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Restriction of Human Polyomavirus BK Virus DNA Replication in Murine Cells and Extracts

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Received 11 February 2009/Accepted 19 March 2009

BK virus (BKV) causes persistent and asymptomatic infections in most humans and is the etiologic agent of polyomavirus-associated nephropathy (PVAN) and other pathologies. Unfortunately, there are no animal models with which to study activation of BKV replication in the human kidney and the accompanying PVAN. Here we report studies of the restriction of BKV replication in murine cells and extracts and the cause(s) of this restriction. Upon infection of murine cells, BKV expressed large T antigen (TAg), but viral DNA replication and progeny were not detected. Transfection of murine cells with BKV TAg expression vectors also caused TAg expression without accompanying DNA replication. Analysis of the replication of DNAs containing chimeric BKV and murine polyomavirus origins revealed the importance of BKV core origin sequences and TAg for DNA replication. A sensitive assay was developed with purified BKV TAg that supported TAg-dependent BKV DNA replication with human but not with murine cell extracts. Addition of human replication proteins, DNA polymerase α-primase, replication protein A, or topoisomerase I to the murine extracts with BKV TAg did not rescue viral DNA replication. Notably, addition of murine extracts to human extracts inhibited BKV TAg-dependent DNA replication at a step prior to or during unwinding of the viral origin. These findings and differences in replication specificity between BKV TAg and the TAGs of simian virus 40 (SV40) and JC virus (JCV) and their respective origins implicate features of the BKV TAg and origin distinct from SV40 and JCV in restriction of BKV replication in murine cells.

Persistent and asymptomatic infections by the human polyomavirus BK virus (BKV) occur in most humans (44) and have been implicated in pulmonary, ophthalmologic, hepatic, autoimmune, neurological, and renal disease (76). Free and integrated BKV genomes also have been detected in human tumor cells and tissues (10, 18); however, their significance in human cancer has not been established (46). The most significant and frequently noted consequence of BKV infection is polyomavirus-associated nephropathy (PVAN) with resulting risk of allograft loss (35). Activation of latent BKV replication in kidney allografts leading to PVAN has been suggested to be caused by inhibition of gamma interferon (1, 6), inflammation or stress, and ischemia/reperfusion (27, 34).

Animal models of BKV-associated diseases would be a great help in dissecting the cause(s) of PVAN and other BKV-related pathologies. Some rodent cells have been reported to be semipermissive for BKV infection (17, 82, 101). BKV infection or expression of the viral early region can cause malignant transformation in cultured cells (53, 72, 101) and hepatocellular carcinomas and renal and other tumors in rodents (21, 38, 88). As in human cells, BKV DNAs are maintained in some rodent cells at low levels, perhaps as episomes (10, 62); however, the processes limiting viral replication have not been defined. Murine polyomavirus (mPyV) DNA replication in mouse kidneys following injury (2, 3) or renal transplant (31) has been suggested to provide a useful model for the study of PVAN. However, replication of BKV in human and murine kidney cells must be better understood to judge the relevance of such models.

Previous studies of simian virus 40 (SV40), mPyV, and JC virus (JCV) have provided numerous insights into the processes of viral DNA replication. The viral large T antigen (TAg) helps to initiate replication by binding to multiple G/A/G)GGC motifs within the core origin, forming a dodecameric structure that distorts duplex DNA, opening DNA on one side and unwinding the other side (81, 95, 107). Single-stranded DNA is coated with replication protein A (RPA), and topoisomerase I relieves torsional stress ahead of the replication fork (7, 37, 86). DNA polymerase α-primase (Pol α-primase) synthesizes short RNA primers that are elongated by DNA polymerase α (28, 65), and leading-strand synthesis is completed by DNA polymerase δ, RPA, proliferating cell nuclear antigen (PCNA), and replication factor C (45, 57, 110). Replication of the lagging strand is mediated by Pol α-primase, DNA polymerase δ, and accessory proteins (69, 102).

In addition to these shared properties, differences in replication between viruses have been reported to occur because of structural variation between viral core origins and TAg pro-
teins (4, 5, 47, 51, 52); differences in TAg acetylation that either stimulate replication (109), or regulate TAg stability (73, 84); selective interactions with the host p180 DNA Pol α-prime

mase and p48 prime subunits (9, 94) and with RPA (103); steps in replication beyond initiation (89); and modulation of replication by 5′ and 3′ cis-acting origin proximal sequences that alter activities of replication proteins and determine chro

matin structure and intranuclear DNA localization (14, 29, 63, 91, 100, 104, 106, 109). Such virus- and host-specific features may affect the outcomes of viral infection.

Despite its importance in human disease, BKV DNA replication in human kidney epithelial cells and its role in PVAN are not well understood. Early studies documented that archetypal BKV isolated from human tissues does not replicate in cultured human cells, and consequently, most analyses have utilized naturally occurring BKV variants with genomic alterations that promote growth in cell culture (58, 96). The consequences of such alterations for BKV infection of humans and possible viral pathogenesis are unknown. Robust BKV DNA replication in cell culture requires a 76-base-pair “core origin” very similar to that of SV40 but which does not by itself suffice for DNA replication (when mediated by SV40 TAg) (23, 24). Binding sites for cellular factors that might activate or modulate replication during PVAN are located in the core origin flanking sequences, termed the enhancer (25, 54, 59, 60). Also, although BKV TAg resembles SV40 TAg in its DNA binding domain (42, 85), helicase domain, and interactions with p53 and pRb (33), physical or functional BKV TAg interactions with cellular replication factors such as Pol α-prime subunits p180 and p48, RPA, and topoisomerase I have not been characterized.

Here, we report sensitive assays of BKV replication and comparative studies of BKV replication in human and murine cells and extracts that point to an early, TAg-dependent step as being a likely cause of the replication block in murine cells and extracts, and we speculate about possible mechanisms.

MATERIALS AND METHODS

Viral infections. Primary human renal proximal tubule epithelial (RPTE) cells (Lonza) were maintained in renal epithelial cell growth medium as previously described (1). Murine embryonic fibroblasts (40) were immortalized using the 3T3 protocol (99) as previously described (16) and were maintained in Dulbecco modified Eagle medium (DMEM) (Gibco/BRL) containing 10% fetal bovine serum (FBS) (HyClone) and supplemented with 100 U/ml penicillin and 100 µg/ml streptomycin (Lonza). Both RPTE and 3T3 cells were grown at 37°C in 5% CO2 in a humidified incubator, BKV strains TU and Prot-2 were propagated as previously described (1). Purified stocks of BKV strain TU were produced by infecting Vero cells (ATCC CCL-81) for 4 weeks and harvesting progeny virus by centrifugation through a 20% sucrose cushion followed by centrifugation in a 1.2- to 1.4-g/cm3 cesium chloride gradient (49).

BKV Proto-2 and BKV TU were used to infect 70% confluent RPTE or 3T3 cells and extracts that point to an early, TAg-dependent step as being a likely cause of the replication block in murine cells and extracts, and we speculate about possible mechanisms.
column volumes of buffer A (50 mM Tris-HCl [pH 7.5], 150 mM NaCl, 5 mM KCl, 0.5% MgCl₂, 0.01% Igepal CA630) and then with 20 column volumes of buffer A containing 5 mM imidazole (pH 7.5). BKV TAg was eluted in buffer A containing 500 mM imidazole (pH 7.5), and fractions containing TAg were pooled and dialyzed against 500 ml of buffer containing 20 mM HEPES [pH 7.5], 5 mM NaCl, 1 mM diethytritol (DTT), 0.1 mM EDTA, and 20% glycerol and stored at −80°C. The purity and amount of BKV TAg were determined by SDS-PAGE using Roti-mark standard (150 to 10 kDa; Carl Roth GmbH & Co.) or precasted molecular weight marker proteins (New England Bioscience). Proteins were detected either by staining with Roti-Blue colloidal Coomassie blue staining reagent (Carl Roth) or by Western blot analysis (68). The purity of BKV TAg was estimated as ∼90% by Coomassie blue staining. SV40 TAg, JCV TAg, and mPyV TAg proteins were purified from extracts of insect High Five cells infected with recombinant baculoviruses by immunoaffinity chromatography using monoclonal antibodies PAb101 and 5F (for SV40 and mPyV TAg, respectively) coupled to protein A-Sepharose or by immobilized metal chelate chromatography (for JCV TAg) using TALon resin as described previously (9, 68).

Cellular replication proteins and extracts. Pol α-primase (80), topoisomerase I (92), and RPA (67, 71) were expressed and purified and their concentrations and activities determined as previously described (66). Logarithmically growing adherent HeLa S3 cells in DMEM and suspension FM3A cells in RPMI supplemented with 10% and 5% FBS, respectively, were collected by centrifugation, washed once with phosphate-buffered saline, and then washed extensively with hypotonic buffer (20 mM HEPES-KOH [pH 7.8], 5 mM potassium acetate, 0.5 mM DTT, and 1× phosphatase and protease inhibitors). Cells were homogenized (Dounce homogenizer B pestle, 20 strokes), adjusted to 50 mM NaCl, and incubated on ice for 30 min. Extracts were clarified by centrifugation twice at 20,000 × g for 30 min and stored at −80°C. Extracts of Human TAg cells containing BKV TAg were prepared as described above except that after centrifugation, extracts were dialyzed into 20 mM HEPES (pH 7.8), 5 mM KCl, and 1 mM DTT with 1× protease inhibitors and stored at −80°C.

In vitro DNA replication assays. Replication of DNAs in vitro was assayed as described by Stadlbauer and coworkers with slight modifications (94). Briefly, the reaction mixtures (30 μl) contained 20 mM HEPES (pH 7.8): 7 mM magnesium acetate; 1 mM DTT; 4 mM ATP; 200 μM each UTP, GTP, and CTP; 50 μM dCTP; 100 μM each dATP, dTTP, and dGTP; 40 mM creatine phosphate di-Tris (pH 7.8); 40 μg/ml creatine kinase plus 5 μCi of [α-32P]dCTP (3,000 Ci/mmol); 0.25 μg test plasmid DNA; FM3A or HeLa cell extract (25 to 75 μg of protein); and purified TAg at the indicated concentrations. After incubation for 60 min at 37°C, reaction products were precipitated with cold 10% (wt/vol) trichloroacetic acid containing 2.5% (wt/vol) sodium pyrophosphate, spotted on glass fiber filters (GF/C; Whatman), washed with 1 M HCl, and analyzed by scintillation counting.

The monoplymerase replication assay (89) was assembled on ice with 0.5 μg of pOriBKV DNA or 0.5 μg of pUC-HS DNA (containing the SV40 replication origin) (79), 50 ng topoisomerase I, 100 ng Pol α-primase, and 1 μg RPA in 30 mM HEPES-KOH (pH 7.8); 7 mM magnesium acetate; 0.1 mM EDTA; 0.5 mM DTT; 200 μM each UTP, GTP, and CTP; 4 mM ATP; 100 μM each dATP, dTTP, and dGTP; and 5 μCi of [α-32P]dCTP (3,000 Ci/mmol; Perkin-Elmer) in 40 μl. Purified BKV or SV40 TAg (0.2 μg) was added to start the reaction, and after incubation for 60 min at 37°C, the reaction products were precipitated with cold 10% (wt/vol) trichloroacetic acid containing 2.5% (wt/vol) sodium pyrophosphate, spotted on glass fiber filters (GF/C; Whatman), washed with 1 M HCl, and analyzed by scintillation counting.

RESULTS

Comparison of BKV infection of human and murine cells. Human RPTE and mouse 3T3 cells were infected with the TU and Proto-2 BKV strains and at 4 and 7 dpi, proteins and low-molecular-weight DNA were extracted and analyzed by Western blotting and quantitative real-time PCR, respectively. Although BKV TAg was expressed in both the human and murine cells, viral DNA replication was observed only in the human RPTE cells, with levels increasing between 4 and 7 dpi (Fig. 1). Incubating the infected murine cells for an additional day did not increase viral DNA, and a fluorescent-focus assay revealed that progeny were produced by infected RPTE cells but not by murine cells (data not shown).

Importance of core origin and flanking sequences for species-specific BKV DNA replication. To assess whether the lack of BKV DNA replication in murine cells is dependent upon core origin or origin flanking sequences, chimeric DNAs with the BKV core origin flanked by BKV (B-B-B) or mPyV sequences (B-B-B, B-B-P, and P-B-P) were constructed (Fig. 2) and their replication activity was analyzed following DNA transfection into cells. Exchanging the BKV early and late flanking region with mPyV sequences reduced the efficiency of chimeric DNA replication in human cells (Fig. 2, compare lane 1 with lanes 2 to 4). When transfected into mouse TCMK-1 cells that support mPyV DNA (P-P-P) replication in the presence of mPyV TAg (Fig. 2, lane 9), no replication of DNAs containing the BKV origin was detected (Fig. 2, lane 5), consistent with the results of the viral infection assays described in Fig. 1. Furthermore, none of the BKV-mPyV chimeric templates were replicated (Fig. 2, lanes 6 to 8). Although BKV and mPyV TAg were expressed at similar levels in TCMK-1 cells (data not shown), BKV TAg did not support replication of any BKV origin tested in murine cells, whereas in these cells, mPyV TAg supported replication of its cognate origin (Fig. 2) and of chimeric DNAs with the mPyV core origin (data not shown).
shown). These data indicate that the BKV core origin and its cognate TAg are primary determinants of the lack of BKV replication in murine cells.

**BKV TAg-dependent DNA replication in human cell extracts.** To define the cause of the lack of BKV replication in murine cells, an in vitro BKV TAg-dependent DNA replication system was established. Since insect cells efficiently express polyomavirus TAg proteins capable of supporting cell-free DNA replication (9, 68, 79, 89, 93, 94, 103), BKV TAg was expressed from baculovirus vectors in High Five insect cells and purified (data not shown). Archetype BKV, which is derived from human patients but does not replicate in cultured cells, was used as the source of DNA for these experiments. The insect cell extracts containing BKV TAg promoted BKV DNA replication in vitro in a dose-dependent manner when mixed with HeLa cell extracts (Fig. 3A, bars 1 to 4). Replication also depended on the presence of the BKV core origin (ori\textsubscript{H11001}) (Fig. 3A, compare bars 2 to 4 with bars 6 to 8). Similar results were obtained with purified BKV TAg (data not shown), whose activity was comparable to that of purified SV40 TAg with DNAs containing the SV40 origin (Fig. 3B, bars 3 and 6).

**Restriction of replication of BKV origin-containing DNA.** Reflecting the lack of viral replication observed in cultured cells, murine cell extracts did not support BKV TAg-dependent replication of BKV DNAs, although these extracts supported mPyV TAg-dependent replication of the mPyV origin (Fig. 4, compare bars 2 and 1) and resembled reactions carried out in parallel with SV40 TAg (Fig. 4, bar 3), and assays including mPyV DNA but lacking mPyV TAg (Fig. 4, bar 4). To ascertain the biochemical basis of the lack of BKV DNA replication, the murine extracts were supplemented with human Pol \(\alpha\)-primase (hPol \(\alpha\)-primase), RPA, or topoisomerase I. No incorporation of deoxynucleoside monophosphates (dNMPs) into BKV origin-containing DNA was observed when hPol \(\alpha\)-primase and BKV TAg were added to murine extracts (Fig. 4, bar 6). In contrast, the addition of hPol \(\alpha\)-primase resulted in SV40 TAg-dependent replication of SV40 origin-containing DNAs to about 60% of the level obtained with mPyV TAg and its cognate origin (Fig. 4, compare bars 7 and 1), consistent with published data (90). Similarly, purified human recombinant RPA (hRPA) added to murine extracts did not support replication of BKV DNA (Fig. 4, bar 9); however, addition of hRPA to the murine extracts stimulated mPyV DNA replication with mPyV TAg (Fig. 4, compare bars 1 and 8), suggesting that RPA levels are limiting in these extracts and that the purified hRPA does not inhibit DNA replication per se. This is consistent with previous data indicating that addition of RPA or *Escherichia coli* single-stranded DNA binding protein stimulated SV40 and mPyV DNA replication, respectively, in mouse cell extracts (26, 94). Addition of purified human recombinant topoisomerase I also did not support incorporation of dNMPs into BKV DNA (Fig. 4, bar 11), but it stimulated mPyV TAg-dependent DNA replication (Fig. 4, compare bars 1 and 10) nearly twofold, suggesting that topoisomerase I is also limiting in murine cell extracts, as has been previously observed in human cell extracts (87).

**Murine proteins inhibit BKV TAg-dependent DNA replication.** The lack of BKV TAg-dependent DNA replication in murine extracts was studied with a monomonomerase replication system comprised of purified hPol \(\alpha\)-primase, RPA, and topoisomerase I (89). Addition of each of these purified human proteins individually did not suffice to support incorporation of dNMPs into BKV DNA (Fig. 4, bar 11), but it stimulated mPyV TAg-dependent DNA replication (Fig. 4, compare bars 1 and 10) nearly twofold, suggesting that topoisomerase I is also limiting in murine cell extracts, as has been previously observed in human cell extracts (87).
5C, compare bar 3 with bar 1), and the addition of human extracts to the SV40 system did not influence replication (compare bars 1 and 2). To determine at what step the inhibition occurs, murine extracts were introduced into the monopolymerase system at different stages of replication (Fig. 6A). When added prior to the addition of TAg and human replication factors, murine extracts inhibited BKV TAg-dependent replication by more than 75% (Fig. 6B, compare bars 2 and 3). In contrast, addition of murine extracts at a later stage (after DNA unwinding but prior to the initiation or the elongation reaction) had a lesser inhibitory effect (Fig. 6B, compare bars 4 and 5 with bars 2 and 3). These findings contrast with those for the SV40 system, where little inhibition by murine extracts is observed, regardless of the time of addition (Fig. 6C). However, these results resemble those recently reported with the SV40 monopolymerase system using polypeptides that interfere with the assembly of the initiation complex (98).

**DISCUSSION**

The regulation of BKV replication in human kidney tissues and the cause(s) of BKV reactivation in allografts leading to
FIG. 5. DNA synthesis with purified human proteins in the presence of BKV and SV40 TAg. (A) Incorporation of dNMPs into DNA containing an SV40 (bar 2) or BKV (bar 4) origin of replication in the presence of 200 ng of the respective viral TAg and 100 ng of purified hPol α-primase, 50 ng topoisomerase I, and 1000 ng RPA was measured (monopolymerase DNA replication system). Vectors without a functional viral origin (ori−) served as negative controls (bars 1 and 3). (B) The effect of human and mouse proteins on DNA synthesis by hPol α-primase was determined with a BKV origin of replication. The incorporation of radioactive dNMPs using the BKV origin of replication as a template was measured in the presence of buffer but no additional proteins or with 15 μg human or mouse cell extracts (bars 1, 2, and 3, respectively). DNA synthesis in the presence of DNA lacking an origin of replication served as a negative control (bar 4). (C) The effect of human and mouse proteins on DNA synthesis by human DNA Pol α-primase was determined with an SV40 origin of replication. The incorporation of radioactive dNMPs was determined in the presence of buffer but no additional proteins or with 15 μg human or mouse cell extracts (bars 1, 2, and 3, respectively). DNA synthesis in the presence of DNA lacking an origin of replication served as a negative control (bar 4). Incorporation of dNMPs into DNA was measured by scintillation counting. DNA synthesis was determined in duplicate and repeated three times. The averages from these experiments and the standard deviations are presented.

PVAN are not understood, and combined with the lack of suitable animal models, this makes the development of effective prevention or interventions difficult. A murine model of BKV infection would be of great benefit; however, BKV DNA is not replicated in murine cells. As shown in Fig. 1, BKV can infect murine cells and low levels of BKV TAg are expressed following infection by virions, but no BKV DNA replication occurs. However, these BKV TAg levels are most likely not the cause for the failure of BKV DNA replication in murine cells, since our previous findings indicate that expression of very low levels of BKV TAg in human RPTE cells suffices for viral replication and progeny production (1). The finding that the presence of BKV TAg is not sufficient for BKV DNA replication in murine cells is consistent with the results of BKV DNA replication in murine cells using overexpressed BKV TAg (Fig. 2 and data not shown). Although BKV TAg levels in murine cells were equivalent to those of mPyV TAg expressed in parallel, BKV TAg does not allow BKV DNA replication, whereas mPyV TAg efficiently supported mPyV DNA replication in these cells (Fig. 2). These data reveal that a factor other than the infection of murine cells and the expression level of BKV TAg is the regulatory step in the species specificity of BKV DNA replication.

Analyses of BKV TAg- and mPyV TAg-dependent replication of their cognate origin-containing DNAs in combination with heterologous flanking sequences revealed that the BKV core origin and TAg are primary determinants of the restriction of BKV replication in murine cells. To study the molecular basis of this restriction, a robust BKV TAg-dependent DNA replication in vitro was established with archetype BKV DNA sequences and analyzed with human and murine cell extracts (Fig. 3). As was observed with cellular assays of BKV DNA replication, murine extracts also did not support BKV TAg-dependent DNA replication. Furthermore, addition of replication-active hPol α-primase, RPA, and topoisomerase I to the murine extracts did not promote BKV DNA replication (Fig. 4), in contrast to similar studies of SV40 and mPyV DNA replication in heterologous systems (9, 94). Additional analyses of BKV TAg-dependent DNA replication with a monopolymerase system comprising these three human proteins revealed that murine cell extracts, but not human cell extracts, inhibit BKV DNA replication at an early stage (Fig. 5 and 6), perhaps during the unwinding of the core origin by BKV TAg. Such inhibition is not observed with SV40 TAg-dependent DNA synthesis in the monomerase system. The inhibitory activities may associate with and/or modify BKV TAg so as to interfere with its origin binding or unwinding activities and are consistent with the cell-based replication assays that indicate the BKV core origin sequences bound by TAg are of primary importance for the restriction of replication in murine cells.

The absence of BKV TAg-dependent DNA synthesis in murine extracts, even with the addition of human replication proteins, suggests that the restriction of BKV DNA replication in murine extracts differs from that observed for JCV and SV40 DNA replication. It is noteworthy that JCV TAg is reported to support replication of DNAs with its cognate origin in murine extracts (89). These differences also might be reflected by the origin selectivity of the virus TAg proteins for their cognate origins. All the experiments with murine extracts were carried out under conditions optimal for mPyV DNA replication, even though factors such as RPA and topoisomerase I are rate limiting (Fig. 4). Moreover, the stimulation of incorporation of dNMPs into mPyV DNA by RPA and topoisomerase I, and into SV40 DNA by hPol α-primase, indicates that these human proteins are functional in murine extracts.

Although introduction of the human replication proteins did not restore BKV DNA replication in murine extracts, one cannot rule out the possibility that the lack of BKV DNA replication in murine cells is mediated by an incompatibility of components of the host DNA replication complex. For instance, a murine initiation factor(s) might stably complex with...
BKV TAg, forming an inactive chimeric complex, analogous to what has been observed with SV40 DNA replication inhibition by polypeptides representing the protein-protein interaction regions of replication proteins (97, 98). Later steps of DNA replication involving other components, such as PCNA, replication factor C, and DNA polymerase ε, required for elongation and polymerase switching might also contribute to the restriction of BKV DNA replication.

The replication assays using chimeric templates suggest that sequences flanking the core origin are unlikely to be primary determinants of the restriction of replication. However, chimeric templates might lack some important “cross talk” interactions between core origin flanking sequences and the core origin, which could be vital for DNA replication in vivo. For example, AP-1 stimulates both mPyV and SV40 DNA replication (29, 30, 39, 56, 100) but inhibits JCV replication (43, 75). NF-1 also has been shown to stimulate SV40 DNA replication (15, 63, 100); however, a closer look into different members of NF-1 family proteins revealed that NF-1D, expressed predominantly in permissive glial cells, stimulated JCV virus replication (61), while NF-1A, expressed predominantly in nonpermissive progenitor and HeLa cells, restricted JCV replication in these cells (74). NF-1 binding sites have been identified on BKV origin flanking sequences (11–13, 54), and AP-1 binding sites were reported in enhancers of BKV strains (54, 55). Their influence on BKV DNA replication needs to be elucidated.

FIG. 6. DNA replication with purified human proteins in the presence of BKV TAg and mouse cell extracts. (A) The replication of polyomavirus DNA was biochemically separated into three consecutive reaction steps: the unwinding, initiation, and elongation reactions. In the presence of RPA, topoisomerase I, ATP, and an ATP-regenerating system, viral TAg unwinds viral DNA at 37°C for 30 min (unwinding reaction). To synthesize primers at the unwound origin of DNA replication (initiation reaction), hPol α-primase and the three remaining ribonucleotides were added, and oligoribonucleotide primers are synthesized during the incubation at 37°C for 30 min, whereas no DNA can be synthesized since deoxynucleoside triphosphates are lacking. Finally, deoxynucleoside triphosphates, which include radioactively labeled dCTP to monitor DNA synthesis via scintillation counting, are added and DNA is synthesized at 37°C for 30 min (elongation reaction). Mouse cell extracts capable of supporting mPyV DNA replication were added to the reactions prior to the specified step (as indicated by the arrows). The addition of buffer served as control for the influence of salt and dilutions. (B) Results of the mononucleoside assay using BKV TAg and template containing the BKV origin of replication. (C) Results of the mononucleoside assay using SV40 TAg and template containing the SV40 origin of replication. In panels B and C, bars 1 and 2 represent dNMP incorporation into DNA in the absence and presence of TAg, respectively, but without mouse proteins. For bar 3, mouse cell extracts were added to reaction components before the addition of hPol α-primase and ribonucleotides (prior to unwinding of DNA). For bar 4, mouse cell extracts were added after the unwinding reaction but before the addition of hPol α-primase (prior to initiation of DNA replication). For bar 5, mouse cell extracts were added after initiation of DNA replication but before addition of deoxynucleoside triphosphates (prior to elongation). Incorporation of dNMPs into DNA was measured by scintillation counting. DNA synthesis was determined in duplicate and repeated three times. The averages from these experiments and the standard deviations are presented.
checkpoint. To override the DNA damage response triggered by viral infection, SV40 TAg targets subunits of the MRN complex for degradation through its interaction with CUL7, an E3 ubiquitin ligase (41, 108, 111). It is possible that BKV TAg does not interact with murine CUL7 and therefore cannot mediate the degradation of the murine MRN complex, whereas this mechanism would be functional in human cells and extracts. In addition, components of the DNA replication machinery also participate in the DNA repair pathway (19, 70, 78). It is possible that the DNA damage response triggered by BKV infection of murine cells might differently modulate these components, causing restriction of BKV replication.

Although BKV is closely related to SV40 and JCV, our data indicate there to be distinct sites at which host-specific replication factors may conflict with viral DNA replication. Detailed analysis of the restriction of BKV DNA replication in murine cells may point to processes that maintain BKV DNA replication at low levels until it is stimulated in the transplant setting to activate DNA replication.

ACKNOWLEDGMENTS

This work was supported by NIH grant R21 AI062848 to W.R.F.; by INTEAS, Health Research Board, Ireland, and Science Foundation Ireland grants to H.P.N.; by NIH grant AI060584 to M.J.I.; by an NUIG fellowship to I.T.; and by the F. G. Novy Fellowship to J.R.A. We thank John Lednicky and W. Deppert for kind gifts of reagents and Sarah Scanlon for advice.

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