilitated by the CXC chemokine receptors CXCR-4 and CXCR-2.<sup>35</sup> CXCR-4 ligands are involved in retaining neutrophils in the bone marrow, but CXCR-2 ligands inhibit CXCR-4 dependant retention thereby encouraging egress.<sup>31, 35</sup> Granulocyte colony-stimulating factor (G-CSF) also helps in the mobilization of neutrophils by altering the balance between ligation of CXCR4 and CXCR2.<sup>35</sup> Ubiquitin, an endogenous CXCR4 agonist, is also released following trauma and burns, controlling the extent of neutrophil release from the bone marrow to the site of injury.<sup>36, 37</sup>

Neutrophils in the peripheral blood can also be rapidly recruited to the site of injury.<sup>38, 39</sup> By their interaction with different damage-associated molecular patterns

(DAMPs), these sentinel cells release different inflammatory cytokines and chemoattractants that recruit additional neutrophils to the inflammatory site.<sup>39, 40</sup> Following sterile trauma, a huge number of immature neutrophils mobilize from bone marrow into the circulation. However, recent investigation on immune response by the immature neutrophils (by checking the ability of phagocytosis and chemotaxis and measuring the expressions of CD14, CD16, Toll-like receptor (TLR)-2 and TLR4) in patients with sterile SIRS, showed that, although immature, these cells can still perform their crucial task as a first line of defence.<sup>41</sup> Studies also described the limitations of these immature neutrophils in the circulation following surgery.<sup>42, 43</sup>

### - Monocytes and Dendritic cells

The mechanisms involved in the egress of monocytes from the bone marrow to the injury site are still poorly understood. In mouse, Tsou *et al* <sup>32</sup> illustrated the recruitment of Ly-6<sup>+</sup> monocytes from the bone marrow into the circulation by a CCR-2 mediated pathway. Additional investigation suggested the spleen as another reservoir of monocyte recruited to damaged tissues.<sup>44</sup> Importantly, emigration of monocytes from the bone marrow to the injured site is independent of neutrophil recruitment.<sup>45</sup>

Decreased expression of HLA-DR by circulating monocytes was also found in patients following trauma, major surgery and burns.<sup>34, 46</sup> Antigen presenting cells (APC), such as dendritic cells (DC), interact with lymphocytes to trigger adaptive immunity. Kawasaki *et al*<sup>47</sup> reported decreased antigen presentation capacity by splenic DC in mice following trauma/haemorrhage. This was associated with decreased expression of MHC class II, IL-12 and IFN-γ. The total number of DCs showed a transient increase after surgery, but decreased at postoperative days 2-3<sup>48</sup>. Recent studies showed decreased myeloid DC (MDC) but no change in plasmacytoid DC (PDC) 3 to 5 days after surgery.<sup>49</sup> MDCs may possibly have been recruited to the surgery site. Circulating MDCs but not PDCs were shown by Maier *et al*<sup>50</sup> to undergo apoptosis. Several studies on DCs from circulating blood showed increased expressions of genes for chemotaxis including CCL5, CXCL5 and CXCL4, antiapoptosis (such as TIMP-1, BCL2), and inflammation (such as NF-κB).<sup>50-53</sup> These results may indicate a role for DC in the recruitment of innate immune cells at the injury site.

Myeloid-derived suppressor cells (MDSC) are a mixed population of myeloid cells capable of suppressing T cell responses. The number of CD14<sup>+</sup> APC cells was found to be significantly elevated for at least 24 hours following surgery with relatively greater increases in CD16<sup>+</sup>, CD80<sup>+</sup> and CD86<sup>+</sup> APCs.<sup>54</sup> More importantly, there was a massive increase in the number of CD14<sup>+</sup>HLADR<sup>-</sup> myeloid derived suppressor cells (MDSCs),<sup>55</sup> indicating their involvement in immunosuppression following trauma.<sup>56</sup> However, further phenotypic and functional studies are needed to confirm the subtype of these CD14<sup>+</sup> APCs and their role in trauma induced immunological responses.

### - Mesenchymal Stromal Cell

Recent investigation show increased numbers of circulatory mesenchymal stromal cells (MSC) in the peripheral blood of burns and trauma patients.<sup>33, 57</sup> There appears to be a direct correlation between the proliferation rate of cultured bone marrow MSCs from patients with multiple traumas and trauma severity.<sup>33</sup> How MSC egress from bone marrow and are recruited to the injury site remains unclear.

# - Lymphocytes

A significant decrease in total CD3+ T-lymphocytes following trauma is associated with a shift from Th1 to Th2 phenotype mediated by regulatory T-cells.<sup>46,</sup> <sup>58</sup> Albertsmeier et al<sup>54</sup> showed a decrease in CD3+, CD4+ and CD28+ T-cell counts immediately after surgery with no changes in CD8+ T cells; CD4+CD25+CD127regulatory T-cells (Treg cells) involved in shifting the T-helper compartment from Th1 to Th2 phenotype were increased. They hypothesized that, suppression of T-cells following trauma is associated with increased Treg cells and MDSCs.<sup>54</sup> In vitro stimulation studies also showed reduced secretion of IFN- $\gamma$ , IL-2 and TNF- $\alpha$  by postoperative T-cells.<sup>46</sup> On the other hand, Munoz et al<sup>59</sup> showed no postoperative changes in CD19+ B lymphocyte counts from the first postoperative day until seventh. Lymphopenia and also the deactivation of cells in stroke patients are indicative of immune-suppression following sterile trauma.<sup>60</sup>

# - Natural Killer Cells

The functional activity of natural killer (NK) cells as a first line responder of innate immunity is decreased following traumatic injury.<sup>61, 62</sup> The number of CD16+CD56+ NK cells is decreased one day after knee joint replacement surgery. This decrease persisted until one week after surgery.<sup>59</sup> Gharehbaghian et al<sup>61</sup> also showed significant suppression of NKp frequencies five days following joint replacement surgery; however the exact CD phenotype of such NKp was not determined.

#### - Other Cells

Postoperative eosinophil counts initially greatly decline and then return to baseline after two days.<sup>63</sup> Decreases may be associated with the increased ACTH secretions through signalling by the HPE-axis.<sup>13, 63</sup> Basophils also decrease one day after surgery and return to normal at day five and there is significant release in histamines.<sup>64</sup> However, phenotypic changes to eosinophils and basophils following major surgeries are poorly documented.

# 4. Changes in Transcript Level

Major trauma triggers the up-regulation of different genes in local neutrophils at the local site. Genes involved include IL-1 receptor antagonist (IL-1RA), IL-18 receptor-1 (IL-18R1), macrophage inflammatory-protein- $3\alpha$ ,

macrophage migration-inhibitory factor (MIF), group-II phospholipase-A2, whereas there is down-regulation of IL-8 receptor-β.<sup>65, 66</sup> Laudanski et al<sup>67</sup> also reported severe trauma induced significant changes in the expression of 2800 genes in monocytes and more than 5500 genes in T-lymphocytes, with an overall increased expression in more than 3000 genes in total WBCs. Circulating monocytes also showed up-regulation in Fas-Ligand, a member of TNF-superfamily.<sup>68</sup> However, since mRNA expression alone cannot provide complete information on the levels and activities of the active molecules in the local site of trauma, mRNA levels need to be confirmed by analysis of the corresponding proteins.

# 5. Changes in plasma biomarkers after sterile trauma

Complex changes in the levels of different cytokines occur after trauma. The most common changes are in IL-6. Miller *et al*<sup>69</sup> showed a post-traumatic decrease of IL-2. IFN- $\gamma$ , and IL-12 levels, whereas IL-10 and IL-4 levels were increased. This was associated with a shift from Th1 to Th2 T-cell responses. A comprehensive list of 109 investigations of changes in cytokine levels at different postoperative times is summarized in Table-2. Increased chemokines play important roles in the recruitment of mononuclear phagocytic cells from the bone marrow and spleen reservoirs to the site of injury.<sup>70</sup> IL-8/CXCL-8 rises transiently following surgery, but drops back to baseline within few days. DAMPs, also called endogenous danger factors or alarmins, are a class of molecule that play vital roles in the recruitment of immune cells to the site of injury. DAMPs are released at the wound site during the trauma period and examples include HMGB-1, S100 proteins, heat shock proteins and  $\alpha$ -defensions.<sup>71</sup> Routine arrays of DAMP concentrations in venous blood would help to better characterize post-traumatic immunity. Following blood loss, haemodilution, inflammation and other reasons, the levels of haemoglobin and haematocrit drop following major surgery.<sup>72</sup> Complement activation produces complement components such as C5a and C3a that increase post-operatively.<sup>73</sup> Release of histones and nucleosomes from damaged cells allows factor VII-activating protease (FSAP) to trigger the complement pathway and generation of C5a.<sup>74</sup> Soluble CD-14, a marker of monocyte activation increases following surgery;<sup>75</sup> whereas decreases are reported in soluble IL-6 receptor (sIL-6R)<sup>76</sup> and soluble gp-130.<sup>77</sup> In knee arthroplasty, Munoz et al<sup>59</sup> showed decreased levels of different immunoglobulins (IgG, IgA, and IgM) at six hours and these remained persistently low for at least three days.

Individuals do not respond equally to surgical trauma/stress due to variations in concentrations of soluble factors. This can be the result of genetic polymorphisms in the promoters of cytokine genes<sup>78, 79</sup> as well as other factors such as race, age, and obesity.<sup>80, 81</sup> For this review, a comprehensive summary of published literature describing post-operative changes in the concentrations of different cytokines, chemokines, and DAMP is presented in Table-2 and detailed in Supplementary Table

S2. At the most frequently used time-point, namely 24 hour, changes are summarized as follows:

- 87 out of 88 studies showed a consistent post-operative increase in IL-6; in one study IL-6 was undetectable.
- No increases were shown in 17% studies of IL-8, 30% studies of IL-10 and 27% studies of IL-1RA. But in the remainder studies all were increased.
- For IL-1 $\beta$ , IL-4, IL-12, IL-13, IL-17, IFN- $\gamma$ , TNF- $\alpha$ , and MCP-1, most studies showed no post-operative changes.
- Decreased IL-2 was shown in 54% (7/13) studies and unchanged in 4 studies.
- IL-5 and MIP-1 $\alpha$  were unchanged in 50% of studies; although there were only 4 studies for IL-5 and 2 for MIP-1 $\alpha$ .
- There was only one study of IL-22 and soluble IL-6 Receptor (sIL-6R) each. IL-22 was increased whereas sIL-6R levels were down.
- For sCD14, all three studies reported postoperative increases.
- HMGB-1 showed elevations in five studies; whereas one study showed no change.
- HSP-27 was studied and showed unchanged by one group. HSP-70, reported by two groups, also showed no postoperative changes. In the only study of HSP-60, levels were decreased.

Table-2: Lists of investigations on changes in biomarker levels in postoperative
venous blood following major surgery

Biomarker Type	Biomarker Name	No. of Investigations				
		Postoperative Changes (1 day) Also see Supplementary Table S2			Total No. of Studies	
		Decrease	No Change	Increase	N.D.	
	IL-1β	3	21	3	5	32
	IL-2	7	4	1	1	13
	IL-6	-	-	87	1	88
Pro-inflammatory	IL-12	2	5	1	-	8
Cytokines	IL-17	-	1	-	1	2
	IFN-γ	1	6	0	-	7
	TNF-α	5	23	6	5	39
	IL-22	-	-	1	-	1
	IL-4	1	4	2	-	7
Anti-inflammatory	IL-5	-	2	2	-	4
Cytokines	IL-10	-	12	28	-	40
	IL-13	-	4	-	-	4
	IL-1RA	-	3	8	-	11
	IL-8	-	7	31	2	40
Chemokines	MCP-1	-	7	2	-	9
	MIP-1a	1	1	-	-	2
	HMGB-1	-	1	5	-	6
DAMPs	HSP-27	-	1	-	-	1
	HSP-60	1	-	-	-	1
	HSP-70	-	2	-	-	2
Others	sIL-6R	1	-	-	-	1
	sCD-14	-	-	3	-	3

Following trauma, necrotic cells release neo-antigens (by different enzymes and post-translational modifications) that trigger autoantibody Immunoglobulin-M (IgM) production by B1a B-cells. A recent study showed that expression of Sialic acid-binding immunoglobulin-type lectin-10 (Siglec-10) by B1a B-cells reduces their expansion and thus prevents the production of IgM.<sup>82</sup> Thus Siglec-10 may play a vital role in alleviating autoimmune reactions. Both systemic and local elevations of soluble CD24 and Siglec-10 were recorded following sterile trauma in our study.<sup>83, 84</sup> This indicates a possible evolutionary advantage of PTI that PTI may suppresses auto-immunity to neo-antigens released from necrotic tissues at the sites of injury.<sup>85</sup>

# 6. Effect of Trauma on Remote Organs

#### - Liver

The liver releases acute phase proteins (APP) after trauma. IL-6 stimulates hepatocytes to release APPs such as C Reactive Protein (CRP), Serum Amyloid-A (SAA), Activated Protein-C, and alpha-1-antitrypsin.<sup>86-88</sup> Elevated APPs stimulate production of cytokine antagonists such as IL-1RA and soluble TNF-receptors, thereby eventually resulting in immune-suppression and favouring wound healing.<sup>89, 90</sup> Although APPs were initially thought to have only pro-inflammatory activities, recent studies suggest that their role following trauma is predominantly anti-inflammatory.<sup>90, 91</sup>

# Gut

Sterile trauma also increases intestinal inter-cellular permeability of the patients. Maintenance of the gut epithelial barrier largely depends on the microbiota. Based on the degree of increases in intestinal permeability, sterile trauma may induce dysbiosis of the microbiome. This may trigger the translocation of bacteria from the gut to the circulation, and increase the risk of systemic PTI. However, whether PTI and increase gut permeability are directly associated with each other, remains to be investigated.<sup>6, 7, 92</sup>

**Figure-1** schematically represents an overview of these complex interactions and attempts to summarize the literature review undertaken. Based on current understanding, Table-3 attempts to compare the key features of sterile trauma in the inflammation phase versus the immunosuppressive phase.

	Immunological Events following Sterile Trauma				
	Activation Phase		Suppression Phase		
	Local	Systemic	Local	Systemic	
Macrophages	Inflammatory M1	-	Anti-inflammatory M2	Anti-inflammatory M2	
	polarization		polarization	polarization	
Monocyte	Inflammatory	Monocytosis	Differentiation into Anti-	Increase in the number of anti-	
•	phenotype		inflammatory phenotype	inflammatory phenotypes.	

 Table 3: Different Immunological phases in local site of trauma and systemic

 circulation following sterile trauma

Neutrophil	Migration into site of	Neutrophilia	Apoptosis of neutrophils at	t Overall increase in number of		
- · · · · · · · · · · · · · · · · · · ·	injury	-	site of injury	neutrophils however probably		
				immature or less active		
Lymphocyte	Inflammatory Th1	Increase in	Anti-inflammatory Th2	Anti-inflammatory Th2		
	pathway	lymphocyte	pathway	pathway		
		apoptosis				
NK cells	-	-	Decrease in NK cell	Decrease in NK cell function		
			function			
DAMPs	Elevated levels of		Elevated levels of DAMPs	DAMPs direct anti-		
	DAMPs trigger		direct anti-inflammatory	inflammatory activities to		
	immediate release of	-	activities to favour wound	favour systemic PTI		
	cytokines to facilitate		healing			
	inflammation					
HPA axis	Immediate HPA-axis		Cortisol and Prostaglandin	Cortisol and Prostaglandin		
	mediated signalling to	-	mediated suppression to	also play role in systemic		
	initiate injury induced		favour healing	suppression		
	responses					
Acute Phase	Immediate release as	Decreased	Trigger local	Trigger immunosuppressive		
Proteins	part of body's	levels of	immunosuppressive	responses		
	inflammatory activities	positive APPs	responses			
MDSCs		Release of	Possible recruitment of	Sustained release of MDSCs		
	-	MDSCs from	MDSCs to the site of injury	probably direct towards		
		the source	to facilitate local healing	systemic suppression		
Cytokines	Immediate release of different		Continued release of anti-	Reductions in inflammatory		
	inflammatory and anti-inflammatory		inflammatory cytokines	cytokines such as IL-1 $\beta$ , IL-2,		
	cytokines		whereas down-regulation of	TNF- $\alpha$ , IFN- $\gamma$ and IL-17A		
			pro-inflammatory cytokine	whereas elevations in anti-		
			release	inflammatory IL-5.		

# **CLINICAL CONSEQUENCES OF PTI**

Recent studies indicate that PTI following surgical trauma enhances patients' vulnerability to acquire exogenous infections from the hospital or endogenous infections associated with microbial translocations from the gut.<sup>6, 7, 60, 93-96</sup> Different forms of nosocomial infections may occur in the immuno-compromised patient in different body compartments.<sup>97</sup> Respiratory tract infections are the most frequent, followed by urinary tract, wound and bloodstream infections.<sup>98</sup> Furthermore, patients may acquire infections with multi-drug resistant bacteria that are resistant to most antibiotics.<sup>99</sup> In extreme situations, PTI may eventually lead to severe sepsis, multiple organ failure and death. A rather alarming view is that the world is heading towards a post-antibiotic era due to the resistance of bacteria such as E. coli, Klebsiella, tuberculosis and gonorrhoea to all known antibiotics. This phenomenon has been termed the "Antibiotic Apocalypse" that may lead to life-threatening dangers even from a routine infection.

Hospital-acquired pneumonia (HAP) is the most common amongst all the nosocomial infections. Pneumonia although mostly caused by Gram-negative bacteria, can also manifest as infections by other bacteria and viruses. Multicentre studies on medical-surgical patients in intensive care units and on patients who acquired infections in ICU revealed the highest episodes of pneumonia; most of which were ventilator associated.<sup>98, 101</sup> Therefore, following surgery and the associated immune-compromised state, patients' vulnerability to acquired pneumonia increases.

The gastrointestinal (GI) tract is another important compartment in our body where infections are often reported following injuries such as blunt trauma and invasive abdominal surgeries. Infections in GI tract also can be manifested with injuries to the spleen, pancreas, and duodenum.<sup>102, 103</sup>

Urinary tract infection (UTI) mostly affects the bladder; however it can also affect the kidney, ureters and urethra. UTI is another common infection in hospital caused by bacteria that enter the urethra and then the bladder. UTI is the second leading cause in ICU patients and it is mostly associated with catheterization.<sup>98</sup> Major surgery most often requires the use of urinary catheters; therefore increasing the chance of UTI.

Bloodstream infections are also common nosocomial infections in hospital particularly from *Staphylococcus Aureus* and coagulase-negative *staphylococci*. This infection can spread into the blood stream by several ways; especially via the intravenous line required in trauma patients.<sup>104</sup>

# **AGGRAVATING FACTORS OF PTI**

PTI can be aggravated by many different biological factors. Blood transfusion, frequently a necessary step following surgery, also aggravates PTI. Transfusion of allogeneic blood or packed red blood cells aggravates PTI, as does transfusion of predeposited autologous blood.<sup>61, 105-107</sup> Haemorrhage is another important aggravating factor which results in immunosuppression even without any major tissue trauma and may increase susceptibility to acquired infections.<sup>108</sup> By releasing pro-inflammatory cytokines IFN- $\gamma$  and IL-2 in vitro with enhanced production of IL-10 by T-cells, haemorrhage may lead to PTI.<sup>109</sup> Surgery itself is a major trigger of physical stress.<sup>110</sup> In addition, factors such as pre-surgical medications and anaesthesia show a direct association with the degree of surgical stresses.<sup>111</sup> Other possible aggravating factors include mTBI, PTSD and stroke.<sup>29, 30, 60</sup> Use of anti-inflammatory/immunosuppressant drugs such as corticosteroids, cyclosporin-A, and anti-TNFa treatments also depress patients' immunity and therefore may also increases susceptibility to acquire infections.<sup>112-114</sup> Another biological factor aggravating PTI is immune-senescence. Elderly patients have weak immunity and are therefore more vulnerable to infections.<sup>115, 116</sup> In animal studies Zacks *et al*<sup>117</sup> previously showed relatively slower muscle regeneration and decreased phagocytic activities in older animals.

# **TREATMENT OF PTI**

Considering all the above events following sterile trauma and the resulting immunosuppression, there has been increasing interest over the last decade in the use of immuno-stimulants to prevent PTI before, during, or after surgical trauma. Commonly used immuno-stimulants can be either functional nutrients or immunotherapeutic agents. Pre-existing malnutrition has a great impact on clinical outcome, and proper nutritional support helps to reverse PTI. Immuno-nutrients prescribed pre-, intra-, and/or post-operatively can help to prevent post-surgical immunosuppression and include: glutamine, arginine, n-acetyl cysteine, branched-chain amino acids, glucan, nucleotides, long-chain n-3 fatty acids, antioxidant vitamins, trace elements, and taurine.<sup>118-122</sup> Patients treated with probiotics may also get substantial benefit following trauma.<sup>123</sup>

Strategies to modulate patients' immunity for better clinical outcome are mostly confined to drugs to prevent overwhelming inflammatory reactions following so-called SIRS. However, without knowing the underlying mechanisms of sepsis or sterile trauma, treatment with drugs may aggravate the clinical situation. This is reflected in the failure of many different clinical trials.<sup>124, 125</sup> Immunotherapeutic agents used include: anti-PD1, mifamurtide, polyinosinic-polycytidylic acid, MF-59, imiquimod, luivac, myrrh, IDR-peptides, IL-7, rIFN- $\gamma$  and IL-15.<sup>125-128</sup>

Although allogeneic and pre-deposited autologous blood transfusion further suppressed PTI,<sup>61</sup> thereby increasing the chance of nosocomial infections, we described the immuno-stimulatory activities of postoperatively salvaged autologous blood transfusion after major surgery.<sup>61, 83</sup> This intriguing observation suggests that yet-to-be-identified natural bio-factors produced at the wound site may exist and if identified and administered intravenously, may have great therapeutic potentials to reverse PTI.

### DISTINGUISHING STERILE TRAUMA FROM SEPSIS

One of the key unanswered questions at the present time in the field of inflammation research is how to distinguish PTI induced by sterile trauma from that induced by sepsis. The SIRS-CARS paradigm, although widely practiced by clinicians, does not distinguish these two conditions. In a recent seminal publication, Savage *et al*<sup>129</sup> analysed the secretory capacity of glial cells stimulated with different Damage Associate Molecular Patterns (DAMPs) to mimic the sterile trauma condition) or the Pathogen Associated Molecular Pattern (PAMP) molecule (LPS) to mimic sepsis. Differences were seen in the secretions of IL-1 $\beta$  namely: elevations when stimulated by PAMPs but unchanged when stimulated by DAMPs. However, secretion of IL-6 and CXCL-1 were identical in both situations. This study thus highlighted the differences in cellular responses following sterile versus non-sterile activation and open new avenues to investigate the consequences of inflammasome activation on these processes.

Johnson *et al*<sup>78</sup> also showed significant variations in the gene expression profile of whole blood cells taken from patients with sterile trauma *versus* early sepsis. Results obtained highlighted numerous genes that were preferentially increased in early sepsis. These unique genes could be subdivided into four broad categories namely: innate immunity, cytokine receptors, T helper cell differentiation, and protein synthesis.<sup>78</sup>

Liu *et al*<sup>130</sup> distinguished certain differences between sterile trauma and sepsis. Thus, following sterile trauma, secreted DAMPs form tri-molecular complex with membrane bound CD24 molecules and the trans-membrane glycoprotein Siglec-10 that inhibits TLR/NLR-mediated inflammation. In contrast, in the case of microbial infections, the sialic acid chain of CD24 molecule is cleaved by the pathogen encoded sialidase enzyme; therefore the tri-molecular complex cannot form, resulting in the TLR/NLR mediated inflammation.<sup>130-132</sup> These *in vivo* mouse models and *in vitro* human studies need to be further validated in a clinical setting.

Both sterile- and infectious- trauma results in similar types of inflammation patterns; although a higher degree of inflammatory signals are recorded following sepsis compared to sterile trauma.<sup>78</sup> This indicates that, following trauma the human body responds by directing the injured organ to be healed by different mechanisms, whereas during sepsis these mechanisms may be altered due to continuous endotoxin (PAMP) induced inflammation.

# CONCLUSION

Sterile trauma involves both local and systemic responses representing a complex balance between pro- and anti-inflammatory mediators. Briefly, at the trauma site, there is an immediate release of endogenous danger signals named damage associated molecular patterns (DAMPs) that increases local production of different cytokines, chemokines, and other soluble factors by resident cells. Subsequently, via DAMPs and chemokines, there is further recruitment of neutrophils, monocytes and mesenchymal stromal cells (MSC) into the local site of inflammation.<sup>39, 133-135</sup> These newly recruited cells differentiate locally into cells secreting predominantly antiinflammatory cytokines.<sup>134, 136</sup> Additionally, neutrophils and monocytes at the injured site are guided by DAMP induced inflammasomes to play a role in apoptosis/necroptosis/NETosis and controlled clearing of damaged tissues.<sup>137</sup> DAMPs also trigger the release of IL-6 that plays an important role in the HPA axis to release steroids and in turn trigger the release of other immunosuppressive mediators to control inflammation at the wound site. IL-6 also induces hepatocytes to release acute phase proteins that mediate post-traumatic anti-inflammatory activities. All the cells and soluble factors at the local site contribute to tissue remodelling and wound healing.<sup>1, 19</sup> These complex local and systemic changes following sterile trauma therefore lead to changes in patients' immune status that would vary between individuals.

Although many gaps in our understanding prevail, an outline is emerging of multiple mechanisms that restrain inflammation and suppress systemic immunity.<sup>138</sup> Suppression of immunity in moments of crisis would appear to have conferred no evolutionary advantage to mammals; but without such restraints, healing would be delayed by activation of tissue destructive inflammatory cascades.

Sterile trauma leads to changes in post-traumatic immunity and therefore a patient may be immune-compromised. Trauma accompanied by infection may also alter the immune status. Moreover, patients with these clinical conditions can be harmed, or put into unnecessary risk, by inappropriate treatment. Figure-2 describes the immune status following trauma and how inappropriate immunomodulatory drugs can result in aggravating a patient's outcome. Figure-2 also describes the benefit of knowing the immune status of patients to treat them with an appropriate immunomodulatory drug. Detailed analysis of immune status of patients therefore needs to be carried out in order to tailor the appropriate therapy. Measuring a patient's immune status individually prior to treatment would appear to be the best clinical approach to achieve the desired clinical outcome.

# ACKNOWLEDGEMENT

We sincerely thank Professor Luke O'Neill, Dept of Biochemistry and Immunology, Trinity College Dublin, Ireland and Professor Manuel Munoz, University of Malaga, Spain for their valuable suggestions and input to this review. This work was supported by Science Foundation Ireland under Grant Numbers SFI09/SRC/B1794 (to RC) and by a Science Foundation Ireland Stokes Professorship SFI07/SK/B1233b (to RC). NI obtained financial support from the Irish Research Council - EMBARK Award (RS/2011/223). The authors acknowledge no conflict of interest.

# **FIGURE LEGENDS:**

Figure 1: Schematic Representation of PTI after Sterile Trauma. This figure schematically illustrates the sequence of events following major trauma. In brief, at the local injury site, release of different DAMPs by the damaged tissues induces viable cells to secrete chemokines such as IL-8, MCP-1, and MIP-1 $\alpha$  and the immediate secretion of inflammatory cytokines such as TNF-a. DAMPs release results in IL-6 and TNF- $\alpha$  production that activate the HPE-Axis to release ACTH. Cortisol, and also PGE2. These events then trigger the secretion of anti-inflammatory biomolecules such as IL-1RA, IL-10, and sTNF-R. Later, IL-6, by virtue of its ability to trigger the release of acute phase proteins (APPs) by the liver, indirectly involved in reducing inflammatory events at the injured site. In parallel with the immediate release of DAMPs, inflammatory cytokines and the activities of resident immune cells, there is recruitment of neutrophils, monocytes, and MSCs from the blood to the injured site. Bone marrow and spleen act as reservoirs for the egression of these cells to the site of injury via- the blood. By releasing AMPs and helping in haemostasis, damaged skin also activates inflammasomes to release DAMPs at the local site. On the other hand, ischaemia-reperfusion injury associated with the surgical procedure increases HIF and ROS expression that also trigger the production of DAMPs and thus play an additional role in recruiting immune cells to the injured site. The ultimate goal of the above events following a major trauma is tissue remodelling and promotion of wound healing.

**Figure 2: Measuring Immune Status helps in choosing appropriate Immunomodulatory drug.** Measuring a patient's immune status based on particular profile of specific pro- and anti-inflammatory proteins will help to indicate if the patient: i) has a stable immune status and medication is not necessary, ii) is at the stage of cytokine storm following trauma or infection when both pro- and antiinflammatory cytokines rise, iii) requires immediate attention with treatment (either immunostimulatory or immunosuppressive drug will be administered), or iv) is not in need of medication but requires monitoring over the coming days. Knowing immune status will provide information following trauma or sepsis that can be monitored throughout the patient's recovery, thereby preventing the risk of unnecessary danger from incorrect therapy.

# **REFERENCES:**

1. Chen GY, Nunez G. Sterile inflammation: sensing and reacting to damage. *Nature Reviews Immunology* 2010; **10**: 826-837.

2. Nightingale F. Florence Nightingale: Measuring Hospital Care Outcomes: Excerpts From the Books Notes on Matters Affecting the Health, Efficiency, and Hospital Administration of the British Army Founded Chiefly on the Experience of the Late War, and Notes on Hospitals. Oakbrook Terrace, Ill: Joint Commission on Accreditation of Healthcare Organizations. Oakbrook Terrace, Ill, 1999.

3. Warkentien T, Rodriguez C, Lloyd B, Wells J, Weintrob A, Dunne JR, et al. Invasive Mold Infections Following Combat-related Injuries. *Clin Infect Dis* 2012; **55**: 1441-1449.

4. Murray CK. Infectious disease complications of combat-related injuries. *Crit Care Med* 2008; **36**: 358-364.

5. Hunter J. A treatise on the blood, inflammation, and gun-shot wounds, by the late John Hunter. To which is prefixed, A short account of the author's life, by his brother-in-law, Everard Home. London: printed by John Richardson, For George Nicol, 1794.

6. Oztuna V, Ersoz G, Ayan I, Eskandari MM, Colak M, Polat A. Early internal fracture fixation prevents bacterial translocation. *Clin Orthop Relat Res* 2006; **446**: 253-258.

7. Oztuna V, Ersoz G, Ayan I, Eskandari MM, Uguz K, Kuyurtar F. Head injury-associated bone fractures induce bacterial translocation: an experimental study. *J Orthop Trauma* 2004; **18**: 92-95.

8. Bone RC, Balk RA, Cerra FB, Dellinger RP, Fein AM, Knaus WA, et al. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. The ACCP/SCCM Consensus Conference Committee. American College of Chest Physicians/Society of Critical Care Medicine. *Chest* 1992; **101**: 1644-1655.

9. Bone RC. Immunologic dissonance: a continuing evolution in our understanding of the systemic inflammatory response syndrome (SIRS) and the multiple organ dysfunction syndrome (MODS). *Ann Intern Med* 1996; **125**: 680-687.

10. Levy MM, Fink MP, Marshall JC, Abraham E, Angus D, Cook D, et al. 2001 SCCM/ESICM/ACCP/ATS/SIS International Sepsis Definitions Conference. *Crit Care Med* 2003; **31**: 1250-1256.

11. Costa A, Benedetto V, Ricci C, Merlin P, Borelli P, Fadda E, et al. Endocrine, hematological and immunological changes in surgical patients undergoing general anesthesia. *Ital J Surg Sci* 1989; **19**: 41-49.

12. Philippou A, Maridaki M, Theos A, Koutsilieris M. Cytokines in muscle damage. *Adv Clin Chem* 2012; **58**: 49-87.

13. Burton D, Nicholson G, Hall G. Endocrine and metabolic response to surgery. *Continuing Education in Anaesthesia, Critical Care and Pain* 2004; **4**: 144-147.

14. Prockop DJ, Oh JY. Mesenchymal stem/stromal cells (MSCs): role as guardians of inflammation. *Mol Ther* 2012; **20**: 14-20.

15. Buvanendran A, Kroin JS, Berger RA, Hallab NJ, Saha C, Negrescu C, et al. Upregulation of prostaglandin E2 and interleukins in the central nervous system and peripheral tissue during and after surgery in humans. *Anesthesiology* 2006; **104**: 403-410.

16. Lu J, Goh SJ, Tng PY, Deng YY, Ling EA, Moochhala S. Systemic inflammatory response following acute traumatic brain injury. *Front Biosci (Landmark Ed)* 2009; **14**: 3795-3813.

17. Cruickshank AM, Fraser WD, Burns HJ, Van Damme J, Shenkin A. Response of serum interleukin-6 in patients undergoing elective surgery of varying severity. *Clinical Science (London)* 1990; **79**: 161-165.

18. Neidhardt R, Keel M, Steckholzer U, Safret A, Ungethuem U, Trentz O, et al. Relationship of Interleukin-10 Plasma Levels to Severity of Injury and Clinical Outcome in Injured Patients. *Journal of Trauma and Acute Care Surgery* 1997; **42**: 863-871.

19. Rock KL, Latz E, Ontiveros F, Kono H. The sterile inflammatory response. *Annu Rev Immunol* 2010; **28**: 321-342.

20. Sorensen OE, Thapa DR, Roupe KM, Valore EV, Sjobring U, Roberts AA, et al. Injuryinduced innate immune response in human skin mediated by transactivation of the epidermal growth factor receptor. *J Clin Invest* 2006; **116**: 1878-1885.

21. Sjogren F, Anderson C. Sterile trauma to normal human dermis invariably induces IL1beta, IL6 and IL8 in an innate response to "danger". *Acta Derm Venereol* 2009; **89**: 459-465.

22. Lin ZQ, Kondo T, Ishida Y, Takayasu T, Mukaida N. Essential involvement of IL-6 in the skin wound-healing process as evidenced by delayed wound healing in IL-6-deficient mice. *J Leukoc Biol* 2003; **73**: 713-721.

23. Kaelin WG, Jr., Ratcliffe PJ. Oxygen sensing by metazoans: the central role of the HIF hydroxylase pathway. *Mol Cell* 2008; **30**: 393-402.

24. Kuhlicke J, Frick JS, Morote-Garcia JC, Rosenberger P, Eltzschig HK. Hypoxia inducible factor (HIF)-1 coordinates induction of Toll-like receptors TLR2 and TLR6 during hypoxia. *PLoS One* 2007; **2**: e1364.

25. Eltzschig HK, Carmeliet P. Hypoxia and inflammation. *N Engl J Med* 2011; **364**: 656-665.

26. Ohta A, Sitkovsky M. Role of G-protein-coupled adenosine receptors in downregulation of inflammation and protection from tissue damage. *Nature* 2001; **414**: 916-920.

27. van Golen RF, Reiniers MJ, Olthof PB, van Gulik TM, Heger M. Sterile inflammation in hepatic ischemia/reperfusion injury: present concepts and potential therapeutics. *J Gastroenterol Hepatol* 2013; **28**: 394-400.

28. Besedovsky HO, del Rey A, Klusman I, Furukawa H, Monge Arditi G, Kabiersch A. Cytokines as modulators of the hypothalamus-pituitary-adrenal axis. *J Steroid Biochem Mol Biol* 1991; **40**: 613-618.

29. Griffin GD, Charron D, Al-Daccak R. Post-traumatic stress disorder: revisiting adrenergics, glucocorticoids, immune system effects and homeostasis. *Clinical & Translational Immunology* 2014; **3**: e27.

30. Griffin GD. Stroke, mTBI, infection, antibiotics and beta blockade: Connecting the dots. *Med Hypotheses* 2015; **85**: 224-229.

31. McDonald B, Kubes P. Cellular and molecular choreography of neutrophil recruitment to sites of sterile inflammation. *J Mol Med* 2011; **89**: 1079-1088.

32. Tsou CL, Peters W, Si Y, Slaymaker S, Aslanian AM, Weisberg SP, et al. Critical roles for CCR2 and MCP-3 in monocyte mobilization from bone marrow and recruitment to inflammatory sites. *J Clin Invest* 2007; **117**: 902-909.

33. Seebach C, Henrich D, Tewksbury R, Wilhelm K, Marzi I. Number and proliferative capacity of human mesenchymal stem cells are modulated positively in multiple trauma patients and negatively in atrophic nonunions. *Calcif Tissue Int* 2007; **80**: 294-300.

34. Kimura F, Shimizu H, Yoshidome H, Ohtsuka M, Miyazaki M. Immunosuppression following surgical and traumatic injury. *Surg Today* 2010; **40**: 793-808.

35. Suratt BT, Petty JM, Young SK, Malcolm KC, Lieber JG, Nick JA, et al. Role of the CXCR4/SDF-1 chemokine axis in circulating neutrophil homeostasis. *Blood* 2004; **104**: 565-571.

36. Majetschak M, Zedler S, Hostmann A, Sorell LT, Patel MB, Novar LT, et al. Systemic ubiquitin release after blunt trauma and burns: association with injury severity, posttraumatic complications, and survival. *J Trauma* 2008; **64**: 586-596.

37. Majetschak M. Extracellular ubiquitin: immune modulator and endogenous opponent of damage-associated molecular pattern molecules. *J Leukoc Biol* 2011; **89**: 205-219.

38. Arancibia SA, Beltran CJ, Aguirre IM, Silva P, Peralta AL, Malinarich F, et al. Toll-like receptors are key participants in innate immune responses. *Biol Res* 2007; **40**: 97-112.

39. Kolaczkowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nature Reviews Immunology* 2013; **13**: 159-175.

40. Williams MR, Azcutia V, Newton G, Alcaide P, Luscinskas FW. Emerging mechanisms of neutrophil recruitment across endothelium. *Trends Immunol* 2011; **32**: 461-469.

41. Drifte G, Dunn-Siegrist I, Tissieres P, Pugin J. Innate immune functions of immature neutrophils in patients with sepsis and severe systemic inflammatory response syndrome. *Crit Care Med* 2013; **41**: 820-832.

42. El-Maallem H, Fletcher J. Effects of surgery on neutrophil granulocyte function. *Infect Immun* 1981; **32**: 38-41.

43. Köller M, Wick M, Muhr G. Decreased Leukotriene Release from Neutrophils after Severe Trauma: Role of Immature Cells. *Inflammation* 2001; **25**: 53-59.

44. Swirski FK, Nahrendorf M, Etzrodt M, Wildgruber M, Cortez-Retamozo V, Panizzi P, et al. Identification of Splenic Reservoir Monocytes and Their Deployment to Inflammatory Sites. *Science* 2009; **325**: 612-616.

45. Henderson RB, Hobbs JA, Mathies M, Hogg N. Rapid recruitment of inflammatory monocytes is independent of neutrophil migration. *Blood* 2003; **102**: 328-335.

46. Hensler T, Hecker H, Heeg K, Heidecke CD, Bartels H, Barthlen W, et al. Distinct mechanisms of immunosuppression as a consequence of major surgery. *Infect Immun* 1997; **65**: 2283-2291.

47. Kawasaki T, Fujimi S, Lederer JA, Hubbard WJ, Choudhry MA, Schwacha MG, et al. Trauma-hemorrhage induces depressed splenic dendritic cell functions in mice. *J Immunol* 2006; **177**: 4514-4520.

48. Ho CS, Lopez JA, Vuckovic S, Pyke CM, Hockey RL, Hart DN. Surgical and physical stress increases circulating blood dendritic cell counts independently of monocyte counts. *Blood* 2001; **98**: 140-145.

49. Henrich D, Maier M, Relja B, Trendafilov P, Schiessling S, Wirth M, et al. Significant Decline of Peripheral Myeloid Dendritic Cells Following Multiple Trauma. *J Surg Res* 2009; **154**: 239-245.

50. Maier M, Geiger EV, Henrich D, Ebrahimi R, Wutzler S, Lehnert M, et al. Apoptosis differs in dendritic cell subsets early after severe trauma. *Hum Immunol* 2009; **70**: 803-808.

51. Geiger EV, Maier M, Schiessling S, Wutzler S, Lehnert M, Marzi I, et al. Subsequent gene expression pattern in dendritic cells following multiple trauma. *Langenbeck's Archives of Surgery* 2013; **398**: 327-333.

52. Maier M, Geiger EV, Henrich D, Bendt C, Wutzler S, Lehnert M, et al. Platelet factor 4 is highly upregulated in dendritic cells after severe trauma. *Mol Med* 2009; **15**: 384-391.

53. Maier M, Wutzler S, Bauer M, Trendafilov P, Henrich D, Marzi I. Altered Gene Expression Patterns in Dendritic Cells After Severe Trauma: Implications for Systemic Inflammation and Organ Injury. *Shock* 2008; **30**: 344-351.

54. Albertsmeier M, Quaiser D, von Dossow-Hanfstingl V, Winter H, Faist E, Angele MK. Major surgical trauma differentially affects T-cells and APC. *Innate Immun* 2015; **21**: 55-64.

55. Filipazzi P, Valenti R, Huber V, Pilla L, Canese P, Iero M, et al. Identification of a new subset of myeloid suppressor cells in peripheral blood of melanoma patients with modulation by a granulocyte-macrophage colony-stimulation factor-based antitumor vaccine. *J Clin Oncol* 2007; **25**: 2546-2553.

56. Cuenca AG, Delano MJ, Kelly-Scumpia KM, Moreno C, Scumpia PO, Laface DM, et al. A paradoxical role for myeloid-derived suppressor cells in sepsis and trauma. *Mol Med* 2011; **17**: 281-292.

57. Mansilla E, Marin GH, Drago H, Sturla F, Salas E, Gardiner C, et al. Bloodstream cells phenotypically identical to human mesenchymal bone marrow stem cells circulate in large

amounts under the influence of acute large skin damage: new evidence for their use in regenerative medicine. *Transplant Proc* 2006; **38**: 967-969.

58. Marik PE, Flemmer M. The immune response to surgery and trauma: Implications for treatment. *Journal of Trauma and Acute Care Surgery* 2012; **73**: 801-808.

59. Munoz M, Cobos A, Campos A, Ariza D, Munoz E, Gomez A. Post-operative unwashed shed blood transfusion does not modify the cellular immune response to surgery for total knee replacement. *Acta Anaesthesiol Scand* 2006; **50**: 443-450.

60. Klehmet J, Harms H, Richter M, Prass K, Volk HD, Dirnagl U, et al. Stroke-induced immunodepression and post-stroke infections: lessons from the preventive antibacterial therapy in stroke trial. *Neuroscience* 2009; **158**: 1184-1193.

61. Gharehbaghian A, Haque KM, Truman C, Evans R, Morse R, Newman J, et al. Effect of autologous salvaged blood on postoperative natural killer cell precursor frequency. *Lancet* 2004; **363**: 1025-1030.

62. Blazar BA, Rodrick ML, O'Mahony JB, Wood JJ, Bessey PQ, Wilmore DW, et al. Suppression of natural killer-cell function in humans following thermal and traumatic injury. *J Clin Immunol* 1986; **6**: 26-36.

63. Roche M, Thorn GW, Hills AG. The Levels of Circulating Eosinophils and Their Response to ACTH in Surgery. *N Engl J Med* 1950; **242**: 307-314.

64. Laroche D, Chrysanthou S, Lefrançois C, Gérard JL, Dubois F, Malet M, et al. Evidence of a progressive degranulation of basophil in subjects operated on without adverse reaction. *Agents Actions* 1992; **36**: C199-C200.

65. Buvanendran A, Mitchell K, Kroin JS, Iadarola MJ. Cytokine gene expression after total hip arthroplasty: surgical site versus circulating neutrophil response. *Anesth Analg* 2009; **109**: 959-964.

66. Abe T, Sakamoto K, Mita S, Kamohara H, Hirano Y-i, Kuwahara N, et al. Surgical Trauma Induces Group II Phospholipase A2 Production by Neutrophils at a Local Site after Surgery. *Clin Biochem* 1998; **31**: 101-106.

67. Laudanski K, Miller-Graziano C, Xiao W, Mindrinos MN, Richards DR, De A, et al. Cellspecific expression and pathway analyses reveal alterations in trauma-related human T cell and monocyte pathways. *Proceedings of the National Academy of Sciences* 2006; **103**: 15564-15569.

68. Sugimoto M, Shimaoka M, Hosotsubo K, Tanigami H, Taenaka N, Kiyono H, et al. Upregulation of Fas ligand (FasL) mRNA expression in peripheral blood mononuclear cells (PBMC) after major surgery. *Clin Exp Immunol* 1998; **112**: 120-125.

69. Miller AC, Rashid RM, Elamin EM. The "T" in trauma: the helper T-cell response and the role of immunomodulation in trauma and burn patients. *J Trauma* 2007; **63**: 1407-1417.

70. Gale LM, McColl SR. Chemokines: extracellular messengers for all occasions? *Bioessays* 1999; **21**: 17-28.

71. Manson J, Thiemermann C, Brohi K. Trauma alarmins as activators of damageinduced inflammation. *Br J Surg* 2012; **99 Suppl 1**: 12-20.

72. Khurana A, Zafar S, Abdul W, Mukhopadhyay S, Mohanty K. Postoperative Reduction in Haemoglobin levels is Related to Rise in Troponin T following Hip Fracture Surgery. *Journal of Bone and Joint Surgery, British Volume* 2012; **94-B**: 024.

73. Burk AM, Martin M, Flierl MA, Rittirsch D, Helm M, Lampl L, et al. Early complementopathy after multiple injuries in humans. *Shock* 2012; **37**: 348-354.

74. Kanse SM, Gallenmueller A, Zeerleder S, Stephan F, Rannou O, Denk S, et al. Factor VII-activating protease is activated in multiple trauma patients and generates anaphylatoxin C5a. *J Immunol* 2012; **188**: 2858-2865.

75. Bastian D, Tamburstuen MV, Lyngstadaas SP, Reikeras O. LBP and sCD14 patterns in total hip replacement surgery performed during combined spinal/epidural anaesthesia. *Scand J Clin Lab Invest* 2011; **71**: 486-491.

76. Kristiansson M, Soop M, Sundqvist KG, Soop A, Suontaka AM, Blomback M. Local vs. systemic immune and haemostatic response to hip arthroplasty. *Eur J Anaesthesiol* 1998; **15**: 260-270.

77. Corbi P, Rahmati M, Delwail A, Potreau D, Menu P, Wijdenes J, et al. Circulating soluble gp130, soluble IL-6R, and IL-6 in patients undergoing cardiac surgery, with or without extracorporeal circulation. *European Journal Cardio-Thoracic Surgery* 2000; **18**: 98-103.

78. Johnson SB, Lissauer M, Bochicchio GV, Moore R, Cross AS, Scalea TM. Gene expression profiles differentiate between sterile SIRS and early sepsis. *Ann Surg* 2007; **245**: 611-621.

79. Stuber F, Klaschik S, Lehmann LE, Schewe JC, Weber S, Book M. Cytokine promoter polymorphisms in severe sepsis. *Clin Infect Dis* 2005; **41 Suppl 7**: S416-420.

80. Silber JH, Rosenbaum PR, Kelz RR, Reinke CE, Neuman MD, Ross RN, et al. Medical and financial risks associated with surgery in the elderly obese. *Ann Surg* 2012; **256**: 79-86.

81. Haider AH, Chang DC, Efron DT, Haut ER, Crandall M, Cornwell EE, 3rd. Race and insurance status as risk factors for trauma mortality. *Arch Surg* 2008; **143**: 945-949.

82. Jellusova J, Wellmann U, Amann K, Winkler TH, Nitschke L. CD22 x Siglec-G doubledeficient mice have massively increased B1 cell numbers and develop systemic autoimmunity. *J Immunol* 2010; **184**: 3618-3627.

83. Islam N, Whitehouse M, Mehendale S, Hall M, Tierney J, O'Connell E, et al. Posttraumatic Immunosuppression is Reversed by Anti-coagulated Salvaged Blood Transfusion; Deductions from studying Immune Status after Knee Arthroplasty. *Clin Exp Immunol* 2014; **177**: 509-520.

84. Islam N. Characterization of post-traumatic immunosuppression and its partial reversal by autologous salvaged blood transfusion. *PhD Thesis, Aran Library, NUI Galway* [28 *January 2015]. 2015. Available at:* <u>http://hdl.handle.net/10379/4868</u>.

85. Darrah E, Andrade F. NETs: the missing link between cell death and systemic autoimmune diseases? *Front Immunol* 2012; **3**: 428.

86. Jawa RS, Anillo S, Huntoon K, Baumann H, Kulaylat M. Analytic Review: Interleukin-6 in Surgery, Trauma, and Critical Care: Part I: Basic Science. *J Intensive Care Med* 2011; **26**: 3-12.

87. Heinrich PC, Castell JV, Andus T. Interleukin-6 and the acute phase response. *Biochem J* 1990; **265**: 621-636.

88. Gabay C, Kushner I. Acute-phase Proteins. eLS: John Wiley & Sons, Ltd; 2001.

89. Gabay C, Kushner I. Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med* 1999; **340**: 448-454.

90. Tilg H, Dinarello CA, Mier JW. IL-6 and APPs: anti-inflammatory and immunosuppressive mediators. *Immunol Today* 1997; **18**: 428-432.

91. Arredouani MS, Kasran A, Vanoirbeek JA, Berger FG, Baumann H, Ceuppens JL. Haptoglobin dampens endotoxin-induced inflammatory effects both in vitro and in vivo. *Immunology* 2005; **114**: 263-271.

92. Pape HC, Dwenger A, Regel G, Auf<sup>'</sup>m<sup>'</sup>Kolck M, Gollub F, Wisner D, et al. Increased gut permeability after multiple trauma. *Br J Surg* 1994; **81**: 850-852.

93. Shorr AF, Jackson WL. Transfusion practice and nosocomial infection: assessing the evidence. *Curr Opin Crit Care* 2005; **11**: 468-472.

94. Faruquzzaman. Positive associations of nosocomial infections in surgical ward with etiological clinical factors. *Bratisl Lek Listy* 2011; **112**: 273-277.

95. Meisel C, Schwab JM, Prass K, Meisel A, Dirnagl U. Central nervous system injuryinduced immune deficiency syndrome. *Nat Rev Neurosci* 2005; **6**: 775-786.

96. Chamorro A, Urra X, Planas AM. Infection after acute ischemic stroke: a manifestation of brain-induced immunodepression. *Stroke* 2007; **38**: 1097-1103.

97. Rabinowitz RP, Caplan ES. Management of infections in the trauma patient. *Surg Clin North Am* 1999; **79**: 1373-1383.

98. Richards MJ, Edwards JR, Culver DH, Gaynes RP. Nosocomial infections in combined medical-surgical intensive care units in the United States. *Infect Control Hosp Epidemiol* 2000; **21**: 510-515.

99. Alanis AJ. Resistance to antibiotics: are we in the post-antibiotic era? *Arch Med Res* 2005; **36**: 697-705.

100. British Broadcasting Corporation. Antibiotic resistance: World on cusp of 'postantibiotic era'. *BBC News Health*; 2015 [19 November 2015]. Available from: <u>http://www.bbc.com/news/health-34857015</u>.

101. Ponce de Leon-Rosales SP, Molinar-Ramos F, Dominguez-Cherit G, Rangel-Frausto MS, Vazquez-Ramos VG. Prevalence of infections in intensive care units in Mexico: a multicenter study. *Crit Care Med* 2000; **28**: 1316-1321.

102. GOINS WA, RODRIGUEZ A, JOSHI M, JACOBS D. Intra-abdominal Abscess After Blunt Abdominal Trauma. *Ann Surg* 1990; **212**: 60-65.

103. Ivatury RR, Zubowski R, Psarras P, Nallathambi M, Rohman M, Stahl WM. Intraabdominal abscess after penetrating abdominal trauma. *J Trauma* 1988; **28**: 1238-1243.

104. Civetta JM, Hudson-Civetta J, Ball S. Decreasing catheter-related infection and hospital costs by continuous quality improvement. *Crit Care Med* 1996; **24**: 1660-1665.

105. Gong MN, Thompson BT, Williams P, Pothier L, Boyce PD, Christiani DC. Clinical predictors of and mortality in acute respiratory distress syndrome: potential role of red cell transfusion. *Crit Care Med* 2005; **33**: 1191-1198.

106. Silverboard H, Aisiku I, Martin GS, Adams M, Rozycki G, Moss M. The role of acute blood transfusion in the development of acute respiratory distress syndrome in patients with severe trauma. *J Trauma* 2005; **59**: 717-723.

107. Karam O, Tucci M, Toledano BJ, Robitaille N, Cousineau J, Thibault L, et al. Length of storage and in vitro immunomodulation induced by prestorage leukoreduced red blood cells. *Transfusion (Paris)* 2009; **49**: 2326-2334.

108. Stephan RN, Kupper TS, Geha AS, Baue AE, Chaudry IH. HEmorrhage without tissue trauma produces immunosuppression and enhances susceptibility to sepsis. *Arch Surg* 1987; **122**: 62-68.

109. Abraham E, Chang YH. Haemorrhage-induced alterations in function and cytokine production of T cells and T cell subpopulations. *Clin Exp Immunol* 1992; **90**: 497-502.

110. Hogan BV, Peter MB, Shenoy HG, Horgan K, Hughes TA. Surgery induced immunosuppression. *Surgeon* 2011; **9**: 38-43.

111. Kawasaki T, Ogata M, Kawasaki C, Okamoto K, Sata T. Effects of epidural anaesthesia on surgical stress-induced immunosuppression during upper abdominal surgery. *Br J Anaesth* 2007; **98**: 196-203.

112. Ali T, Kaitha S, Mahmood S, Ftesi A, Stone J, Bronze MS. Clinical use of anti-TNF therapy and increased risk of infections. *Journal of Drug, Healthcare and Patient Safety* 2013; **5**: 79-99.

113. Gea-Banacloche JC, Opal SM, Jorgensen J, Carcillo JA, Sepkowitz KA, Cordonnier C. Sepsis associated with immunosuppressive medications: an evidence-based review. *Crit Care Med* 2004; **32**: S578-590.

114. Hansen BL, Rohr N, Svendsen V, Olsen H, Birkeland SA. Bacterial Urinary Tract Infection in Cyclosporine-A Immunosuppressed Renal Transplant Recipients. *Scand J Infect Dis* 1988; **20**: 425-427.

115. Aw D, Silva AB, Palmer DB. Immunosenescence: emerging challenges for an ageing population. *Immunology* 2007; **120**: 435-446.

116. Ginaldi L, Loreto MF, Corsi MP, Modesti M, De Martinis M. Immunosenescence and infectious diseases. *Microbes and Infection* 2001; **3**: 851-857.

117. Zacks SI, Sheff MF. Age-related impeded regeneration of mouse minced anterior tibial muscle. *Muscle Nerve* 1982; **5**: 152-161.

118. Kudsk KA. Immunonutrition in surgery and critical care. *Annu Rev Nutr* 2006; **26**: 463-479.

119. Helminen H, Raitanen M, Kellosalo J. Immunonutrition in elective gastrointestinal surgery patients. *Scand J Surg* 2007; **96**: 46-50.

120. Stechmiller JK, Childress B, Porter T. Arginine immunonutrition in critically ill patients: a clinical dilemma. *Am J Crit Care* 2004; **13**: 17-23.

121. Zheng Y, Li F, Qi B, Luo B, Sun H, Liu S, et al. Application of perioperative immunonutrition for gastrointestinal surgery: a meta-analysis of randomized controlled trials. *Asia Pac J Clin Nutr* 2007; **16 Suppl 1**: 253-257.

122. Calder PC. Immunonutrition in surgical and critically ill patients. *Br J Nutr* 2007; **98 Suppl 1**: S133-139.

123. Klaenhammer TR, Kleerebezem M, Kopp MV, Rescigno M. The impact of probiotics and prebiotics on the immune system. *Nature Reviews Immunology* 2012; **12**: 728-734.

124. Boomer JS, Green JM, Hotchkiss RS. The changing immune system in sepsis: Is individualized immuno-modulatory therapy the answer? *Virulence* 2014; **5**: 45-56.

125. Hotchkiss RS, Monneret G, Payen D. Immunosuppression in sepsis: a novel understanding of the disorder and a new therapeutic approach. *Lancet Infect Dis* 2013; **13**: 260-268.

126. Hancock RE, Nijnik A, Philpott DJ. Modulating immunity as a therapy for bacterial infections. *Nature Reviews Microbiology* 2012; **10**: 243-254.

127. Scanzello C, Figgie M, Nestor B, Goodman S. Perioperative Management of Medications Used in the Treatment of Rheumatoid Arthritis. *HSS J* 2006; **2**: 141-147.

128. Hutchins NA, Unsinger J, Hotchkiss RS, Ayala A. The new normal: immunomodulatory agents against sepsis immune suppression. *Trends Mol Med* 2014; **20**: 224-233.

129. Savage CD, Lopez-Castejon G, Denes A, Brough D. NLRP3-Inflammasome Activating DAMPs Stimulate an Inflammatory Response in Glia in the Absence of Priming Which Contributes to Brain Inflammation after Injury. *Front Immunol* 2012; **3**: 288.

130. Liu Y, Chen GY, Zheng P. CD24-Siglec G/10 discriminates danger- from pathogenassociated molecular patterns. *Trends Immunol* 2009; **30**: 557-561.

131. Chen GY, Chen X, King S, Cavassani KA, Cheng J, Zheng X, et al. Amelioration of sepsis by inhibiting sialidase-mediated disruption of the CD24-SiglecG interaction. *Nat Biotechnol* 2011; **29**: 428-435.

132. Chen GY, Tang J, Zheng P, Liu Y. CD24 and Siglec-10 selectively repress tissue damage-induced immune responses. *Science* 2009; **323**: 1722-1725.

133. Shi C, Pamer EG. Monocyte recruitment during infection and inflammation. *Nature Reviews Immunology* 2011; **11**: 762-774.

134. Han Z, Jing Y, Zhang S, Liu Y, Shi Y, Wei L. The role of immunosuppression of mesenchymal stem cells in tissue repair and tumor growth. *Cell and Bioscience* 2012; **2**: 8.

135. McDonald B, Pittman K, Menezes GB, Hirota SA, Slaba I, Waterhouse CC, et al. Intravascular danger signals guide neutrophils to sites of sterile inflammation. *Science* 2010; **330**: 362-366.

136. Arnold L, Henry A, Poron F, Baba-Amer Y, van Rooijen N, Plonquet A, et al. Inflammatory monocytes recruited after skeletal muscle injury switch into antiinflammatory macrophages to support myogenesis. *J Exp Med* 2007; **204**: 1057-1069.

137. Kaczmarek A, Vandenabeele P, Krysko DV. Necroptosis: the release of damageassociated molecular patterns and its physiological relevance. *Immunity* 2013; **38**: 209-223.

138. Medzhitov R. Origin and physiological roles of inflammation. *Nature* 2008; **454**: 428-435.