

Provided by the author(s) and University of Galway in accordance with publisher policies. Please cite the published version when available.

Title	Catalytic potential of selected metal ions for bioleaching, and potential techno-economic and environmental issues: a critical review
Author(s)	Pathak, Ashish; Morrison, Liam; Healy, Mark G.
Publication Date	2017-01-06
Publication Information	Pathak, Ashish, Morrison, Liam, & Healy, Mark Gerard. (2017). Catalytic potential of selected metal ions for bioleaching, and potential techno-economic and environmental issues: A critical review. Bioresource Technology, 229, 211- 221. doi: http://dx.doi.org/10.1016/j.biortech.2017.01.001
Publisher	Elsevier
Link to publisher's version	http://dx.doi.org/10.1016/j.biortech.2017.01.001
Item record	http://hdl.handle.net/10379/6294
DOI	http://dx.doi.org/10.1016/j.biortech.2017.01.001

Downloaded 2024-04-29T06:04:54Z

Some rights reserved. For more information, please see the item record link above.



1Published as: Pathak, A., Morrison, L., Healy, M.G. 2017. Catalytic potential of2selected metal ions for bioleaching, and potential techno-economic and environmental3issues: a critical review. Bioresource Technology 229: 211 – 221.4<u>http://dx.doi.org/10.1016/j.biortech.2017.01.001</u>5

6 Catalytic potential of selected metal ions for bioleaching, and potential techno-

7 economic and environmental issues: A critical review

- 8 Ashish Pathak^a*, Liam Morrison^b, Mark Gerard Healy^a
- ^a Civil Engineering, College of Engineering and Informatics, National University of Ireland,

10 Galway, Ireland.

- ^b Earth and Ocean Sciences, School of Natural Sciences and Ryan Institute, National
- 12 University of Ireland, Galway, Ireland.
- 13

*Corresponding author. Tel.: +353 91495364

15 Email-<u>ashish.pathak@nuigalway.ie</u>

16

17 Abstract

Bioleaching is considered to be a low-cost, eco-friendly technique for leaching valuable 18 19 metals from a variety of matrixes. However, the inherent slow dissolution kinetics and low metal leaching yields have restricted its wider commercial applicability. Recent 20 advancements in bio-hydrometallurgy have suggested that these critical issues can be 21 22 successfully alleviated through the addition of a catalyst. The catalyzing properties of a variety of metals ions (Ag⁺, Hg⁺⁺, Bi⁺⁺⁺, Cu⁺⁺, Co⁺⁺ etc.) during bioleaching have been 23 successfully demonstrated. In this article, the role and mechanisms of these metal species in 24 catalyzing bioleaching from different minerals (chalcopyrite, complex sulfides, etc.) and 25

waste materials (spent batteries) are reviewed, techno-economic and environmental challenges associated with the use of metals ions as catalysts are identified, and future prospectives are discussed. Based on the analysis, it is suggested that metal ion-catalyzed bioleaching will play a key role in the development of future industrial biohydrometallurgical processes.

31

32 *Keywords:* Bioleaching; catalyst; metals; leaching; minerals

33

34 1. Introduction

Bioleaching is a low-cost, green technology for leaching metals from a variety of 35 36 minerals and waste materials (Zeng et al., 2016). Bioleaching has several advantages compared to conventional hydrometallurgy (the leaching of metals with acids or bases) or 37 pyrometallurgy (the leaching of metals using thermal treatment), which have high energy 38 requirements, require a large amount of chemicals, and also produce high levels of 39 40 environmental pollution (Zeng et al., 2013). Currently, a significant portion of many minerals/ores are processed at industrial scale using bioleaching (Walting, 2015). For 41 example, approximately 20-25% of the world's total copper (Cu) is produced using 42 bioleaching (Brierley, 2008). A number of successful commercial bioleaching operations 43 44 exist worldwide, and a Cu bioleaching plant (Morenci mine, USA) with a capacity of up to 230,000 tonnes/year is currently operational (Panda et al., 2015a). 45

Although bioleaching offers many advantages, the relatively slow dissolution kinetics and low metal leaching yield are critical factors that hinder its large-scale application. For example, laboratory bioleaching experiments may last for more than 300

days in order to obtain reasonable metal extraction efficiencies (Munoz et al., 2007), and in
large industrial operations such as Cu⁺⁺ bioleaching, some researchers have taken up to 900
days to obtain a Cu yield of just 60% (Clark et al., 2006). Therefore, research efforts have
been directed towards improving the efficiency of the bioleaching process by improving
process dissolution kinetics and metal leaching yields.

54 The rate of reaction and bioleaching yield may be improved considerably by the 55 addition of suitable catalysts. A 'catalyst' is a substance that lowers the activation energy and thereby increases the rate of reaction. A variety of metal ions (Ag⁺, Hg⁺⁺, Bi⁺⁺⁺, Cu⁺⁺, 56 Co⁺⁺, etc.) and non-metallic catalysts (activated carbon, light illumination, waste 57 newspapers, polyethylene glycol, etc.) have been used to improve bioleaching efficiency 58 59 (Ballester et al., 1990; Liu et al., 2015; Niu et al., 2015; Panda et al., 2015a; Zhou et al., 2015). However, the use of metal ions as a catalyst has gained most attention, as non-60 metallic catalysts have produced relatively poor bioleaching yields of as low as 12.5% 61 (Zhang et al., 2016) and also require large amounts of catalysts of up to 2500 g/kg of ore 62 (Dong et al., 2013) to obtain significant leaching yields. In comparison, metal ions have 63 excellent catalytic properties and therefore it is expected that in future, metal ions will have 64 65 a significant influence on the development of a suitable bioleaching process at commercial scale. 66

Previous review articles on bioleaching have primarily dealt with the fundamentals and mechanism of bioleaching (Donati et al., 2016), the microorganisms involved and the types of minerals bioleached (Das et al., 2011; Panda et al., 2015b; Donati et al., 2016), and the mineral-microbial interaction (Diao et al., 2014). However, to date, the role and potential of metal ions in promoting bioleaching efficiency has not been reported. Since the

application of microbial-assisted bioleaching is emerging, and to date, no systematic review 72 73 has addressed the potential of metal ions as catalysts in bioleaching, the aim of this paper is to (1) identify the metal ions which have the potential in catalyzing bioleaching of different 74 ores/minerals/waste/end-of-life materials (2) describe the role and catalytic mechanism of 75 76 these metal ions and how they improve the bioleaching efficiency of 77 minerals/ores/waste/end-of-life materials (3) select the most appropriate metal ion for 78 particular ores/minerals (4) evaluate the different techno-economic issues and 79 environmental challenges associated with the use of these metal ions, which need to be 80 overcome before the process may be applied on an industrial scale. This information will 81 assist metallurgists in understanding the catalytic properties of metal ions for improving the 82 efficiency of the bioleaching process.

83

84 2. Bioleaching mechanisms and microorganisms involved

Bioleaching involves extraction of metals from mineral ores using biological means 85 (Karthikeyan et al., 2015). A wide variety of microorganisms 86 such as chemolithoautotrophic bacteria, heterotrophic bacteria, archea and fungi, play an important 87 role in bioleaching (Panda et al., 2015b). Chemoautotrophic bacteria such as 88 Acidithiobacillus thiooxidans (At. thiooxidans) and Acidithiobacillus ferrooxidans (At. 89 90 ferrooxidans) are the most dominant and industrially used microorganisms to extract the metals from ore and minerals (Feng et al., 2016). Bioleaching microorganisms derive the 91 92 energy required for their growth from the oxidation of ferrous iron and reduced (inorganic) 93 sulfur compounds in acidic environments. During bioleaching, microorganisms catalyze the oxidation of ferrous iron (Fe^{++}) and reduced sulfur compounds as per Eqs. (1-2). The 94

oxidation of Fe^{++} and reduced sulfur compounds lead to the generation of biologically produced ferric ion (Fe^{+++}) and sulfuric acid (H_2SO_4), respectively (Ma et al., 2017). The generated H_2SO_4 and/or Fe^{+++} act as oxidants and oxidize the metal sulphides (redoxolysis) and/or solubilize the metal sulfides and oxides as per Eqs. (3-5).

99
$$2\text{FeSO}_4 + \text{H}_2\text{SO}_4 + \frac{1}{2}\text{O}_2 \xrightarrow{\text{Microorganisms}} (\text{Fe})_2(\text{SO}_4)_3 + \text{H}_2\text{O}$$
 (1)

100 $S^0 + H_2O + 1.5O_2 \xrightarrow{\text{Microorganisms}} H_2SO_4$ (2)

101
$$H_2SO_4 + MeS \longrightarrow H_2S + MeSO_4$$
 (3)

$$102 \quad H_2SO_4 + MeO \longrightarrow H_2O + MeSO_4 \tag{4}$$

103
$$Fe_2(SO_4)_3 + MeS + H_2O + 3/2O_2 \longrightarrow Me^{++} + SO_4^{--} + 2FeSO_4 + H_2SO_4$$
 (5)

104 Species of *Acidithiobacilli* thrive in acidic conditions (pH 0.5-3) and are also 105 tolerant of high concentrations of a variety of metal ions present in different leaching 106 environments. Other microbial species may be used in bioleaching, including *Acidophillum* 107 *cryptum*, *Thiobacillus thioparus* and *Halothiobacillus halophilus*. However, they are not 108 widely studied, as they only grow at a relatively high pH range (2.5-8.0) in which efficient 109 leaching does not take place (Chen and Lin, 2009; Xu et al., 2010; Vainshtein et al., 2015).

Besides chemoautotrophic bacteria, heterotrophic bacteria and fungi have also been used for the treatment of non-sulphides and oxide materials. However, in these cases, metal leaching is performed as a result of production of organic acids and complexing compounds secreted into the system (Vakilchap et al., 2016). A detailed overview of the different microorganisms involved in various bioleaching environment is provided by Krebs et al. (1997) and Panda et al. (2015b), amongst others.

116

117 **3. Bottlenecks in bioleaching process**

In recent years, the mining industry has made significant efforts to develop eco-118 119 friendly and low cost bio-hydrometallurgical operations (Walting, 2015). However, certain bottlenecks still exist which hinder its wider commercial applicability. The process kinetics 120 are currently much too slow for it to be economical. Longer periods of operation are 121 122 required compared to traditional methods of leaching to obtain reasonable yields (Clark et 123 al., 2006). For example, Cerda and Ohlbaum, 2008 reported 65-80% Cu recoveries from 124 chalcosite and covellite after a bioleaching time of up to 500 days in industrial operations. 125 Laboratory-scale experiments, performed in column reactors under mesophilic conditions, 126 also found that more than 300 days of bioleaching was required to obtain 7.3 to 27.1% of 127 Cu recovery from enargite and covellite-rich composites (Lee et al., 2015). Further, the 128 efficiency of the process is affected by atmospheric conditions and slight changes in parameters such as temperature can adversely affect the process efficiency (Karimi et al., 129 130 2010). In addition, with industrial wastes (such as spent batteries), bioleaching is mostly conducted with low pulp density i.e. 1% w/v (Zeng et al., 2013). The process efficiency 131 132 decreases at higher pulp density due to the presence of alkaline wastes or metal toxicity to leaching microorganisms (Niu et al., 2015). The maintenance of optimum particle size in 133 the reactor is another concern, as the presence of very fine particles (<25 µm) can 134 135 negatively impact the activity of cells due to attrition caused by increased particle-particle 136 collision (Nemati et al., 2000).

137

4. Role and catalytic mechanism of metal ion in bioleaching

Different materials have been reported to catalyze the bioleaching through varying
modes of action (Table 1). Generally, physical, chemical and electrochemical processes are

involved in the dissolution of minerals in aqueous solutions. In the case of physical 141 142 processes, there is no chemical transformation and salts are water soluble. The dissolution 143 of NaCl in water is such an example. In chemical processes, the crystalline solid is 144 insoluble in water, but becomes soluble in the presence of a certain ion in solution. 145 Chemical dissolution of minerals proceeds in two steps: (1) a physical process which establishes an equilibrium between the aqueous phase and mineral solids, and (2) a 146 147 chemical reaction (in the vicinity of the solid) between the ions that just left the solid and 148 the reagent present in the aqueous phase. The third process involved in the dissolution of 149 minerals is electrochemical leaching. These are coupled redox reactions, which change the oxidation state of the minerals and accelerates the dissolution in the leaching medium 150 151 (Habashi et al., 1983). Among these processes of mineral dissolution, the electrochemical mechanism is well established and considered to be the most important. Habashi (1983) 152 postulated that the electrochemical mechanism of sulfide dissolution is affected by the (1) 153 presence of impurities in the crystalline structure that modify the conductivity of the 154 155 mineral solid (2) formation of galvanic couples on the basis of the various mineralogical species present in the ore, and (3) presence of certain ions in solution, which, as they come 156 into contact with the surface of the solid, modify its electrochemical behavior. Most 157 research has been focused on this third factor i.e., the addition of suitable catalytic ions 158 159 which form a redox couple to influence the reactions between the metal sulphide and the oxidizing medium. Using this mechanism, metal ions modify the electrochemical behavior 160 and catalyze the dissolution of sulfides in the leaching medium. Ballester et al. (1990, 1992) 161 162 also suggested that dissolution of different metals from mineral sulfides can be promoted by the addition of soluble external ions. These metal ions cause the formation of a metal 163

sulfide, which dissolves the original sulfide mineral either by galvanic action or substitution in crystal lattice. However, the catalyze effect of a particular metal ion during leaching will be different for each mineral/ore. This is due to fact that different mineral sulfides have different rest potentials (mV), which will impact the galvanic interaction between the metal ion catalyst and mineral solids. In addition, each metal ion poses a unique ability to activate the particular mineral sulfide surface, displacing the ions from the target mineral solid.

170 The first study on the catalytic effects of metal ions in the leaching medium 171 (without bacteria) was conducted by Scott and Dyson (1968), who studied the effect of various metal ions on the dissolution kinetics of ZnS. Further studies were conducted by 172 Mulak (1987), who examined the catalytic effect of Cu^+ , Cu^{++} and Fe^{3+} during HNO₃ 173 leaching of synthetic Ni₃S₂, and observed that these metal ions catalyzed the dissolution of 174 mineral solids by several orders of magnitude. Since then, several studies on the catalytic 175 effect of metal ions on the dissolution of metals in the leaching medium have been reported 176 in the literature (both in the presence of microorganisms - bioleaching - and without 177 microorganisms). The first of these bioleaching studies was reported by Mcelroy and 178 Duncan (1974), who found that the addition of small quantities of Ag⁺ during chalcopyrite 179 bioleaching with At. ferrooxidans dramatically enhanced the Cu leaching yield as opposed 180 to without Ag^+ . 181

182

183 5. Catalytic role and mechanisms of different metal ions

184 5.1 Role of Ag^+

A variety of metal ions such as Ag⁺, Hg⁺⁺, Bi⁺⁺⁺, Cu⁺⁺, Co⁺⁺, etc. have been used successfully as catalysts in various bioleaching studies (Table 2). Among these metal ions,

 Ag^+ has received the most attention. Different Ag compounds such silver sulfate (Ag₂SO₄), 187 silver nitrate (AgNO₃) and silver chloride (AgCl), have been used as a source of Ag⁺ in the 188 bioleaching medium. Although Ag⁺ has been found to be efficient in enhancing the 189 dissolution kinetics and leaching yields of metals from a wide variety of minerals and 190 matrices, the majority of these bioleaching studies using Ag⁺ have been reported for Cu-191 192 bearing ore and/or minerals such as chalcopyrite (Kuwazawa and Miura 2012; Abdollahi et 193 al., 2015). Bioleaching of chalcopyrite (the principal source of Cu in the mining industry) is 194 difficult due to the formation of secondary solid phases and jarosite-type precipitates, 195 which contribute to surface passivation (Feng et. al., 2015; Panda et al., 2015b). The role of Ag⁺ during chalcopyrite bioleaching is to form a film of silver sulfide (Ag₂S) on the 196 197 chalcopyrite surface, which alleviates the surface passivation through the formation of a less tenacious and more porous layer of elemental sulfur (S^0) . The new film, therefore, 198 improves the semi-conductor properties of chalcopyrite, and facilitates the electronic 199 transfer to the oxidant. The presence of Ag₂S in the bioleached chalcopyrite has been 200 confirmed through X-ray diffraction by Hu et al. (2002) (Suppl. Fig. A1). Scanning 201 Electron Microscopy (SEM) micrographs have shown the formation of Ag₂S precipitates 202 203 on the chalcopyrite surface (Abdollahi et al., 2014) (Suppl. Fig. A2).

Ag⁺ initially forms Ag₂S on the chalcopyrite surface, followed by Fe^{3+} mediated oxidation:

206
$$\operatorname{CuFeS}_2 + 4\operatorname{Ag}^+ \longrightarrow \operatorname{Cu}^{++} + \operatorname{Fe}^{++} + 2\operatorname{Ag}_2 S$$
 (6)

207 $2Ag_2S + 4Fe^{+++} \longrightarrow 4Ag^+ + 4Fe^{++} + 2S^0$ (7)

The silver ion recirculates between the solution and solid phase reactions, and the overallsum of the chalcopyrite reaction yields elemental S:

- 210 $CuFeS_2 + 4Fe^{+++} \longrightarrow Cu^{++} + 5Fe^{++} + 2S^0$ (8)
- 211 Elemental Ag can also form during the process:

212 $CuFeS_2 + 4Ag^+ \longrightarrow Cu^{++} + Fe^{++} + 4Ag^0 + 2S^0$ (9)

The role of bioleaching bacteria in this process is to oxidize the Fe^{++} into Fe^{+++} , which oxidizes the Ag₂S film.

The first report on the catalytic effect of Ag⁺ during bioleaching was published as a 215 216 US patent developed by the researchers at the British Columbia Research Council (Mcelroy 217 and Duncan, 1974). In this patent, it was reported that during bioleaching with At. ferrooxidans, the addition of only 0.5 g Ag⁺/kg of ore promoted the Cu yield to 88% 218 compared to 50% without Ag⁺. It was also found that instead of soluble silver salts, 219 insoluble salts such as Ag₂S can also be used. However, the quantity of Ag⁺ required to 220 achieve a significant increase in bioleaching yield will be relatively higher using Ag₂S 221 $(0.700-7.0 \text{ g Ag}^+/\text{kg of chalcopyrite concentrate}).$ 222

Since, Ag⁺ salts are expensive, researchers also tried to use other forms of Ag in order to develop a cost-effective bioleaching process. Hu et al. (2002) conducted bioleaching using Ag-concentrate and found that addition of 20 g Ag-concentrate/kg of ore dramatically increased the Cu yield to 67% compared to only 24.3% without using Agconcentrate. The promising results achieved using relatively cheaper Ag-bearing concentrates suggested that the industrial application of the Ag-catalyzed process can be enhanced using a cheaper source of Ag.

230 Most of these initial studies using Ag⁺ as catalysts were conducted using mesophilic 231 bacteria such as *At. ferrooxidans, At. thiooxidans and Leptospirilum ferrooxidans (L.* 232 *ferrooxidans).* However, it has been reported that rate of bioleaching in different minerals

by thermophilie and moderate thermophiles is higher than mesophiles (Devecei et al., 2004) 233 due to faster reaction kinetics. Romano et al. (2001) reported that bioleaching yield of 234 chalcopyrite under mesophilic conditions was about 50%, whereas it increased to 100% 235 236 with thermophilic microorganism after three weeks of experiments. To harness the higher kinetics, Ag⁺-catalyzed bioleaching was also explored with moderate thermophilic and 237 238 thermophilic microorganisms. Gomez et al. (1999) used shake flask bioleaching with mixed 239 moderate thermophilic bacteria, and reported that the Cu yield increased three-fold in the presence of 0.1 to 0.5 g Ag⁺/kg compared to without the addition of Ag⁺. Moreover, the 240 catalyzing effects of Ag⁺ were higher in moderate thermophilic bioleaching compared to 241 242 mesophilic bioleaching (Abdolllahi et al., 2015). Gomez et al. (1999) reported that in the presence of Ag⁺, it took only 5 days of moderate thermophlic bioleaching to obtain about 90% 243 Cu yield from chalcopyrite-molybdenite concentrate, whereas it took more than 10 days to 244 achieve a similar Cu yield with mesophilic bioleaching. In addition, relatively smaller 245 amounts of Ag^+ (0.77 g Ag^+ / kg of ore) were required to obtain a similar Cu yield using 246 moderate thermophilic bioleaching compared to mesophilic bioleaching (6.7 g Ag⁺/ kg of 247 ore). Similar findings were observed by Abdollahi et al. (2014) in the case of Ag⁺-catalyzed 248 249 thermophilic bioleaching, who found that 75% Cu was leached from the concentrate compared to only 27% with mesophilic temperatures. The reasons for the different effects 250 251 of catalyst at different temperatures were not explained in these bioleaching studies. However, in the case of Ag⁺-catalyzed chemical leaching (without bacteria) a number of 252 factors such as higher solubility of silver complex, higher regeneration of Ag₂S by Fe³⁺ and 253 254 reduced jarosites precipitation, were found to be responsible for higher leaching yield (Guo

et al., 2011). Further studies are required to understand the mechanism and biochemicalaspects of catalyst-based bioleaching at different temperatures.

Most of the initial bioleaching studies using Ag⁺ were conducted in shake flasks. 257 However, the type of reactor may play an important role, as process efficiency may suffer 258 259 in large reactors such as columns due to the restriction of gas transfer in the columns (static 260 system) compared to shake flasks and stirred reactors (agitated types) (Munoz et al., 2007). 261 Nevertheless, column studies give useful information on the effect of different variables on 262 bioleaching process for long periods of time and under conditions close to those found in heap leaching practice. Munoz et al. (2007) investigated the effectiveness of Ag⁺-catalyzed 263 bioleaching process in a stirred reactor and column reactors for treating low-grade Cu ore. 264 265 They found that the addition of 7.1 g of Ag^+/kg of ore in the stirred tank yield 89.3% of Cu compared to 28% without Ag⁺. Similarly, in the column reactor, they found that at different 266 concentrations of Ag^+ , the Cu recovery was at least double that in the absence of Ag^+ (30%). 267 Moreover, the particle size of ore was also found to be an important parameter and use of 268 smaller ore size favors the higher bioleaching yield due to relatively higher surface area of 269 the ore. Therefore, during silver-catalyzed bioleaching, the economics of crushing to finer 270 size versus Cu extraction would have to be evaluated to determine an optimum particle size. 271

Although the majority of studies using Ag^+ as a catalyst were conducted with Cubearing minerals and ore, researchers also exploited the catalytic properties of Ag^+ for treating materials other than Cu. For instance, Guo et al. (2011) tested the process effectiveness with the As-bearing mineral, realgar (As₂S₂), using both mesophilic (*At. ferrooxidans*) and thermophilic *Sulfobacillus sibiricus* (*S.s*), and observed that in the absence of Ag^+ , very little As₂S₂ was leached during bioleaching with either *At.*

ferrooxidans (0.2-0.5%) or S.s (1.5-2.2%). On the contrary, the addition of Ag⁺ enhanced 278 279 the dissolution of As to 24.4% and 15.3% using At. ferrooxidans and S.s, respectively.

As Ag is a potentially toxic metal (David et al., 2008), the addition of Ag⁺ during 280 bioleaching has been reported to cause a decrease in the bacterial growth rate and 281 282 subsequent iron-oxidation rate. To overcome this problem, a process in which corn steep liquor (CSL) is added along with Ag⁺, was recently patented (Kuwazawa and Miura, 2012). 283 284 The authors reported that CSL acted as a source of organic nitrogen to the leaching media 285 and due to its addition, the iron-oxidizing capacity of iron-oxidizing bacteria was improved. This caused a significant improvement in the leaching yield of Cu. It was found that the 286 addition of 10 mg/L of Ag⁺ and 0.1 g/L of CSL led to dissolution of more than 80% of Cu 287 as compared to less than 10% in the absence of Ag^{+.} They also observed that CSL can 288 either be added as an immersion solution or as powder. 289

However, until recently, the catalytic use of Ag⁺ was limited to treat natural mineral 290 and ores. In a recent study, it was reported that the bioleaching yield of industrial waste 291 material and end-of-life products such as spent lithium batteries, can also be improved by 292 the addition of Ag⁺ (Zeng et al., 2013). It was observed that the addition of only 2.0 g of 293 Ag^+/kg of ore with At. ferrooxidans, led to dissolution of 98.4% Co. In the absence of Ag^+ , 294 the leaching yield of Co was only 43.1%. Zeng et al. (2013) found that the catalytic role of 295 Ag⁺ involved promoting the formation of the AgCoO₂ intermediate. This intermediate 296 $AgCoO_2$ was oxidized by bio-produced Fe^{3+} , resulting in higher yield of Co. The role of 297 the bacteria is to oxidize the Fe^{2+} back to Fe^{3+} . Chen and Lin (2009) also utilized the 298 catalyzing property of Ag⁺ for decontamination of riverine sediments that were 299 contaminated with different metals (Cu, Ni, Zn, Pb and Mn). In this study, a higher pH 300

reduction rate was observed during bioleaching with Ag^+ . It was also observed that the Ag^+ added was incorporated into the lattice of the initial sulfide through a cationic interchange reaction. This resulted in the short lag phase and high metal yield during bioleaching as opposed to without Ag^+ . The best results were achieved by the addition of 1.5 g of Ag^+/kg of sediment, beyond which a decrease in leaching rate constant for these metals was observed.

307



Although Hg⁺⁺ is considered to be extremely hazardous in the natural environment, 309 its catalytic properties have been exploited by many researchers to improve bioleaching 310 311 kinetics. Ballester et al. (1990) was perhaps one of the first researchers to report the catalytic effects of Hg⁺⁺ during bioleaching of the complex sulfide concentrate (FeS₂, 312 CuFeS₂, ZnS and PbS) using At. ferrooxidans. Their study suggested that the addition of 2 313 g Hg++/kg of concentrate (in the form of HgSO4) during bioleaching enhanced the 314 dissolution of Cu to 80% compared to only 25% without Hg⁺⁺ from the complex sulfide 315 concentrate. These authors speculated that the catalytic effect of Hg⁺⁺ was due to its 316 tendency to replace the Cu from the sulfide lattice. The sulfide formed as a result of the 317 displacement reaction on the surface of the complex sulfide concentrate, acted as a cathode 318 319 with respect to the chalcopyrite, leading to the rapid dissolution of the mineral chalcopyrite. The changes in chalcopyrite surface during Hg⁺⁺ catalyzed bioleaching were examined 320 using SEM and auger electron spectroscopy (AES) (Gomez et al., 1997). They reported that 321 Hg⁺⁺ were incorporated onto the mineral surface and formed a thin coating of HgS. The 322 standard Gibb's energy ($\Delta G^0 r$) calculated for the reaction between chalcopyrite and Hg⁺⁺ 323

has also shown that the formation of HgS is thermodynamically possible. The mechanism through which Hg^{++} catalyzes bioleaching is presented in Eqs. (10-12). As per reaction mechanism, Hg^{++} first reacts with chalcopyrite and produces a thin coating of HgS.

327
$$CuFeS_2 + 2Hg^{++} \longrightarrow 2HgS + Cu^{++} + Fe^{++}$$
 (10)

The produced HgS can be easily dissolved by bioprdouced Fe^{+++} , which also leads to the regeneration of Hg⁺⁺.

$$330 \quad Fe^{+++} + HgS \longrightarrow Hg^{++} + S^0 + Fe^{++}$$
(11)

The produced Fe^{++} is oxidized to Fe^{+++} by leaching bacteria, which is recirculated in the leaching media.

$$333 \quad \mathrm{Fe}^{++} \xrightarrow{\mathrm{bacteria}} \mathrm{Fe}^{+++} + 1 \,\mathrm{e}^{-} \tag{12}$$

Ballester et al. (1992) further argued that the concentration of Hg^{++} is an important 334 parameter that determines the efficiency of the bioleaching process. By employing different 335 concentrations of Hg⁺⁺ ranging from 0.04 to 1 g Hg⁺⁺/kg of ore, they found that 0.2 g 336 Hg^{++}/kg of ore was the optimum Hg^{++} concentration for enhancing the bioleaching yields of 337 Cu and Zn from the complex sulfide. Moreover, using 0.2 g Hg^{++}/kg of ore, the bioleaching 338 period was also shortened to about 8 days compared to 21 days for the non-Hg⁺⁺ inoculated 339 bioleaching experiment to achieve a similar Cu yield (20%). These authors also postulated 340 that a too low Hg⁺⁺ concentration (0.04 g Hg⁺⁺/kg of ore) during bioleaching may not 341 induce desired changes, whereas a concentration beyond 0.2 g Hg⁺⁺/kg of ore may not 342 further improve the dissolution kinetics. This is due to the fact that at higher concentrations, 343 the Hg⁺⁺ may impart toxicity to mesophilic Acidithiobacilli, which can suppress the 344 leaching. The toxic effects of Hg⁺⁺ are more pronounced with thermophilic archaea 345 (Sulfolobus sp.), which poses less resistance to metal toxicity compared with mesophilic 346

microorganisms. The toxic effect of Hg⁺⁺ at higher concentration during bioleaching was 347 also highlighted by Escudro et al. (1993), who confirmed that 0.1 g Hg⁺⁺/kg of ore is the 348 optimum concentration at which Hg⁺⁺ successfully enhanced the Cu yield from complex 349 sulfide to 58% compared to only 29% using 2.0 g Hg⁺⁺/kg of ore. Besides Cu leaching, the 350 addition of Hg⁺⁺ has also been shown to promote the simultaneous dissolution of Zn from 351 complex sulfide, and about 85% of Zn was leached in the presence of 0.1 g Hg⁺⁺/kg of ore 352 compared to 65% without Hg⁺⁺. Moreover, the dissolution kinetics (250 hours instead of 353 354 400 hours) were greatly improved for achieving similar leaching of Zn (65%).

- 355
- 356 5.3 Role of Bi^{+++}

Along with Ag⁺⁺ and Hg⁺, the catalytic properties of Bi⁺⁺⁺ during bioleaching were 357 also first documented by Ballester et al. (1990). In their study on bioleaching of sphalerite 358 and complex sulfide concentrate using At. ferrooxidans, the authors observed that the 359 addition of Bi⁺⁺⁺ as Bi(NO₃)₃ had an catalyzing effect on Zn dissolution from sphalerite and 360 about 78.4% of Zn was bioleached in the presence of Bi⁺⁺⁺ compared to only 50% without 361 it. The catalytic effect of Bi⁺⁺⁺ was explained on the basis of its tendency to influence 362 electrochemical dissolution process, so that it added the anodic behavior of the mineral. In 363 another study, Mier et al. (1994) observed that the role of Bi⁺⁺⁺ during bioleaching was to 364 suppress the formation of phosphate compounds of Fe^{3+} , which otherwise precipitate in the 365 leaching medium. This results in higher availability of free Fe³⁺ and, therefore, higher 366 oxidizing properties during bioleaching, which are helpful in dissolution of Cu from 367 chalcopyrite. The mechanism through which Bi⁺⁺⁺ catalyzes the chalcopyrite bioleaching is 368 presented in Eqs. (13-15) (Mier et al., 1994). Eq. (13) is purely a chemical reaction, 369

whereas Eq. (14) becomes prominent when bacterial growth takes place. The role of bioleaching bacteria in this process is to oxidize the Fe^{++} and S^0 into Fe^{+++} and H_2SO_4 , respectively. The produced Fe^{+++} oxidizes the chalcopyrite, whereas H_2SO_4 maintains the acidity of the leaching medium.

374
$$CuFeS_2 + 4H^+ + O_2 \longrightarrow Cu^{++} + Fe^{++} + 2H_2O + 2S^0$$
 (13)

375
$$CuFeS_2 + 4Fe^{+++} \longrightarrow Cu^{++} + 5Fe^{++} + 2S^0$$
 (14)

The role of Bi^{+++} is to combine with PO_4^{3-} and prevent the possible formation of ferric phosphate precipitates (FePO₄, FeHPO₄, etc.), which otherwise precipitate in the leaching medium and reduce the oxidation potential of Fe⁺⁺⁺/Fe⁺⁺ couple.

$$379 \quad \text{Bi}^{+++} + \text{PO}_4^{3-} \longrightarrow \text{Bi}\text{PO}_4 \tag{15}$$

However, it has also been reported that the addition of Bi^{+++} during mesophilic bioleaching increased the bioleaching rate of both chalcopyrite and sphalerite, but not the final leaching yield (Ballester et al., 1992). The catalytic role of Bi^{+++} has also been investigated by these researchers during thermophilic bioleaching of chalcopyrite. It was reported that the addition of Bi^{+++} enhanced the Cu yield to 90% compared to 80% without it and that the presence of Bi^{+++} also promoted bacterial growth (Ballester et al., 1992).

386

387 5.4 Role of Cu^{++}

Many researchers have tested the catalytic properties of Cu^{++} during bioleaching due to its relatively cheaper cost and easy availability. Chen et al. (2008) used Cu^{++} as a catalyst for treating a Zn containing mineral 'marmatite'. It was reported that in the presence of 5 g Cu^{++}/kg of ore, about 73% Zn was leached compared to 65% without Cu^{++} . The enhanced leaching yield of Zn was attributed to the electrochemical effect of Cu^{++} on bioleaching. It was postulated that during bioleaching, some Cu^{++} substitutes Zn^{++} and bonded the crystal lattice of marmatite. The CuS formed during bioleaching shows higher electrode potential and improves the conductivity of the mineral surface, so Zn dissolves preferentially as an anode. In addition, iron sulfide, elemental sulfur and jarosite were formed on the marmatite surface during bioleaching. These products formed a passivated layer, which suppressed the bioleaching process. The Cu⁺⁺ catalyzed the bio-oxidation of S⁰, which eliminated the barrier and led to the increased bioleaching of marmatite.

400 In addition to promoting the bioleaching efficiency of natural ores and minerals, Cu⁺⁺ may also be effective in assisting the bioleaching of an industrial end-of-life waste 401 402 material such as spent lithium-ion batteries (Zeng et al., 2012). Zeng et al. (2012) reported that almost 99.9% of Co present in a sample was bioleached in the presence of 75 g of Cu⁺⁺ 403 / kg of ore after 10 days of bioleaching, whereas only 43.1% of Co was bioleached in the 404 same period of time without Cu⁺⁺. The catalytic action of Cu⁺⁺ was due to the formation of 405 an intermediate product (CuCo₂O₄) on the LiCoO₂ surface as a result of cationic exchange 406 reactions. This intermediate CuCo₂O₄ was easily dissolved by bio-produced Fe⁺⁺⁺, leading 407 to the higher bioleaching yield of Co as per Eqs. (16-18). 408

409
$$\operatorname{Cu}^{++} + 2\operatorname{LiCoO}_2 \longrightarrow \operatorname{CuCo}_2O_4 + 2\operatorname{Li}^+$$
 (16)

410
$$CuCo_2O_4 + 6Fe^{+++} \longrightarrow 6Fe^{++} + Cu^{++} + 2O_2 + 2Co^{++}$$
 (17)

411
$$4Fe^{++} + O_2 + 4H^+ \longrightarrow 4Fe^{+++} + 2H_2O$$
 (18)

- The changes in the particle size and morphology of bioleached residues in the presence of
 Cu⁺⁺ are presented in Fig. A3 (suppl.).
- 414 Besides spent lithium-ion batteries, spent Zn-Mn batteries have also been treated 415 successfully in the presence of Cu^{++} . Niu et al. (2015) reported that the Cu^{++} has the ability

to enhance the dissolution kinetics of Zn and Mn from spent Zn-Mn batteries. Moreover, 416 417 this process can be performed at a higher pulp density (10%), which may enhance the 418 industrial application of the process. It was also reported that the leaching kinetics followed the chemical reaction-controlled model. While conducting bioleaching in the presence of 8 419 g Cu⁺⁺/kg of ore, the leaching yield of Zn increased to 62.5% compared to 47.7% without 420 Cu⁺⁺. Similarly, the bioleaching yield of Mn also increased to 62.4% compared to 30.9% 421 without Cu⁺⁺. It was observed that the Cu⁺⁺ formed a possible intermediate CuMn₂O₄, 422 which was oxidized by bio-produced Fe^{3+} . 423

- 424
- 425 5.5 Role of Co^{++}

Very few studies have used Co⁺⁺ as catalytic ion in bioleaching operation, owing to 426 its higher cost compared to Cu⁺⁺ and Bi⁺⁺⁺. Moreover, At. ferrooxidans is susceptible to 427 higher concentrations of Co⁺⁺ (Mohseni et al., 2011). The catalytic properties of Co⁺⁺ were 428 recognized in one of the earlier bioleaching studies conducted on the use of different metal 429 ions as a catalyst. Ballester et al. (1990) reported that the addition of Co⁺⁺ in the form of 430 CoSO₄.7H₂O significantly improved the bioleaching yield of Zn from the sphalerite 431 concentrate. After bioleaching for 400 hours with At. ferrooxidans, only 50% Zn could be 432 leached from the sphalerite concentrate. The addition of 2 g Co⁺⁺/kg of ore during 433 bioleaching enhanced the leaching yield of Zn from 50% to 68.4%. Similar findings were 434 made by Escudero et al. (1993), who conducted bioleaching of a complex sulfide 435 containing Cu, Pb and Zn. Bioleaching was conducted in a stirred glass reactor using a 436 437 mixed culture of bacteria (At. ferrooxidans, At. thiooxidans, L. ferrooxidans). The results showed that the bioleaching yield of Cu was only 22% in the absence of a catalyst. 438

However, there was a drastic increase in the Cu recovery in the presence of Co^{++} and the Cu leaching yield reached to 52% compared to 22% without Co^{++} . A similar increase was also observed in the case of Zn, where about 92% of Zn was leached in the presence of Co⁺⁺ compared to 65% without it. The catalytic effect of Co⁺⁺ was also attributed to its electrochemical interaction with the mineral matrix, which favored the anodic behavior of the sulfides during its reaction.

445

446 **6. Key challenges**

Metal ion-catalyzed bioleaching offers great opportunities for the recovery of metals from a variety of minerals and ores. During bioleaching, these metal ions exhibit catalytic properties at wide range of operating conditions (Table 3). However, despite its high efficiency, to date, the process has not been developed for larger scale applications. Some of the key challenges which need to be overcome are now detailed.

452

453 *6.1 Process economy*

Generally, the capital costs of bio-mining operations are less than those of smelting 454 operations. Dew et al. (1997) reported that the capital costs for a bio-oxidation plant of gold 455 ores was about two-fold lower than roasting or pressure oxidation, and that operating costs 456 457 were also lower. In the case of Cu bioleaching, the operating costs (in 2002) for leaching dumps and in-place ore was calculated to be between US \$ 0.18 and US \$ 0.22/ pound of 458 cathode copper (Pradhan et al., 2008). These costs are competitive in comparison to the unit 459 460 costs of smelting/refining. However, operating costs of bioleaching operations may vary depending on a number of factors such as plant location, cost of services at particular sites 461

etc. (Du Plessis et al., 2007). Cognizance of operating costs (such as addition of catalytic 462 463 ions) is therefore important, considering that most of the metal catalysts used during bioleaching are precious or economically valuable commodities. Munoz et al. (2007) 464 concluded that Ag⁺-catalyzed bioleaching is economical and technically attractive. They 465 reported that the cost of Ag^+ addition per unit cost of Cu in the ore equates to only 6% of 466 467 the commercial price of Cu. Evaluating the effectiveness of this process in the current 468 scenario, the approximate price of Ag in the third quarter of 2016 (June-Sept 2016) is USD 469 \$ 670/kg (NASDAQ). Similarly, the average price of Cu in the third quarter of 2016 at the 470 London Metal Exchange (LME) is about USD \$4.85/kg. Therefore, the cost of Ag addition (1g Ag/kg of Cu) per unit cost of Cu in the ore equates to 0.67 USD /kg of Cu (1 g Ag \times 471 472 USD \$ 670/1000 g). This is equivalent to only 14% (USD 0.67/ USD 4.85×100) of the current international price of Cu. A detailed analysis involving the cost of different metal 473 474 catalyst added during bioleaching, their concentrations and their performance in increasing the bioleaching yield, are provided in Table 4. The cost has been calculated by considering 475 price (in USD \$/kg) of metals catalyst (June-Sept 2016) as 670, 4.85, 26, 2.3, 0.018, 2.42 476 for Ag⁺, Cu⁺⁺, Co⁺⁺, Zn⁺⁺, Bi⁺⁺⁺ and As⁺⁺⁺, respectively (NASDAQ, LME, 477 478 metalprices.com). The analyses suggest that considering the performance along with the cost, Ag⁺ and Cu⁺⁺ are the most effective catalysts among all catalysts used. In comparison, 479 catalysts such as Co⁺⁺, Bi⁺⁺⁺ and Hg⁺⁺ are not so effective and, in addition, few studies 480 481 using them have been reported, which precludes a definitive conclusion to be made. Between Cu^{++} and Ag^{+} , Ag^{+} is preferable, as it can be used for treating wide varieties of 482 483 minerals (Cu-bearing minerals, complex sulfide, spent batteries, Arsenopyrite etc.) as opposed to Cu⁺⁺, which is primarily found to catalyze only spent batteries in bioleaching. 484

Moreover, the efficiency of Ag^+ is better compared to Cu^{++} . An increase of more than 40% 485 486 in the bioleaching yield of a target metal is routinely evident in most of these studies when Ag⁺ is used as a catalyst. Relatively few amounts of Ag⁺ are required to induce the desired 487 catalytic effect. A concentration in the range of 0.5-1.0 g Ag⁺/kg of ore seems to be the 488 489 optimum concentration required to catalyze a wide variety of minerals and ore. Moreover, in the case of a commercially important mineral such as chalcopyrite, the cost of Ag⁺ 490 addition can be as low as 4.42% of the current cost of Cu (Table 4). Therefore, Ag⁺ seems 491 492 to be the best candidate among all catalysts, considering its cost of addition and 493 effectiveness. Future studies should explore its effectiveness for large-scale application. However, considering its effectiveness in terms of the cost of the addition of metal ions, the 494 495 process seems attractive for large-scale application, but to drive a satisfactory cost analysis of the overall process, the cost of safe disposal, recovery of metal ions from treated 496 497 residue/leachate, establishment of catalyst storage facilities, acclimatization of microorganisms to metal ions etc., will have to be added to the total cost. Annual 498 499 maintenance and management costs will also be incurred. Hence, this process seems less costly in terms of the cost of metal ion used, but could prove equally costly in terms of 500 overall cost of the process compared to traditional bioleaching. Unfortunately, inadequate 501 502 information is available in the literature on the economic aspects of metal ion-mediated 503 bioleaching processes, which limits the scope of a detailed cost-benefit analysis. Therefore, the economic and commercial viability of this process remains a knowledge gap which 504 requires urgent attention. 505

506

507 6.2 Toxicity to leaching microorganisms

One of the major challenges associated with metal ion-catalyzed bioleaching is the 508 509 possible toxic/inhibitory effects of these metal ions to bioleaching microorganisms. It has 510 been observed that most of the metal ions used as catalysts are potentially toxic to 511 mesophilic Acidithiobacilli (David et al., 2008; Mohseni et al., 2011). However, the 512 concentration of particular metal ions at which it becomes toxic/inhibitory varies for each 513 microorganism species and strains. The inhibitory concentrations of various catalytic ions 514 to bioleaching microorganisms are shown in Table 5. It can be seen that the iron oxidation rate of At. ferrooxidans was found to be inhibited at as low as 0.1 mg/L of Ag⁺/Hg⁺⁺. In the 515 case of Ag⁺, the inhibition has been explained through a mixed mechanism in which Ag⁺ 516 replaces Fe⁺⁺ in the active site of the oxidizing enzyme in the bacterial cell (De et al., 1996). 517 518 The accumulation of Ag on At. ferrooxidans cells has been confirmed during leaching of sulfide ore mineral (Fang et al., 2014). Besides Ag^+ and Hg^{++} , other commonly used metal 519 ions such as Cu⁺⁺ and Bi⁺⁺⁺ are also known to inhibit the bacterial iron-oxidation capacity. 520 However, their inhibitory effects are observed at comparatively higher concentrations 521 compared to Ag⁺. The special ability of Acidithiobacilli to tolerate relatively higher Cu 522 concentrations during bioleaching is due to having in its genome several genes that are 523 related to Cu homeostasis, such as those coding for CopA efflux ATPases, Cus efflux 524 systems and copper chaperones (Latorre et al., 2016). 525

526 Considering these issues, the identification of an appropriate concentration of these 527 metal ions is very important in the development of a successful bioleaching process. A low 528 concentration of these metal ions may not necessarily induce desirable changes, whereas a 529 higher concentration may impart toxicity to these microorganisms. A possible solution to 530 overcome this problem is to screen and employ those microbial species which have higher tolerance to these metal ions. Silver tolerant *Acidithiobacilli* sp. with the ability to grow in relatively higher concentration of Ag^+ , have been reported (Wu et al., 2007). Another method is to use the adapted culture which shows less inhibition in the presence of these metal ions. This can be achieved through repeated sub-culturing in progressively higher levels of these metal ions. Alternatively, the use of mutant strains produced through microwave induced mutation can yield *Acidithiobacillus sp*. with higher tolerance to metal ions (Wu et al., 2007).

538

539 6.3 Recovery of metal ions

As most of the metal ions used as catalysts during bioleaching are highly valuable metal commodities, their recovery at the end of bioleaching process is an important consideration for improving the cost-effectiveness of the process. There are different processes through which these metal species can be recovered, depending on the type of metal ions used and chemical state of that metal in the leaching system. The choice of treatment technology also depends upon whether a metal species is left in solid residue or is present as soluble species in solution.

The major process reported for recovery of different metal species from solid residue is leaching in which a lixiviant is added to the metal-containing materials which leach the metal. Thiourea leaching, leaching with thiosulfate, ozone leaching or bioleaching, are the major leaching techniques which have been successfully reported for leaching different metal ions $(Ag^+/Hg^{++}/Cu^{++}/Co^{+++})$ from a variety of primary minerals and spent materials. For example, Li et al. (2012) employed a thiourea leaching process to recover Ag from the printed circuit boards of waste mobile phones. A leaching solution having 24 g/L

thiourea and 0.6% of Fe^{3+} at room temperature was found to leach about 50% Ag within 2 554 555 hours. On the other hand, cementation, biosorption, ion exchange, electro-winning and solvent extractions are the major techniques which have been reported for the recovery of 556 these metals from a variety of liquid streams. Theoretically, at the end of bioleaching 557 558 processes, metal ions used as catalysts may be present either in solution or left in solid residue. However, most of the studies conducted, primarily with Ag⁺ as a catalyst, have 559 reported that Ag⁺ added during bioleaching remained in the solid residue (Romero et al., 560 561 2003).

Relatively few studies have been conducted for recovery of metal ions either from 562 563 the actual leached solid residue or leachate. In one of these studies, Palencia et al. (1998) 564 developed a silver catalyzed IBES process (indirect bioleaching with effect separation) and studied the recovery of the Ag^+ from the solid residue remaining after the Fe_2SO_4 leaching 565 566 of a chalcopyrite-sphalerite concentrate. Their study suggested that total recovery of the Ag⁺ added as a catalyst (2 mg/g concentrate) plus 93% of the Ag originally present in the 567 568 concentrate, can be obtained by leaching a leach residue (1 wt% of pulp density) with a 200 g/L NaCl-0.5 M H₂SO₄ at 90 °C and for 2 hours. Palencia et al. (1998) also found that to 569 achieve higher Ag recoveries, the previous removal of S^0 formed during bioleaching is 570 required. A similar acid-brine leaching process - the 'BRISA process' - has been 571 developed (Romero et al., 2003) in which the solid residue is leached in an acid-brine 572 medium with 200 g/L of NaCl and either HCl or H₂SO₄. Romero et al. (2003) postulated 573 that it is possible to obtain total recovery of the Ag^+ added as a catalyst plus 75% of the Ag 574 575 originally present in concentrate (44 mg/kg) by leaching the residue with a 200 g/L NaCl-0.5 M H₂SO₄ medium at 90 °C and 10% of pulp density in two stages of 2 hours each. Few 576

studies or processes have been developed to recover the catalytic metal ions from bioleached residue. The reported process, developed by Palencia et al. (1998) and Romero et al. (2003), also dealt primarily with recovery of Ag^+ from the chalcopyrite bioleaching. The studies of other valuable ions (Hg^{++} , Cu^{++} , Bi^{+++} etc.) used as catalysts to recover metals from other industrially important minerals and end-of-life industrial materials are absent. Therefore, in order to develop a successful commercial process, more in-depth studies using different metal ions have to be conducted under different process conditions.

584

585 6.4 Environmental safety

Large quantities of metal ions are going to be used as catalysts in order to develop 586 587 an industrial-scale bioleaching operation. Almost all metal ions employed as catalysts during bioleaching are considered toxic, and their safe transport, storage and disposal at the 588 end of the bioleaching process, is a critical challenge to the mining industry. Meanwhile, 589 there are increasingly stringent regulations against environmental pollution of different 590 591 toxic metal ions, and many countries have banned the permanent storage and management of surplus production of these metals. For example, the European Union (EU) implemented 592 an export ban on Hg in 2008 and enacted a further ban and policies for the permanent 593 storage and management of surplus Hg produced in EU countries in 2011 (The Zero 594 Mercury Working Group, 2007). Metallic ions such as Hg⁺⁺ and Ag⁺ are known for their 595 toxicity to marine, microbial, invertebrate and vertebrate communities (including humans) 596 including biomagnification via food chains, and have caused a number of public health 597 issues (Yang et al., 2012; O' Reily et al., 2016). The predominantly used Ag⁺ are 598 biologically active and promptly interact with proteins, free anions and receptors on 599

mammalian, eukaryotic cell membranes, assimilated in the human body and enters the 600 601 systemic dissemination as a protein complex (Lansdown, 2006; Katharina, 2011). Similarly, Hg is the most harmful metal known to humans (Tang et al., 2015) and divalent mercury 602 (Hg^{++}) is considered the most toxic form of Hg. Hg^{++} is highly reactive and binds to the 603 604 amino acid cysteine in proteins. Therefore, it has become a great concern to recover/remove 605 these species from the final effluent or residues and to ensure that they comply with the 606 stringent disposal standards. The US Environmental Protection Agency (USEPA) standards 607 for the disposal of hazardous wastes provide a disposal limit of Hg below 0.2 mg/L. 608 Similarly, the USEPA allowable disposal limit for safe disposal of Ag is 5 mg/L (USEPA, 609 2013). The EU (drinking water) regulation, 2014 (EU Regulations, 2014) also provides a 610 safe Hg limit of 0.001 mg/L for potable water. This regulation also prescribed a safe Cu limit in potable water of 2 mg/L. Therefore, it's a challenge to the mining industry to 611 612 formulate a robust policy in order to recover these metal ions from bioleached residue, 613 maintain adequate storage facilities and safety procedures, and achieve safe disposal of the treated residue. 614

615

616 **7.0 Future perspectives**

Although bioleaching is well understood, the commercial application of the process is still limited in the extractive metal industry, owing to its slow kinetics. It is foreseen that innovations such as the addition of metal ions as catalysts, may greatly improve the overall bioleaching efficiency and economics of large-scale industrial operations by promoting faster process kinetics and higher metal yields. Currently, studies on metal ion-catalyzed bioleaching are mostly conducted at laboratory-scale, and have been conducted with

primary minerals and ores. Studies conducted with secondary resources (such as end-of-life 623 624 products, industrial waste materials, mining wastes, spent batteries etc.) are limited and hence, research is needed to find out the process efficacy with these secondary resources. It 625 is important to note that most of these secondary resources contain a significant amount of 626 627 highly valuable metals and hence, their efficient processing through bioleaching will 628 achieve the goal of a "circular economy" (Ellen MacArthur Foundation, 2015). In terms of catalytic ions, Ag⁺ has proven to be the most exciting due to its positive effect on a wide 629 630 range of minerals (chalcopyrite, galena, complex sulfide, spent batteries etc.). There is evidence that Cu-bearing minerals and ore are currently the most amenable for Ag⁺ 631 catalyzed bioleaching. Considering the fact that a significant amount of Cu is already 632 produced industrially using bioleaching (Panda et al., 2015b), Ag⁺-catalyzed bioleaching 633 (with At. ferrooxidans) offers the greatest potential to extract Cu from low-grade minerals 634 and concentrates in a more efficient and cost-effective manner. Moreover, Ag⁺ can be 635 added to the leaching medium either as a finely divided metal or in the form of different 636 637 salts. However, types of salts used may affect the overall bioleaching process due to their differential solubility in the leaching media and their toxicity towards bioleaching 638 microorganisms (Mclerov et al., 1974). Silver salts such as AgCl, Ag₂SO₄ and AgNO₃ are 639 preferable as they are water soluble, although insoluble salts such as Ag₂S may also be used 640 but the quantity of Ag⁺ required to achieve the same yield will be relatively higher using 641 insoluble salts (Mcleroy et al., 1974). Among water soluble silver salts, AgNO₃ may be 642 preferable to AgCl and Ag₂SO₄ as the latter have a tendency to precipitate as 643 644 argentojarosite depending upon the media composition (Cordoba et al., 2009; Abdollahi et al., 2014). In terms of toxicity, Ag₂SO₄ has been found to be more toxic to bioleaching 645

microorganisms compared to AgCl. This was attributed to its relatively higher solubility in 646 647 aqueous solution, which leads to higher bioavailability of soluble silver ion to bioleaching 648 microorganisms (Sato et al., 2000). However, there are currently criteria for choosing these salts and the choice seems to be largely governed by its availability and cost. There is a 649 paucity of studies concerning the effect of different Ag⁺ salts on bioleaching process. 650 Studies with metal salts other than Ag⁺ (Hg⁺⁺, Bi⁺⁺⁺, Cu⁺⁺, Co⁺⁺) are also absent. Future 651 652 studies should explore this aspect. Moreover, studies on the combined use of metal ion 653 catalysts and non-metal catalysts or bi-metal ion catalyst are also absent. The simultaneous use of metal ion (such as Ag⁺) and non-metal catalyst (such as activated carbon) compared 654 to a single catalyst, may have the potential to induce greater catalysis owing to synergistic 655 656 effect. Similarly, the use of bi-metallic catalysts, in combination or in series, may also promote better bioleaching yields. Future studies should explore their possible efficacy and 657 reaction mechanisms during bioleaching. 658

It is also important to note that At. ferrooxidans is currently the most used 659 660 microorganism owing to its relatively higher tolerance to different metal ions generally present in bioleaching media. However, as each mineral and ore have different chemical 661 compositions, other leaching microorganisms such as thermophilic bacteria, archaea, and 662 fungi, under different process conditions, may also prove to be useful agents for gaining 663 664 better efficiency. Issues with the safe usage of many metal ions have been a concern with regulatory bodies and the public. Therefore, robust strategies for safe handling and disposal 665 need to be identified in accordance with the prevailing local and international laws. 666 Following this review, it is recommended that the focus of the future work should be on (1) 667 investigating the efficiency of processes with different secondary resources (2) evaluating 668

the use of different types of microorganisms (3) examining the process efficiency in larger
scale reactors, and (4) evaluating different cost-effective processes to recover metal ions
from treated bioleached residue and metal laden liquor.

672

673 **8. Conclusions**

The analysis suggests that addition of different metal ions $(Ag^+, Hg^{++}, Cu^{++}, etc.)$ 674 increases bioleaching efficiency remarkably. Although most of the metal ion-catalyzed 675 676 bioleaching studies have obtained promising results, techno-economic challenges still exist 677 and efforts are needed to overcome these challenges in order to make the technology 678 feasible for large-scale application. Moreover, the feasibility and sustainability of these 679 processes have not been demonstrated on a large-scale and there remains a need to scale-up investigations, which should include estimations of cost and environmental impact. This is 680 681 a critical aspect in the development of future industrial metal ion-catalyzed biohydrometallurgical processes. 682

683

684 Acknowledgements

This work was financially supported by a grant from Irish Research Council(GOIPD/2015/589).

- 687
- 688

689

690

692 **References**

- 1. Abdollahi, H., Noaparast, M., Shafaei, S.Z., Manafi, Z., Munoz, J.A., Tuovinen, O.H.,
- 694 2015. Silver-catalyzed bioleaching of copper, molybdenum and rhenium from a 695 chalcopyrite molybdenite concentrate. Int. Biodeter. Biodegr. 104, 194-200.
- 696 2. Abdollahi, H., Shafaei, S.Z., Noaparast, M., Manafi, Z., Niemela, S.I., Tuovinen, O.H.,
- 697 2014. Mesophilic and thermophilic bioleaching of copper from a chalcopyrite-698 containing molybdenite concentrate. Int. J. Mine. Proces. 128, 25–32.
- Ballaster, A., Gonzales, F., Blazguez, M.L., Mier, J.L., 1990. The influence of various
 ions in the bioleaching of metal sulphides. Hydrometallurgy 23, 221-235.
- 4. Ballester, A., Gonzalez, F., Blaz quez, M.L., Gomez, C., Mier, J.L., 1992. The use of
 catalytic ions in bioleaching. Hydrometallurgy 29, 145-160.
- 5. Brierley, C.L. 2008. How will biomining be applied in future? Trans. Nonfer. Met. Soc.
 China, 18(6), 1302-1310.
- 6. Cancho, L., Blazquez, M.L., Ballester, A., Gonzalez, F., Munoz, J.A., 2007. Bioleaching
 of a chalcopyrite concentrate with moderate thermophilic microorganisms in a
 continuous reactor system. Hydrometallurgy 87, 100-111.
- 708 7. Cerda, P.A.M., Ohlbaum, R.B.B., 2008. Process to increase the bioleaching speed of
 709 ores or concentrates of sulfide metal species, by means of continuous inoculation with
 710 leaching solution that contains isolated microorganisms, with or without presence of
 711 native microorganisms. US Patent No. US20080127779A1.
- 8. Chang, Li. L., Jin, L.X., Zhen, Y.N., Yi, Y., Chen, Y.M., 2012. Effect of sodium
 chloride on sulfur speciation of chalcopyrite bioleached by the extreme thermophile *Acidianus manzaensis*. Bioresour. Technol. 110, 462-470.
- 715 9. Chen, S., Qin, W.Q., Qiu, G.Z., 2008. Effect of Cu^{2+} ions on bioleaching of marmatite.
- 716 Tran. Nonferrous Met. Soc. China. 18, 1518-1522.
- 717 10. Chen, S.Y., Lin, J.G., 2009. Enhancement of metal bioleaching from contaminated
 718 sediment using silver ion. J. Haz. Mate. 161, 893-899.
- 11. Clarke, M.E., Batty, J.D., van Buuren, C.B., Dew, D.W., Eamon, M.A., 2006.
 Biotechnology in mineral processing: Technological breakthroughs creating value.
 Hydrometallurgy 83, 3-9.

- 12. Cordoba, E.M., Munoz, J.A., Blazquez, M.L., Gonzalez, F., Ballester, A., 2009.
 Comparative kinetic study of the silver-catalyzed chalcopyrite leaching at 35 and 68 °C.
 Int. J. Miner. Process. 92, 137–143.
- 13. Das, A., Modak, J.M., Natrajan, K.A., 1997. Studies on multi-metal ion tolerance of
 thiobacillus ferrooxidans. Mine. Engg. 10, 743-749.
- 14. Das, A.P., Sukla, L.B., Pradhan, N., Nayak, S., 2011. Manganese biomining. Bioresour.
 Technol. 102, 7381-7387.
- 15. David, D.J., Pradhan, D., Das, T., 2008. Evaluation of iron oxidation rate of *Acidithiobacillus ferrooxidans* in presence of heavy metal ions. Min. Proces. Extractive
 Metall. 117, 56-61.
- 16. De, G.C., Oliver, D.J., Pesic, B.M., 1996. Effect of silver on the ferrous iron oxidizing
 ability of *Thiobacillus ferrooxidans*. Hydrometallurgy 41, 211–229.
- 17. Deveci, H., Akcil, A., Alp, I., 2004. Bioleaching of complex zinc sulphides using
 mesophilic and thermophilic bacteria: comparative importance of pH and iron.
 Hydrometallurgy 73, 293–303.
- 18. Dew, D.W., Lawson, E.N., Broadhurst, J.L., 1997. In: D. E. Rawlings (Ed.), Biomining:
 Theory, Microbes and Industrial Processes, Springer-Verlag, Berlin, pp. 45–80.
- 19. Diao, M., Taran, E., Mahler, S., Nguyen, A.V., 2014. A concise review of nanoscopic
 aspects of bioleaching bacteria–mineral interactions. Adv. Colloid. Interface Sci. 212,
- 741 45–63.
- 20. Donati, E.R., Castro, C., Urbieta, M.S., 2016. Thermophilic microorganisms in
 biomining. World J. Microbial. Technol. 32, 1-8.
- 21. Dong, Y.B., Lin, H., Zhou, S., Xu, X., Zhang, Y., 2013. Effects of quartz addition on
 chalcopyrite bioleaching in shaking flasks. Mine. Eng. 46-47, 177-179.
- 22. Du Plessis, C.A., Batty, J.D., Dew, D.W., 2007. Commercial applications of
 thermophilie bioleaching. In: D.E. Rawlings, D.B. Johnson (Eds.). 2007, Biomining,
 Springer, pp. 57-80.
- 23. Ellen MacArthur Foundation. 2015. Growth within: a circular economy vision for a
 competitive Europe. Report by the Ellen MacArthur Foundation, the McKinsey Centre

- for Business and Environment and the Stiftungsfonds für Umweltökonomie undNachhaltigkeit (SUN), June 2015.
- 24. Escudero, M.E., Gonzilez, F., Blizquez, M.L, Ballester, A., Gomez, C., 1993. The
 catalytic effect of some cations on the biological leaching of a Spanish complex sulphide.
- 755 Hydrometallurgy 34, 151-169.
- 25. EU-European Union (Drinking Water) Regulations., 2014, S.I. No. 122 of 2014.
- 757 26. Fang, F., Zhong, H., Jiang, F., Zhan, X., 2014. Catalytic effect of silver on bioleaching
 758 of arsenopyrite. Int. J. Chem. Eng. Appl. 5, 474-478.
- 759 27. Feng, S., Yang, H., Wang, Wu., 2015. Improved chalcopyrite bioleaching by *Acidithiobacillus sp.* via direct step-wise regulation of microbial community structure.
 761 Bioresour. Technol. 192, 75-82.
- 762 28. Feng, S., Yang, H., Wang, Wu., 2016. Insights to the effects of free cells on community
 763 structure of attached cells and chalcopyrite bioleaching during different stages.
 764 Bioresour. Technol. 200, 186-193.
- 765 29. Gomez, E., Ballester, A., Blazquez, M.L., Gonzalez, F., 1999. Silver-catalysed
 766 bioleaching of a chalcopyrite concentrate with mixed cultures of moderately
 767 thermophilic microorganisms. Hydrometallurgy 51, 37-46.
- 30. Gomez, E., Roman, E., Blazquez, M.L., Ballester, A., 1997. SEM and AES studies of
 chalcopyrite bioleaching in the presence of catalytic ions. Mine. Eng. 10, 823-835.
- 31. Grogan, D.W., 1989. Phenotypic characterization of the archaebacterial genus *sulfolobus*:
 comparison of five wild type strains. J. Bacteriol. 171, 6710-6719.
- 32. Guo, P., Zhang, G., Cao, J., Li, Y., Fang, Z., Yang, C., 2011. Catalytic effect of Ag⁺ and
 Cu²⁺ on leaching realgar (As₂S₂). Hydrometallurgy 106, 99-103.
- 33. Habashi, F., 1983. Dissolution of minerals and hydrometallurgical processes. The Sc.
 Nat. 70, 403-411.
- 34. He, Z., Gao, F., Zhong, H., Hu, Y., 2009. Effect of L-cysteine on Ni-Cu sulphide and
 marmatite bioleaching by *Acidithiobacillus caldus*. Bioresour. Technol. 100, 1383-1387.
- 35. Hu, Y., Qiu, G., Wang, J., Wang, D., 2002. The effect of silver-bearing catalysts on
 bioleaching of chalcopyrite. Hydrometallurgy 64, 81–88.

- 36. Hua, Y., Qiu, G., Wang, J., Wang, D., 2002. The effect of silver-bearing catalysts on
 bioleaching of chalcopyrite. Hydrometallurgy 64, 61-88.
- 37. Johnson, D. B., Ghauri, M. A., Said, M. F., 1992. Isolation and characterization of an
 acidophilic, heterotrophic bacterium capable of oxidizing ferrous iron. Appl. Environ.
 Microbiol. 58, 1423–1428.
- 785 38. Karimi, G.R., Rowsona, N.A., Hewitt, C.J., 2010. Bioleaching of copper via iron
- oxidation from chalcopyrite at elevated temperatures. Food Bioprod. Process. 88, 21–25.
- 39. Karthikeyan, O.P., Rajasekar, A., Balasubramanian, R., 2015. Bio-oxidation and biocyanidation of refractory mineral ores for gold extraction: A review. Crit. Rev. Environ.
- 789 Sci. Technol. 45, 1611-1643.
- 40. Katharina, M.F., 2011. Give silver a shine. Nat. Chem. 3, 178.
- 41. Krebs, W., Brombacher, C., Bosshard, P.P., Bachofen, R., Brandel, H., 1997. Microbial
 recovery of metals from solids. FEMS Microbiol. Rev. 20, 605-617.
- 42. Kuwazawa, T., Miura, A., 2012. Method of leaching copper sulphide. US Patent no. US
 8,277,538 B2.
- 43. Lansdown, A.B., 2006. Silver in health care: antimicrobial effects and safety in use.
 Current Prob. Derm. 33, 17–34.
- 44. Latorre, M., Ehrenfeld, N., Cortés, M.P., Travisany, D., Budinich, M., Aravena, A.,
- González, M., Bobadilla-Fazzini, R.A., Parada, P., Maass, A., 2016. Global
 transcriptional responses of *Acidithiobacillus ferrooxidans* Wenelen under different
 sulfide minerals. Bioresour. Technol. 200, 29-34.
- 45. Lee, J, Acar, S., Doerr, D.L., Brierley, J.A., 2015. Comparative bioleaching and
 mineralogy of composited sulfide ores containing enargite, covellite and chalcocite by
 mesophilic and thermophilic microorganisms. Hydrometallurgy 105, 213-221.
- 46. Li, J.Y. Xu, X. L., Liu, W.Q., 2012. Thiourea leaching gold and silver from the printed
 circuit boards of waste mobile phones. Waste Manage. 32, 1209-1212.
- 47. Li, S., Zhong, H., Hu, Y., Zhao, J., He, Z., Gu, G., 2014. Bioleaching of a low-grade
 nickel-copper sulfide by mixture of four thermophiles. Bioresour. Technol. 153, 300–
 306.

- 48. Liu, W., Yang, H.Y., Song, Y., Tong, L.L., 2015. Catalytic effects of activated carbon
 and surfactants on bioleaching of cobalt ore. Hydrometallurgy 152, 69–75.
- 811 49. LME, <u>www.lme.com</u> (accessed on 30th June 2016).
- 812 50. Ma, L., Wang, X., Feng, X., Liang, Y., Xiao, Y., Hao, X., Yin, H., Liu, H., Liu, X., 2017.
- 813 Coculture microorganisms with different initial proportions reveal the mechanism of
- chalcopyrite bioleaching coupling with microbial community succession, Bioresour.
- 815 Technol. 223, 121-130.
- 51. McElroy, R.O. and Duncan, D.W., 1974. Copper extraction by rapid bacteriological
 process. US Patent No. 3,856,913.
- 818 52. Metalprices.com, https://www.metalprices.com/metal/arsenic/arsenic-99-usa (accessed
 819 on 12th December 2016).
- 53. Mier, J.L., Ballester, A., Gonzalez, F., Blazquez, M.L., Gomez, E., 1996. The influence
 of metallic ions on the activity of *sulfolobus BC*. J. Chem. Tech. Biotechnol. 65, 272280.
- 54. Mier, J.L., Gomez, C., Ballester, A., Blazquez, M.L., Gonzalez, F., 1994. Effect of silver
 and bismuth on bioleaching of copper sulphide concentrates with thermophilic
 microorganisms. In: Hydrometallurgy 94 (International Symposium), July 11-15,
 Cambridge, England.
- 55. Mohseni, S., Marzban, A.R., Sepehr, S., Hosseinkhani S, Karkhaneh M, Azimi A., 2011.
 Investigation of some heavy metals toxicity for indigenous *Acidithiobacillus ferrooxidans* isolated from Sarcheshmeh copper mine. Jundishapur J. Microbiol. 4, 159166.
- 56. Mulak, W., 1987. The catalytic action of cupric and ferric ions in nitric acid leaching of
 Ni₃S₂. Hydrometallurgy 17, 201–214.
- 57. Munoz, J.A., Dreisinger, D.B., Cooper, W.C., Young, S.K., 2007. Silver catalyzed
 bioleaching of low-grade copper ores. Part III: Column reactors. Hydrometallurgy 88,
 35-51.
- 58. Nakazawa, H., Fujisawa, H., Sato, H., 1998. Effect of activated carbon on the
 bioleaching of chalcopyrite Concentrate. Int. J. Mine. Process. 55, 87–94.
- 838 59. NASDAQ, http://www.nasdaq.com/markets/silver.aspx (accessed on 30th June 2016).

- 60. Nemati, M., Lowenadler, J., Harrison, S., 2000. Particle size effects in bioleaching of
 pyrite by acidophilic thermophile *Sulfolobus metallicus* (BC). Appl. Microbiol.
 Biotechnol. 53, 173-179.
- 842 61. Nguyen, V.K., Lee, J.U., 2014.Catalytic effect of activated charcoal on microbial
 843 extraction of arsenic and heavy metals from mine tailings. Geosci. J. 18, 355-363.
- 62. Niu, Z., Huang, Q., Wang, J., Yang, Y., Xin, B., Chen, S., 2015. Metallic ions catalysis
- for improving bioleaching yield of Zn and Mn from spent Zn-Mn batteries at high pulp
 density of 10%. J. Haz. Mate. 298, 170–177.
- 63. O'Reilly, S.B., Schierl, R., Nowak, D., Siebert, U., William, J.F., Owi, F.T., Ismawati Ir,
 Y., 2016. A preliminary study on health effects in villagers exposed to mercury in a
 small-scale artisanal gold mining area in Indonesia. Environ. Res. 149, 274–281.
- 64. Palencia, I., Romero, R., Carranza, F., 1998. Silver catalyzed IBES process: Application
 to a Spanish copper–zinc sulphide concentrate. Part 2. Biooxidation of the ferrous iron
 and catalyst recovery. Hydrometallurgy 48, 101–112.
- 65. Panda, S., Biswal, A., Mishra, S., Panda, P.K., Pradhan, N., Mohapatra, U., Sukla, L.B.,
 Mishra, B.K., Akcil, A., 2015a. Reductive dissolution by waste newspaper for enhanced
 meso-acidophilic bioleaching of copper from low grade chalcopyrite: A new concept of
 biohydrometallurgy. Hydrometallurgy 153, 98-105.
- 857 66. Panda, S., Akcil, A., Pradhan, N., Deveci, H., 2015b. Current scenario of chalcopyrite
 858 bioleaching: A review on the recent advances to its heap-leach technology. Bioresour.
 859 Technol. 196, 694–706.
- 860 67. Pradhan, N., Nathsarma, K.C., Rao, K.S., Sukla, L.B., Mishra, B.K., 2008. Heap
 861 bioleaching of chalcopyrite: A review. Mine. Engg. 21, 355–365.
- 862 68. Romano, P., Blázquez, M.L., Alguacil, F.J., Munoz, J.A., Ballester, A., González, F.,
- 2001. Comparative study on the selective chalcopyrite bioleaching of a molybdenite
 concentrates with mesophilic and thermophilic bacteria. FEMS Microbiol. Lett. 196, 71–
 75.
- 866 69. Romero, R., Mazuelos, A., Palencia, I., Carranza, F., 2003. Copper recovery from
 867 chalcopyrite concentrates by the BRISA process. Hydrometallurgy 70, 205–215.

- 868 70. Sand, W., Gehrke, T., Hallmann, R., Rhode, K., Sobotke, B., Wentzien, S., 1993. In:
 869 Torma AE, Wey JE, Lakshmanan, (Eds.), In-situ bioleaching of metal sulfides: The
- 870 importance of *Leptospirillum ferrooxidans*. Biohydrometallurgical Technologies. TMS
- 871 Press, Warrendale, Pennsylvaia, pp. 15-27.
- 872 71. Sato, H., Nakazawa, H., Kudo, Y., 2000. Effect of silver chloride on the bioleaching of
 873 chalcopyrite concentrate. Int. J. Mine. Process. 59, 17-24.
- 874 72. Schippers, A., Hedrich, S., Vasters, J., Drobe, M., Sand, W., Willscher, S., 2014.
 875 Biomining: metal recovery from ores with microorganisms. Adv. Biochem. Eng.
 876 Biotechnol. 141, 1–47.
- 877 73. Scott, T., Dyson, N., 1968. The catalyzed oxidation of zinc sulfide under acid pressure
 878 leaching conditions. Trans. Metal. Soc. AIME. 242, 1815–1821.
- 74. Tang, J., Lv, J., Gong, Y., Huang, Y., 2015. Preparation and characterization of a novel
 graphene/biochar composite for aqueous phenanthrene and mercury removal. Bioresour.
 Technol. 196, 355-363.
- 75. The Zero Mercury Working Group/Health and Environment Alliance/Health Care
 Without Harm Europe/European Environmental Bureau, EU Mercury Export Ban
 Advances, Safer Liquid Mercury Storage Now Being Considered, 2007.
- 76. Tuovinen, O.H., Puhakka, J., Hiltunen, P. and Dolan, K.M., 1985. Silver toxicity to
 ferrous iron and pyrite oxidation and its alleviation by yeast extract in cultures of *Thiobacillus ferrooxidans*. Biotechnol. Lett. 7, 389-394.
- 888 77. USEPA (United State Environmental Protection Agency). 2013. Land Disposal
 889 Restrictions, 40 CFR 268.
- 78. Vainshtein, M., Abashina, T., Bykov, A., Repina, A., Kaparullina., 2015. Formate
 supplementation can increase nickel recovery by *Halothiobacillus halophilus*. World J
 Microbiol. Biotechnol. 31, 535-537.
- 893 79. Vakilchap, F., Mousavi, S.M., Shojaosadati, S.A., 2016. Role of Aspergillus niger in
- recovery enhancement of valuable metals from produced red mud in Bayer process.
- Bioresour. Technol. 218, 991-998.
- 896 80. Walting, H., 2015. Review of biohydrometallurgical metal extraction from polymetallic
 897 mineral resources. Minerals. 5, 1-60.

- 898 81. Wu, X.L., Qiu, G.Z., Gao, J., Ding, J.N., Kang, J., Liu, X.X., 2007. Mutagenic breeding
- 899 of silver-resistant *Acidithiobacillus ferrooxidans* and exploration of resistant mechanism.
- 900 Trans. Nonferrous Met. Soc. China. 17, 412-417.
- 901 82. Xu, A.L., Xia, J.L., Zhang, S., Yang, Y., Nie, Z.Y., Qiu, G.Z., 2010. Bioleaching of
- 902 chalcopyrite by UV-induced mutagenized *Acidiphilium cryptum* and *Acidithiobacillus*
- 903 *ferrooxidans*. Trans. Nonferrous Met. Soc. China. 20, 315-321.
- 83. Yag, H.M., Liu, H., Wu, H. and Wang, M., 2012. Photochemical removal of gaseous
 elemental mercury in a dielectric barrier discharge plasma reactor. Plasma Chem.
 Plasma Process. 32, 969–977.
- 84. Zeng, G., Deng, X., Luo, S., Luo, X., Zou, X., 2012. A copper-catalyzed bioleaching
 process for enhancement of cobalt dissolution from spent lithium-ion batteries. J. Haz.
 Mate. 199-200, 164–169.
- 85. Zeng, G., Luo, S., Deng, X., Li, L., Au, C., 2013. Influence of silver ions on bioleaching
 of cobalt from spent lithium batteries. Mine. Eng. 49, 40–44.
- 86. Zeng, J., Gou, M., Tang, Y.Q., Li, G.Y., Sun, Z.Y., Kida, K., 2016. Effective
 bioleaching of chromium in tannery sludge with an enriched sulfur-oxidizing bacterial
 community. Bioresour. Technol. 218, 859-866.
- 87. Zhang, R., Wei, D., Shen, Y., Liu, W., Lu, T., Han, C., 2016. Catalytic effect of
 polyethylene glycol on sulfur oxidation in chalcopyrite bioleaching by *Acidithiobacillus*
- 917 *ferrooxidans*. Mine. Eng. 95, 74-78.
- 88. Zhou, S., Gan, M., Zhu, J., Li, Q., Jie, S., Yang, B., Liu, X., 2015. Catalytic effect of
 light illumination on bioleaching of chalcopyrite. Bioresour. Technol. 182, 345-352.
- 920
- 921
- 922
- 923
- 924

- 926
- 927

928	Figure	captions
-----	--------	----------

929	Fig. A1: XRD of the bioleached residue confirming the presence of Ag_2S (Hu et al., 2002)
930	Fig. A2: SEM micrographs (A and B) and EDAX mapping (C and D) showing Ag
931	precipitation on bioleached residues of molybdenite concentrate (Abdollahi et al.,
932	2014.
933	Fig. A3: SEM images of residues leached (A) without Cu^{++} (B) in the presence of Cu^{++}
934	(Zeng et al., 2012)
935	
936	List of Tables
937	Table 1: Different catalytic agents reported for promoting bioleaching efficiency.
938	Table 2 : Metal ions used as catalyst in different bioleaching studies.
939	Table 3 : Operating conditions reported for metal ion catalyzed bioleaching.
940	Table 4: Comparative cost and performance of the metal ion catalysts during bioleaching.
941	Table 5: Inhibitory concentration of catalytic ions to different leaching microorganisms.
942	
943	
944	
945	
946	
947	
948	
949	
950	

Catalytic Agents	Microorganisms	Conc. of the catalyst (g catalyst /kg of ore)	Reaction time (Day)	Yield (%)	Ore/ Minerals	References
Activated carbon	At. ferrooxidans	0-625	19	42 (Cu)	Chalcopyrite concentrate	Nakazawa et al., 1998
Activated carbon	At. ferrooxidans, At.thiooxidans	12	25	79 (Cu)	Copper sulfide ores	Zhang et al., 2007
Activated charcoal	At. thiooxidans, At. ferrooxidans	80	16.7	86 (Cu)	Mine tailings	Nguyen and Lee, 2014
Activated carbon	At. ferrooxidans, At. thiooxidans, L. ferrooxidans	10	18	90.8 (Co)	Cobalt ore	Liu et al., 2015
Ag^+	At. ferrooxidans	6.7	30	93 (Cu)	Chalcopyrite -molybdnite	Abdollahi et al., 2015
Cu ⁺⁺	At. thiooxidans, L. ferriphilum	8.0	13	62.5 (Zn), 62.4 (Mn)	Spent Zn- Mn batteries	Niu, et al., 2015
Cysteine	A. caldus	10	19	25 (Cu)	Ni-Cu sulfide	He et al., 2009
Cysteine	A. brierleyi , A. manzaensis, M. sedula JCM, S. metallicus JCM	10	16	81.4 (Cu), 83.7 (Ni)	Ni-Cu sulfide	Li et al., 2014
Quartz	<i>At. ferrooxidans</i> LD-1	2500	32	53.5 (Cu)	Chaclopyrite	Dong et al., 2013
Sodium chloride	A. manzaensis YN-25	66	10	82 (Cu)	Chalcopyrite	Chang et al., 2012
Waste newspaper	At. ferrooxidans	20	6	99.13 (Cu)	Chalcopyrite	Panda et al., 2015a
Polyethyle ne glycol	At. ferrooxidans	09	21	12.5 (Cu)	Chalcopyrite	Zhang et al., 2016

Table 1: Different catalytic agents reported for enhancing bioleaching efficiency

Metal ion	Salt	Conc. (g catalyst	Reaction time	Bioleaching yields (%)		Ore	References	
		/kg of ore)	(Day)	Cu	Zn	Co		
				Cu				
Ag^+	Ag_2SO_4	2.0	16.7	90	84		Complex sulfide	Ballester et al., 1990
Ag^+	Ag_2SO_4	0.2	21	60			Complex sulfide	Ballester et al., 1992
Ag^{+}	AgNO ₃	6.7	30	93			Chalcopyrite- molybdnite	Abdollahi et al., 2015
Ag^{+}	Ag ₂ SO4	1.0	180	44.4			Low grade	Munoz et al., 2007
Ag^+	AgNO ₃	2	7			98.4	Spent lithium	Zeng et al., 2013
Hg ⁺⁺	HgSO ₄	2	16.7	32.5	59		Sphalerite/ complex sulfide	Ballester et al., 1990
Hg^{++}	HgSO ₄	0.2	21	23			Complex sulfide	Ballester et al., 1992
Hg^{++}	HgSO ₄	0.8	22	58	85		Complex sulfide	Escudro et al., 1993
Bi ⁺⁺⁺	Bi(NO ₃) ₃	2.0	16.7	28	78.4		Sphalerite/ complex sulfide	Ballester et al., 1990
Bi ⁺⁺⁺	Bi(NO ₃) ₃ . 5H ₂ O	0.2	21	18			Complex sulfide	Ballester et al., 1992
Cu ⁺⁺	CuSO₄∙ 5H₂O	75	10			99.9	Spent lithium ion batteries	Zeng et al., 2012
Cu ⁺⁺	CuSO ₄ ·	8.0	13		62.5		Spent Zn-Mn batteries	Niu et al., 2015
Co ⁺⁺	5H ₂ O CoSO ₄ . 7H ₂ O	2	16.7	55	90.5		Complex sulfide	Ballester et al., 1990

 Table 2: Metal ions used as catalyst in different bioleaching studies

Metal	Salt	Conc. of	Medium	Pulp	Temp.	Materials	References
ions		the	pН	density	(°C)	bioleached	
		catalyst		(%)			
		(g/kg)					
Agʻ	AgNO ₃	0-1	2.5	10.67	35	Chalcopyrite	Mcelroy and
. +		-	•	_		~	Duncan, 1974
Ag	Ag_2SO_4	2	2.0	5	35	Sphalerite/	Ballester et al.,
. +						Complex sulfide	1990
Agʻ	AgNO ₃	0.2-20	2.0	1	35	Spent lithium	Zeng et al.,
l						batteries	2013
Ag^{-}	-	4	2.0	0.5	35	Realgar	Guo et al.,
							2011
Ag^+	-	0.1	2.0	2	30	Arsenopyrite	Fang et al.,
							2014
Ag^+	AgNO ₃	0-33.3	1.6	3.0	32	Chalcopyrite-	Abdolllahi et
						Molybednite	al., 2015
Hg^{++}	HgSO ₄	2	2.0	5	35	Sphalerite and	Ballester et al.,
						Complex sulfide	1990
Hg^{++}	HgSO ₄	0.8	2.0	5	35	Complex sulfide	Escudro et al.,
							1993
Bi ⁺⁺⁺	Bi(NO ₃)	2	2.0	5	35	Sphalerite and	Ballester et al.,
	3					Complex sulfide	1990
Bi ⁺⁺⁺	Bi(NO ₃)	10		1	35	Copper-sulfide	Mier et al.,
	3.					concentrate	1994
	$5H_2O$						
Cu^{++}	$CuSO_4\cdot$	0.750	2.9	1	35	Spent lithium-	Zeng et al.,
	$5H_2O$					ion batteries	2013
Cu^{++}	CuSO₄∙	8	1.0	10	35	Spent Zn-Mn	Niu et al.,
	511.0					batteries	2015
C_{c}^{++}	$3\Pi_2 O$	0.0	2.0	5	25	Complex sulfile	Equidare et al
CO	CUSU4. 711 O	0.8	2.0	3	33	Complex sunde	
	/п20						1773

Table 3: Operating conditions reported for metal ion catalyzed bioleaching.

Catalyst	Mineral/ore	Amount of catalyst (g catalyst /kg of ore	Increment in bioleaching yield after catalyst addition (%)	Cost of catalyst addition to ore (A) in US \$	Cost (%) wrt. to the price of target metal = (A/Current price of target	Reference
				/kg of ore	metal in US \$	
Ag^+	Chalcopyrite	0.5	38 (Cu)	0.34	<u>kg</u>)*100 6.9	Mcelroy and Duncan, 1974
Ag^+	Complex sulphide	2.0	63.5 (Cu)	1.34	27.6	Ballester et al., 1990
Ag^+	Chalcopyrite	0.32	50 (Cu)	0.21	4.42	Hu et al., 2002
Ag^+	Chalcopyrite	1.0	44.4 (Cu)	0.67	13.8	Munoz et al., 2007
Ag^+	Spent lithium ion batteries	2.0	55.3 (Co)	1.34	5.2	Zeng et al., 2013
Ag^+	Chalcopyrite- molybdnite	6.7	40 (Cu)	4.49	92.5	Abdollahi et al., 2015
Bi ⁺⁺⁺	Sphalerite	2.0	28.4 (Zn)	0.018	0.8	Ballester et al., 1990
Bi ⁺⁺⁺	Chalcopyrite	10	10 (Cu)	0.09	1.9	Ballester et al., 1992
Cu ⁺⁺	Marmatite	5.0	8 (Zn)	0.024	1.1	Chen et al., 2008
Cu ⁺⁺	Spent lithium ion batteries	75	56 (Co)	0.36	1.4	Zeng et al., 2012
Cu ⁺⁺	Spent zinc batteries	8.0	15 (Zn)	0.04	1.7	Niu, et al., 2015
Co ⁺⁺	Sphalerite	2.0	18.4 (Zn)	0.052	2.3	Ballester et al., 1990
Co ⁺⁺	Complex sulfide	2.0	30 (Cu)	0.052	1.1	Escudero et al., 1993
Co++	Complex sulfide	2.0	27 (Zn)	0.052	2.3	Escudero et al., 1993

Table 4: Comparative cost and performance of the metal ion catalysts during bioleaching

Catalytic	Salt	Toxic	Affecting	Reference
ion		concentration	Microorganisms	
		(mg/L)		
Ag^+	AgNO ₃	0.10	At. ferrooxidans	Tuovinen et al., 1985
Ag^+	Ag_2SO_4	0.2	L. ferrooxidans	Johnson et al., 1992
Ag^{+}	-	0.9	S. solfataricus	Grogan, 1989
Ag^{+}	Ag_2SO_4	20	S. Bc	Munoz et al., 1998
Hg^{++}	-	0.1	At. ferrooxidans	David et al., 2008
Hg^{++}	-	0.4	S. solfataricus	Grogan, 1989
Bi	Bi(NO) ₃ ⁻⁵ H ₂ O	9000	S. BC	Mier at al., 1996
Cu^{++}	CuSO ₄ ·5H ₂ O	5000	At. thiooxidans,	Niu et al., 2015
			L.ferriphilum	
Cu^{++}	CuSO ₄	10,000	At. ferrooxidans	Das et al., 1997
Cu^{++}	CuSO ₄	3177	Acidiphilum cryptum	Johnson et al., 1992
Co	CoSO4 ⁻⁷ H ₂ O	5300	S. BC	Mier at al., 1996
Co	CoSO4 ⁻⁷ H ₂ O	2000	L. ferrooxidans	Sand et al., 1993.

Table 5: Inhibitory concentration of catalytic ions to different bioleaching microorganisms