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<td>Doheny, HC, Houlihan, DD, Ravikumar, N, Smith, TJ, Morrison, JJ</td>
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Human Chorionic Gonadotrophin Relaxation of Human Pregnant Myometrium and Activation of the BKCa Channel

HELEN C. DOHENY, DIARMAID D. HOULIHAN, NANDINI RAVIKUMAR, TERRY J. SMITH, AND JOHN J. MORRISON

Department of Obstetrics and Gynaecology (H.C.D., D.D.H., N.R., J.J.M.), National University of Ireland Galway, Clinical Science Institute, University College Hospital Galway, Galway, Ireland; and National Centre for Biomedical Engineering Science (T.J.S.), National University of Ireland, Galway, Ireland.

The uterorelaxant effect of human chorionic gonadotrophin (hCG) is regarded as an important mediator in maintenance of uterine quiescence during pregnancy with clinical potential for tocolysis, the mechanisms of which are unknown. The large conductance calcium-activated K⁺ channel (BKCa) is ubiquitously encountered in human uterine tissue and plays a significant role in modulating myometrial cell membrane potential and excitability. The objective of this study was to investigate the involvement of BKCa channel function in the response of human myometrial cells to hCG. Single electrophysiological BKCa channel recordings from freshly dispersed myocytes were obtained in the presence and absence of increasing hCG concentrations. Isometric tension studies, investigating the effects of hCG on isolated myometrial contractions, in the presence and absence of the BKCa channel blocker, iberiotoxin, were performed. The hCG significantly increased the open-state probability of these channels in a concentration-dependent manner [control 0.036 ± 0.01; 1 IU/ml hCG 0.065 ± 0.014 (P = 0.262); 10 IU/ml hCG 0.111 ± 0.009 (P = 0.001); and 100 IU/ml hCG 0.098 ± 0.004 (P = 0.007)]. In vitro functional studies demonstrated that hCG exerted a significant concentration-dependent relaxant effect on human myometrial tissue. This effect was significantly attenuated by preincubation with iberiotoxin (P < 0.05). These findings outline that activation of BKCa channel activity may explain the potent uterorelaxant effect of hCG. (J Clin Endocrinol Metab 88: 4310–4315, 2003)

The factors regulating myometrial quiescence during human pregnancy, and the subsequent transformation to a state of maximal excitability during labor, are poorly understood (1–3). Knowledge pertaining to such factors is essential to both an understanding of the physiology of human parturition and the clinical management of human labor and disorders thereof. The major clinical problem associated with human labor is that of preterm labor, which is the leading cause of perinatal mortality and morbidity in the developed world (1, 4, 5).

Human chorionic gonadotropin (hCG) is a heterodimeric glycoprotein produced primarily in the placenta (6). Although the importance of hCG in maintenance of early pregnancy has been widely accepted, recent reports have highlighted a potential role of hCG in maintaining uterine quiescence in the third trimester. The hCG exerts a potent concentration dependent inhibitory effect on human myometrial contractions in vitro (7). The hCG receptors in human myometrium are down-regulated following the onset of labor, in both term and preterm deliveries (7, 8). Although possible explanations for the inhibitory effect of hCG in human myometrial smooth muscle have included a direct reduction of intracellular calcium availability (9, 10), down-regulation of myometrial gap junctions (11) and inhibition of uterotonic agents such as eicosanoids (8), little is known of the endogenous mechanisms of hCG regulation of uterine contractility during pregnancy.

The state of contractility of the myometrium is ultimately dependent on both the concentration of and sensitivity to intracellular calcium (2, 12). Cell membrane ion channels play a major role in regulation of smooth muscle activity by controlling cell membrane potential and in the regulation of intracellular calcium concentration. For myometrium, Ca²⁺, Na⁺, K⁺, and Cl⁻ channels all contribute to the regulation of the resting membrane potential and the action potential, and both directly and indirectly contribute to the regulation of intracellular free Ca²⁺ concentrations (12). The K⁺ channels are the largest category of ion channels in the cell and hence are of primary importance in the regulation of membrane potential and cell excitability in the myometrium. The large conductance calcium-activated potassium channel (BKCa or maxi-K channel) is the predominant K⁺ channel type encountered in nonpregnant and pregnant human myometrium (14, 15) and has been reported as playing a significant role in modulating myometrial function in both animal models and in humans (12, 16). Although it is established that BKCa channel function in smooth muscle is closely linked to membrane conductance and cellular excitability, little is known about the actual mechanisms or role of BKCa modulation by endogenous substances. This study was undertaken to investigate whether hCG exerted an effect on open-state probability of BKCa in single channel electrophysiological recordings of human uterine myocytes and, second, to evaluate whether the relaxant effect of hCG on myometrial

Abbreviations: BKCa, Calcium-activated K⁺ channel; hCG, human chorionic gonadotropin; IßTX, iberiotoxin; NPo, mean open probability; PSS, physiologic saline solution; TEA, tetraethylammonium.
contractions was altered by BK_{Ca} blockade in functional studies using iberiotoxin (IbTX).

**Materials and Methods**

*Tissue collection*

Biopsies of human myometrium were obtained from women undergoing elective cesarean section in the third trimester of pregnancy in the Department of Obstetrics and Gynecology, University College Hospital, Galway, Ireland. The biopsies were excised from the midline portion of the upper lip of the incision in the lower uterine segment. Women who had received exogenous progastalandins, oxytocin, or corticosteroids were excluded from the study. Recruitment was by written informed consent. Ethics committee approval for the study was obtained from the Research Ethics Committee, University College Hospital, Galway. For electrophysiological studies, tissue samples were immediately placed in sterile Ham’s F-12 medium (Sigma-Aldrich, Dublin, Ireland) supplemented with 100U/mL penicillin and 100 μg/mL streptomycin (Sigma-Aldrich). Tissue samples for isometric recordings were placed in fresh Krebs-Henseleit physiologic saline solution (PSS) of the following composition (mmol/liter): 140 KCl, 110 NaCl, 1.2 MgSO_4_, 1.2 CaCl_2_, 12 KH_2PO_4_, 25 NaHCO_3_, and 11 glucose (Sigma-Aldrich). Tissue was stored at 4°C and used within 6 h of collection.

*Tissue dispersion/primary human myometrial cells*

The preparation of single myometrial cells for electrophysiological recordings was performed using methodology previously described (17, 18). Freshly isolated myometrial cells were obtained by enzymatic digestion of finely minced myometrium with 2 mg/ml collagenase (type IA, 300–400 U/mg) (Sigma-Aldrich) in Hank’s buffered salt solution. The incubation with enzyme was performed at 37°C for 2 h followed by centrifugation (1000 revolutions per minute) in 50% Percoll (Sigma-Aldrich) for 10 min. The cell layer was removed, washed, and spun in physiological solution to remove excess red blood cells. The cell suspension was then triturated and filtered through an 80-μm nylon mesh filter. Single cells were placed in a recording chamber (Warner Instrument Corp., Hamden, CT) and electrophysiological experiments begun immediately. Morphologically, freshly dissociated uterine myocytes were characterized by a long, slender fusiform shape. All myocytes used for this work were relaxed and adhered to the bottom of the recording chamber with no additional substrate.

*Electrophysiological recordings of the BK_{Ca} channel*

Single-channel recordings using the cell-attached configuration of the patch clamp technique were performed. Several drops of cell suspension were placed in the recording chamber containing a solution of the following composition (mmol/liter): 140 KCl, 0.1 CaCl_2_, 0.1 MgCl_2_, 25 NaHCO_3_, and 30 glucose (pH 7.4, 22–25°C). Single potassium channel currents were measured in cell-attached patches by filling the patch pipette (2–5 MΩ) with Ringer solution composed of (mmol/liter): 140 NaCl, 5 KCl, 1.2 MgSO_4_, 0.1 CaCl_2_, and 10 HEPES, and making a gigaohm seal on a single myocyte. Voltage across the patch was controlled by setting the cellular membrane potential to 0 mV using a high-K+ extracellular solution. Average channel activity in patches was measured as mean open probability (NP_s). The effects of hCG (1, 10, and 100 IU/mL) on channel activity were measured by recording about 10 sec of continuous recording at +40 mV before and 15 min after drug treatment. Currents were filtered at 1 kHz and digitized at 10 kHz. In all experiments, voltage-clamp and voltage-pulse generation were controlled with an Axopatch 200-A patch clamp amplifier (Axon Instruments, Inc., Union City, CA), and data were acquired and analyzed with pClamp 8. For statistical purposes and to ensure accuracy, NP_s was calculated for single BK_{Ca} channels at a depolarized potential in which channel openings can be clearly differentiated from other channel species. BK_{Ca} channels are easily visualized at +40 mV because there is virtually no contamination with other channel species. In addition, at such depolarized potentials, there is no other ion channel expressed in these cells with an amplitude of 7–9 pA, making statistical analysis of BK_{Ca} channel activity more accurate (19). Moreover, recording single-channel activity at more depolarized potentials helps compensate the artificial depression of activity otherwise encountered from recording channel activity at room temperature (22–25°C) (19).
BKCa channel activity

Single-channel recordings from freshly dispersed myocytes, using the cell attached configuration of the patch clamp technique, revealed ion channel activity with an amplitude average of 7–10 pA at +40 mV identifying the BKCa channel (19). NS1619, a specific BKCa channel opener, elicited potent channel activation at applied concentration of 20 μmol/liter (Fig. 1). Furthermore, application of 10 mmol/liter tetraethylammonium (TEA), which exhibits selectivity for BKCa channels, reversed the NS1619 activation of this channel. These experiments identified the BKCa channel as the predominant K+ channel expressed in human myometrial myocytes, consistent with previous studies on these cells (13, 14, 16).

A representative recording, demonstrating minimum BKCa channel activity under control conditions (NPo = 0.042), is shown in Fig. 2. Cumulative additions of hCG resulted in a potent activation of BKCa channel activity in a concentration-dependent fashion, as illustrated. BKCa channel activity for this recording was increased following the addition of 1 (NPo = 0.088), 10 (NPo = 0.121), and 100 (NPo = 0.107) IU/ml hCG, respectively. The average NPo (Fig. 3B) before and after addition of hCG was as follows: control 0.036 ± 0.01 (n = 5); 1 IU/ml hCG 0.065 ± 0.014 (n = 5; P = 0.262); 10 IU/ml hCG 0.111 ± 0.009 (n = 5; P = 0.001); and 100 IU/ml hCG 0.098 ± 0.004 (n = 4; P = 0.007). The hCG stimulated the BKCa channel with a maximal activation effect observed at a concentration of 10 IU/ml, and the effect appeared to plateau at higher concentrations. Open-state channel activity was stimulated 2- and 3-fold by 1 and 10 IU/ml hCG, respectively; however, subsequent addition of 100 IU/ml hCG did not elicit significant further effect. In general, there was a 5- to 10-min latency period before observation of hCG-stimulated channel activity, and this effect appeared to be maximal within 15–20 min. In all patches this activity persisted until either seal integrity was lost, or the experiment was terminated.

In vitro contractility

The hCG exerted a cumulative inhibitory effect on contractility in isolated pregnant human myometrium (n = 6). Figure 4A demonstrates a representative recording of oxytocin (0.5 nmol/liter) elicited myometrial contractility, and in Fig. 4B the effects of cumulative additions of hCG (0.1, 0.5, 1, 5, and 10 IU/ml) to the tissue bath are shown. ANOVA

**FIG. 1.** Representative recordings from the same membrane patch (+40 mV) in the cell-attached configuration and after addition of 20 μmol/liter NS1619 and subsequent addition of 10 mmol/liter TEA. TEA reversed NS1619-stimulated channel activity.

**FIG. 2.** Human CG stimulates BKCa channel activity in human myometrial smooth muscle cells. Representative continuous recordings were recorded from the same cell attached patch (+40 mV) before (control) and 15 min after cumulative exposure to 1, 10, and 100 IU/ml hCG, respectively. Channel openings are upward deflections from the baseline (closed state) (dashed line).

**FIG. 3.** Human CG opens BKCa channel activity in human myometrial smooth muscle cells in a concentration-dependent manner. A, Activity plot of BKCa channel open probability in a single cell-attached patch before and 15 min after exposure to cumulative doses of hCG (1, 10, and 100 IU/ml). Vertical bars are channel activity during a 100-msec test pulse to +40 mV. Recording time under each condition was 10 sec, as indicated on the time axis. Breaks in the time axis represent drug incubation periods when channel activity was not recorded. Record of drug exposure is indicated by horizontal lines above the histogram. B, Each bar represents the average channel open probability obtained from cell attached patches (+40 mV) before and 15 min after 1, and subsequent additions of 10 and 100 IU/ml hCG, respectively. *, Significant increase in channel activity, compared with control (n = 5; P < 0.01).
revealed a significant effect on contractility of both addition of hCG (group 1 vs. group 2; \( P < 0.05 \)) and increasing hCG concentrations. Post hoc analysis showed a statistically significant relaxant effect of hCG at bath concentrations of 1 (\( n = 5; P < 0.01 \)), 5 (\( n = 6; P < 0.01 \)), and 10 (\( n = 6; P < 0.01 \)) IU/ml when compared with respective control values measured from strips exposed to oxytocin alone.

Incubation of strips with IbTX, in addition to oxytocin, did not significantly alter the integrals of contractile activity measured (group 1 vs. group 3; \( n = 5; P = 0.83 \)). However, addition of IbTX, before bath addition of hCG, significantly attenuated the relaxant effect exerted by hCG. In Fig. 4C a representative recording demonstrating the effects of preincubation with the BK\(_{Ca}\) channel antagonist IbTX, on the uterorelaxant effects of hCG, is shown. Comparison of the inhibitory effects of hCG on the oxytocin-induced contractility of myometrial strips, in the presence or absence of IbTX, revealed a significant difference across the groups (group 2 vs. group 4; \( n = 6; P < 0.05 \)). Post hoc analysis revealed that preincubation with IbTX (100 nmol/liter) significantly attenuated the uterorelaxant effect of hCG at all test concentrations, resulting in no significant difference when compared with control values [group 3 (\( n = 5 \)) vs. group 4 (\( n = 6 \)); \( P > 0.05 \)] at all hCG bath concentrations. In Fig. 5 the effects of cumulative additions of hCG (0.1, 0.5, 1, 5, and 10 IU/ml) on contractions of myometrial strips, in the absence and presence of IbTX, are shown in graphical form alongside the integrals observed in control strips.

**Discussion**

The concept that hCG plays a role in maintenance of human uterine quiescence throughout pregnancy, and particularly in the third trimester, has been recently highlighted (7, 8). Based on hCG receptor quantification data, using mRNA and protein studies (8), it has been suggested that this relaxant effect of hCG is down-regulated at the time of labor. The exact mechanism, or mechanisms, of the inhibitory effect of hCG in myometrial smooth muscle are unknown, but direct reduction in intracellular calcium availability (10, 22) and down-regulation of gap junctions have previously been suggested (11). The potential inhibition of unknown uterotonnic agents by hCG has also been speculated (8). Our data demonstrate a clear and potent activation of the BK\(_{Ca}\) channel, directly by hCG, in human myometrium during pregnancy, at both the single ion channel level and in functional studies. Activation of the BK\(_{Ca}\) channel results in an efflux of K\(^+\) ions from the intracellular compartment, drawing the cell membrane potential closer to the potassium equilibrium potential and thereby reducing cellular excitability and contractility. These results provide a clear explanation of mechanism of action of the uterorelaxant effect of hCG; however, inhibition via other pathways is also possible. Our findings also account for the direct reduction in intracellular calcium elicited by hCG in myometrial tissue (10) by inactivation of...
voltage-gated calcium channels because of hyperpolarization of the cell membrane via BK_{Ca} channel activation.

In our experiments, single-channel patch clamp studies clearly identified BK_{Ca} channels in the isolated myocytes and demonstrated that hCG significantly increases their open state probability. These studies are consistent with previous reports indicating that the BK_{Ca} channel is the predominant potassium channel in primary myocytes from human pregnant myometrium (13–15). Because of its large conductance, and high density of expression, this channel plays an important role in the regulation of membrane potential and cellular excitability of the myometrium and hence has been reported as playing a significant role in modulating myometrial function in both animal models and humans. Initial links between the myometrial BK_{Ca} channel and parturition were evident with the report outlining that it apparently loses its Ca^{2+} and voltage sensitivity in human myometrium with the onset of labor (13). In addition, NS1619, the specific BK_{Ca} channel activator, exerts a potent relaxant effect on pregnant human myometrium, in vitro experiments (16). Using both inside-out and outside-out configurations of the patch clamp technique, NS1619 appears to act directly on myometrial BK_{Ca} channels and stimulates channel activity by increasing their time spent in the open state (16). The findings from this study indicate that hCG exerts its utero-relaxant effect in a manner similar to that of NS1619.

The results from single BK_{Ca} channel recordings concur with the data obtained from isometric tension recordings with human myometrial strips. After preincubation with IbTX, the contractility measured following hCG exposure, was not significantly different to that of control strips (i.e. strips exposed to oxytocin alone or oxytocin and IbTX). This confirms the fact that BK_{Ca} channel function is intrinsic to myometrial contractions and also providing evidence that this channel plays a central role in hCG-mediated relaxation. In our study, after the equilibration period, the strips were exposed to oxytocin to achieve regular rhythmic contractions, which are reproducible over a period of hours and hence serve as a reliable control, unlike spontaneous contractions in vitro (23). In our experiments, there was no significant difference observed in integrals of contractility in strips exposed to oxytocin alone vs. those exposed to oxytocin and IbTX, before bath addition of hCG.

Although the physiological and clinical importance of endogenous factors regulating myometrial quiescence during pregnancy are obvious, the potential use of hCG, administered exogenously, as a therapeutic agent for preterm labor, has been recently debated (7, 24, 25). Whole hCG exhibited potent inhibition of PGF_{2alpha}–induced preterm delivery in mice, and the effect was dose dependent (25). There are no reliable data in relation to clinical studies, to our knowledge. Our findings support the hypothesis that clinical tocolysis using hCG remains a possibility, with a predominant mechanism of action via BK_{Ca} channel opening in myometrial cells.

We did not address the possibility that the effects outlined in our results may be altered by labor onset. This remains an exciting study for further evaluation of both hCG effector mechanisms, and BK_{Ca} channel activity, in myometrial tissue obtained during spontaneous labor, with similar clinical criteria as outlined for this study.

In conclusion, we report the first direct evidence that hCG exerts a potent uterorelaxant effect in human pregnancy via BK_{Ca} channel activation. These novel data raise further questions about a central role of hCG, and BK_{Ca} channel function, in association with uterine quiescence and parturition. These findings support the growing hypothesis that hCG may confer therapeutic benefit in preterm labor management.

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Address all correspondence and requests for reprints to: Professor John J. Morrison, Department of Obstetrics and Gynaecology, National University of Ireland Galway, Clinical Science Institute, University College Hospital Galway, Galway, Ireland. E-mail: john.morrison@nuigalway.ie.

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**References**


