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Categorization of tars from fast pyrolysis of pure lignocellulosic compounds at high temperature

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Abstract

This study presents how the yields of different tar compounds from pure lignocellulosic compound respond to the change in temperature and residence time. Experiments were carried out with a drop tube furnace in the temperature range from 800 to 1250°C. The tar composition was characterized by gas chromatography with a flame ionization detector and mass spectrometry using a dual detector system. Longer residence time and higher heat treatment temperatures increased the soot formation and decreased the tar yields. Soot yields from lignin samples were greater than soot yields from holocellulose pyrolysis. The dominating products in tars from pyrolysis of all lignocellulosic compounds were benzene and toluene. Cellulose and hemicellulose pyrolysis produced greater amount of oxygenates in tars, whereas lignin tar was rich in phenols, polycyclic hydrocarbons and naphthalenes. Simultaneous reduction of tar and soot was achieved by impregnation of lignin

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from wheat straw with alkali metals. The OPLS-DA model can accurately explain the differences in tar composition based on the experimental mass spectrometry data.

Keywords: fast pyrolysis, lignocellulosic compounds, potassium, tar, principal component analysis

1. Introduction

Lignocellulosic biomass has a potential to replace fossil fuels in the production of liquid hydrocarbons and chemicals [1]. Biomass gasification offers high conversion efficiency and the possibility to handle different lignocelluloses [2]. Owing to the high volatile content in biomass, a major challenge in biomass gasification is usually tar content in syngas, which decreases the cold gas efficiency and requires cost-intensive cleaning systems. Entrained flow gasification (EFG) process generates little or no tar due to high operating temperatures although it produces considerable amount of soot.

In order to reduce the soot yield from EFG process it is necessary to understand how tar composition and yield are correlated with fuel composition and operating conditions since soot is formed as a result of the reactions between PAHs. Tars are defined as a complex mixture of hydrocarbons with single or multiple rings [3]. Primary tars are generated during devolatilization at the temperatures of 400-500°C, followed by a series of cracking reactions [4]. Primary tars are predominantly oxygenated organic compounds similar to the original structures of lignocellulosic compounds. Primary tars from cellulose and hemicellulose are levoglucosan, xylose and non-aromatic oxygenated compounds (ketones, aldehydes, and alcohols) while primary tars from lignin

20 contain mainly phenolic compounds [5–7]. Secondary reactions take place
21 both heterogeneously and homogeneously, and include intra-particle crack-
22 ing of tar on the char surfaces [4, 8]. The secondary tars are mainly composed
23 of phenols and olefins [9]. The tertiary products are generated above 700°C
24 and subdivided into alkyl aromatics and larger polycyclic aromatic hydrocar-
25 bons e.g. benzene, toluene, naphthalene, pyrene, etc. [10–12]. The molecular
26 structure of tertiary tars cannot be found in natural biomass and can emerge
27 from small molecule fragments as allyl-, aryl-, and alkyl radicals, which re-
28 sult from cycloaddition reactions according to Diels-Alder and followed by
29 aromatization due to dehydrogenation respectively dehydration [9]. As well
30 known from a number of studies on the combustion of gaseous and liquid fuel
31 combustion [13], polycyclic aromatic hydrocarbons (PAH) are thought to be
32 the main precursors of soot [14, 15]. Some studies suggested to represent tar
33 with two lumped compounds of primary tar (acetol and catechol) [6, 16] or
34 by three lumped compounds (acetol, toluene) [5]. Palma et al. [6] suggested
35 that the most abundant tar compounds in wood gasification were catechol, *o*-
36 cresol and salicylaldehyde from lignin. The kinetic model of Jess [17] showed
37 that benzene is the key component of thermal decomposition of aromatic
38 hydrocarbons, and naphthalene is a precursor of soot formation. In order to
39 reduce the yield of soot during gasification, it is necessary to understand how
40 tar composition and yield are correlated with fuel composition and operating
41 conditions.

42 The formation of tars depends strongly on the operational conditions of
43 gasification [18]. A number of studies investigated the effect of heating rate,
44 temperature and pressure on the yields and composition of primary tar under

45 fast pyrolysis conditions. The large particle sizes and increasing pressure led
46 to the tar yield decrease in wood and cellulose pyrolysis [8, 19]. Furthermore,
47 the yields and average molecular weight of tars from pyrolysis of cellulose,
48 lignin and pinewood decreased with lowering heating rates and increasing al-
49 kali content (potassium and sodium) [20, 21]. The proportion of heavier tars
50 in pinewood pyrolysis increased with the higher temperatures and longer resi-
51 dence time due to the enhanced polymerization reactions between aromatic
52 tar compounds to form larger PAHs [22].

53 Despite of numerous previous studies on fast pyrolysis of various feed-
54 stocks at high temperatures [22–25], few studies, have systematically inves-
55 tigated how the chemical and structural variance of biomass affects the tar
56 yield and composition. It is difficult to identify how the tar yield and compo-
57 sition are affected by the origin of the feedstock. Therefore, a basic study of
58 fast pyrolysis using major biomass components is important, and beneficial
59 for the optimization of industrial processes such as entrained-flow gasifiers.
60 The specific objectives of this study were: (1) to obtain knowledge about
61 lignin type (softwood and wheat straw), and holocelluloses influence on the
62 tar and soot yields during high-temperature fast pyrolysis, (2) to relate the
63 product distribution of tar compounds to the soot yields at different resi-
64 dence times and temperatures, and (3) to quantify as many components of
65 tar samples as possible using GC-FID and GC-MS techniques.

66 **2. Materials and methods**

67 The effects of lignocellulosic compounds and potassium on the product
68 yields and composition were investigated in the drop tube reactor as de-

69 scribed previously [26]. Two types of organosolv lignin made from softwood
70 and wheat straw (purity > 94 %) were provided by BOC Sciences. Cellulose
71 Avicel[®] (purity > 99.9 %) and xylan from beechwood (purity > 90 %) were
72 supplied by Sigma-Aldrich. The purity of xylan was additionally improved
73 from 90 % to 96.6 % by a three step procedure involving a strong alkali treat-
74 ment, bleaching and acetylation. The effect of potassium on the product
75 yield and composition was investigated by wheat straw lignin impregnation
76 with potassium nitrate. The lignocellulosic compounds were reacted in the
77 drop tube reactor (DTF) at temperatures of 800-1250°C. The quantitative
78 analysis of tar compounds which were identified by GC-MS was performed
79 using GC-FID. The response factors for the known tar compounds were de-
80 termined.

81 *2.1. Original fuel characterization*

82 The ultimate and proximate analysis of the lignocellulosic compounds
83 was carried out at University of Agder, Department of Science and Engineer-
84 ing and shown in Table 1.

Table 1: Proximate, ultimate and ash analyses of cellulose, xylan from beechwood (hemicellulose), lignin from softwood and lignin from wheat straw.

	Cellulose	Xylan from beechwood	Lignin from softwood	Lignin from wheat straw
Proximate and ultimate analysis (% on dry basis)				
Moisture ^a	6	6	4.1	3.8
Ash (550 °C)	0.3	3.7	1.3	3.6
Volatiles	94.9	81.6	67.3	66.3
HHV ^b	18	14	26.4	26.7
LHV ^b	16.1	12.2	24.9	25.2
C	42.3	39.6	59.9	61.1
H	6.3	6	5.5	5.6
O	50.9	52.3	31.9	28.2
N	0.3	0.2	1.2	0.8

^a wt. % (as received) ^b in MJ kg⁻¹

85 The ash content of lignocellulosic compounds was determined using a
86 standard ash test at 550°C, according to the procedure described in DIN EN
87 14775. Lignin from wheat straw was rich in Na, Si and Fe (Na: 0.7 wt.%
88 of the dry material; Si: 0.4 wt.% of the dry material; Fe: 0.1 wt.% of the
89 dry material), whereas lignin from softwood contained a smaller fraction of
90 Na (0.4 wt.% of the dry material). Xylan from beechwood after purification
91 contained low amounts of Na and Ca (Na: 2 wt.% of the dry material; Ca:
92 0.6 wt.% of the dry material), as shown in supplementary Table S-1. The
93 effect of potassium on the product yield and composition was investigated
94 by wheat straw impregnation with potassium nitrate (Alfa Aesar, purity
95 > 99%), which was diluted in deionized water and added to 50 mg lignin.
96 The impregnated lignin had a similar potassium content to the wheat straw
97 (1.9%, db) [27]. The addition of alkali nitrates did not affect the carbon
98 balance compared to the impregnation with alkali carbonates, and nitrates

99 can be easily decomposed by a simple increase of temperature [28].

100 *2.1.1. ^{13}C solid state NMR spectroscopy*

101 Solid-state NMR analysis was carried out on a Bruker Avance 400 NMR
102 spectrometer (9.4 T) operating at Larmor frequencies of 400.13, 100.58 and
103 79.48 MHz for ^1H and ^{13}C respectively. All experiments were conducted using
104 a double resonance probe equipped with 4 mm (o.d.) rotors. Samples were
105 analyzed at room temperature by single-pulse (SP) magic angle spinning
106 (MAS) as well as cross polarization (CP) MAS [29] utilizing high-power ^1H
107 two-pulse phase-modulated decoupling (TPPM) [30] during acquisition and
108 employing a spin-rate of 9 kHz. The ^{13}C CP/MAS spectra were recorded
109 using a recycle delay of 8 s, a contact time of 1 ms, an acquisition time of
110 45.9 ms and 4096 scans, whereas the ^{13}C SP/MAS spectra were recorded using
111 a recycle delay of 128 s, an acquisition time of 45.9 ms and 1080 scans. All
112 ^{13}C NMR spectra were referenced to the carbonyl resonance of an external
113 sample of α -glycine at 176.5 ppm.

114 *2.1.2. FTIR Spectroscopy*

115 The lignin samples were analyzed by a Thermo Nicolet 6700 FTIR spec-
116 trometer equipped with a Golden gate (diamond) ATR accessory and DTGS
117 (KBr) detector. All absorbance spectra were obtained in the 4000-600 cm^{-1}
118 range by 100 scans at 4 cm^{-1} resolution. For background, 200 scans were
119 acquired. A good contact between sample and ATR-crystal surface was en-
120 sured by the pressure device of the unit (up to 30000 psi) [31]. All samples
121 were measured in triplicate.

122 *2.2. Fast pyrolysis in drop tube reactor*

123 The lignocellulosic compounds were reacted at three particle residence
124 times (800°C: 0.12, 0.25, 0.5 s; 1000°C: 0.1, 0.2, 0.42 s; 1250°C: 0.09, 0.17,
125 0.35 s) in a laminar drop tube reactor. The DTF setup was described in
126 detail by Trubetskaya et al. [26]. The residence time was varied by using dif-
127 ferent reactor lengths (0.72, 1.06 and 2.12 m), while the flow rate of feed gases
128 was kept constant. Reaction products were separated into coarse particles
129 (mainly char and fly ashes), fine particles (mainly soot and ash aerosols), per-
130 manent gases, and tars. Soot particles passing the cyclone (cut size 2.5 μm)
131 were captured from the product gas flow by a grade QM-A quartz filter
132 with a diameter of 50 mm (Whatman, GE Healthcare Life Science). The
133 tars from the product gas flow were collected in three serially connected gas
134 washing bottles, each of which was filled with 30 ml of methanol (HiPer-
135 Solv CHROMANORM[®], purity > 99.8%). The temperature of methanol
136 was kept at -50°C by the cooling bath Proline Edition X (LAUDA Dr. R.
137 Wobser GmbH, Germany).

138 *2.3. Tar analysis*

139 For the semiquantification of annotated substances, 5 μl of an internal
140 standard (Chlorobenzene, Sigma-Aldrich) was injected in the whole volume
141 of tars dissolved in methanol. Prior to the GC-FID analysis, a 1.5 ml aliquot
142 was pipetted into the autosampler screw cap vial and stored in the freezer
143 at -20°C. The quantitative analysis of tar compounds was performed on
144 a gas chromatograph 7820A (Agilent Technologies, USA) equipped with a
145 flame ionization detector (GC-FID) and DB-EUPAH capillary column (30 m

length, 0.25 mm internal diameter, 0.25 μm film thickness). The temperatures of the injector and detector were kept at 250°C and 300°C, respectively. The column temperature program ran from 50 to 280°C. After holding the oven temperature at 50°C for the first 2 min the temperature was increased to 160°C at a rate of 1.5°C min⁻¹, then to 230°C at a rate of 6°C min⁻¹, and then to 280°C at a heating rate of 8°C min⁻¹ and before it was hold for another 5 min. Nitrogen was used as a carrier gas with a constant flow rate of 1 ml min⁻¹. Data acquisition and processing were performed using Agilent OpenLAB CDS EZChrom A.02.02 (Agilent Technologies, USA). Certain species were calibrated at four levels with solutions of known concentration and 5 replicates per level. Prior to the quantitative analyses in GC-FID, the tar compounds were annotated using a dual detector system GC-MS 5975C TAD Series / GC-FID 7890A (Agilent Technologies, USA). The column temperature and carrier gas settings were kept the same as those used in GC-FID analysis. The mass spectrometer with a quadrupole type analyzer scanned the range from m/z 35 to m/z 250 resulting in a scan rate of 6.22 scans s⁻¹. The mass spectrometer was operated at unit mass resolution. A 0.5 μl of sample was injected at a 4:1 split ratio. The collected spectra were exported from Chemstation E.02.00.493 (Agilent Technologies, USA) to NetCDF and further processed by the statistical software "R" 2.15.2 [32] that can acquire and align the data, correct baseline, set time-window and perform multivariate analysis [33]. The multivariate analysis using MCR-AR algorithm yielded deconvoluted mass spectra with the well-resolved overlapping peaks [34], which were imported into the mass spectra library software NIST MS Search 2.0 [35]. The area of peaks was normalized to 100% within

171 each sample and the mean of triplicate measurements was calculated. The
172 peaks with mass spectra similarity higher than 80% were used in the tar
173 quantification. The relative response factors (RRFs) were determined for
174 each compound in tar samples using MatLab (version 8.6, MathWorks Inc.).

175 *2.4. Multivariate data analysis*

176 Prior to multivariate analysis, the resulting intensities of all tar com-
177 pounds were normalized to the intensity of the internal standard. Multivari-
178 ate analysis was carried out using the SIMCA software (Umetrics AB, version
179 14.0) to access the effects of different lignocellulosic compounds and residence
180 time using 0.72 m- and 1.06 m-long reactor tube on the composition of tar
181 samples, which were grouped in carbon dioxide, aliphatic hydrocarbons and
182 their derivatives (paraffins, olefins, acetylene, alkanenitriles, cycloolefins),
183 aromatic hydrocarbons (toluene, benzene, ethylbenzene, styrene, etc.), aro-
184 matic nitrogen-containing hydrocarbons, triophenes, oxygenates (ester, alco-
185 hol, aldehyde), furans, naphthalenes, polyaromatic hydrocarbons, and un-
186 known species. Orthogonal Projections to Latent Structures Discriminant
187 Analysis (OPLS-DA) is a development of partial least squares (PLS) with
188 the aim to increase the interpretability of models by separating the variation
189 that is related to the response from the variation that is unrelated to the
190 response. Using the same settings, the prediction properties are the same for
191 PLS and OPLS [36]. Both methods involve the construction of a regression
192 model maximizing the covariance between the descriptor variables (X) and
193 the response, i.e. the dependent variable, y. Additionally, OPLS performs a
194 filtering step, which captures structured variation not related to the response
195 but overlapping with the related variation, in one or more orthogonal compo-

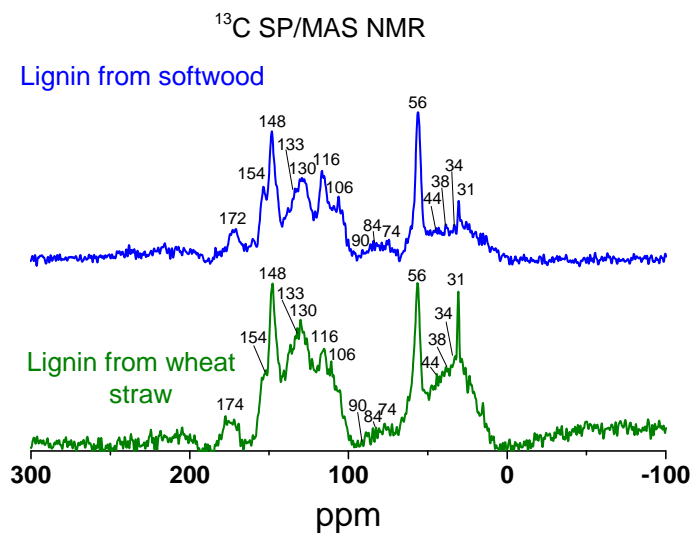
196 nents. The loadings of the descriptor variables on those components indicate
197 the origin of the uncorrelated, also called orthogonal, variation. The varia-
198 tion correlated with the response can be interpreted by the loadings on the
199 predictive component of the OPLS model. OPLS-DA has been largely used
200 in the -omics context, and it is now the multivariate linear model of choice
201 for classification/discrimination [37, 38]. The term classification is used when
202 the objective is to classify new objects into one of two or more possible classes
203 (e.g. cellulose, hemicellulose, softwood lignin, wheat straw lignin). The term
204 discrimination is used for the two-class case, in which the objective is to sep-
205 arate two classes and investigate the causes for class separation. In OPLS
206 the vector y is a continuous variable; in two-class discrimination, OPLS-DA
207 y is categorical and, thus, defined as a dummy vector of 0/1 for the two-class
208 case (for the multiple- y case, it is a dummy matrix with a 0/1 vector per
209 class), describing class belonging. R^2X values of the predictive and orthogo-
210 nal components are measures of the structured fraction of the original data
211 variation describing the response and the fraction not correlated with the
212 response. The quality of an OPLS-DA model is described by the R^2Y value,
213 i.e. the correlation between the observed and predicted values for the studied
214 response, and the Q^2 value, i.e. the correlation between the observed and
215 cross-validated predicted response. The higher the R^2Y and Q^2 value the
216 better the response can be described and predicted as a function of the de-
217 scriptor variables, respectively. The R^2X , R^2Y , and Q^2 values are normalized
218 to have an upper limit of 1. The low end is around 0 but the results of the
219 cross validation may cause Q^2 to be negative when no model is found [39].
220 The confidence level was set at 95 %. The model accuracy is evaluated by the

221 goodness of fit (R^2Y) for the X matrix and goodness of prediction by cross
222 validation (Q^2) [40]. Both R^2Y and Q^2 parameters vary in the range from
223 0 to 1, with the upper limit of 1 for a perfect fit and 0.5 for an acceptable
224 fit [41]. The number of components in all models was chosen using the aut-
225 ofit approach in SIMCA software that maximizes the relevant Q^2 statistics,
226 while ensuring that all calculated components are significant and stable to
227 resampling. The output of OPLS-DA model was visualized using both score
228 scatter and loading plots. Each point in the score scatter plot represents
229 a tar sample from pyrolysis of cellulose, hemicellulose, softwood lignin and
230 wheat straw lignin using 0.72 m-long or 1.06 m-long reactor tube. Each col-
231 umn in the loading plot corresponds to a GC-MS peak colored according to
232 its identified class of chemical structure.

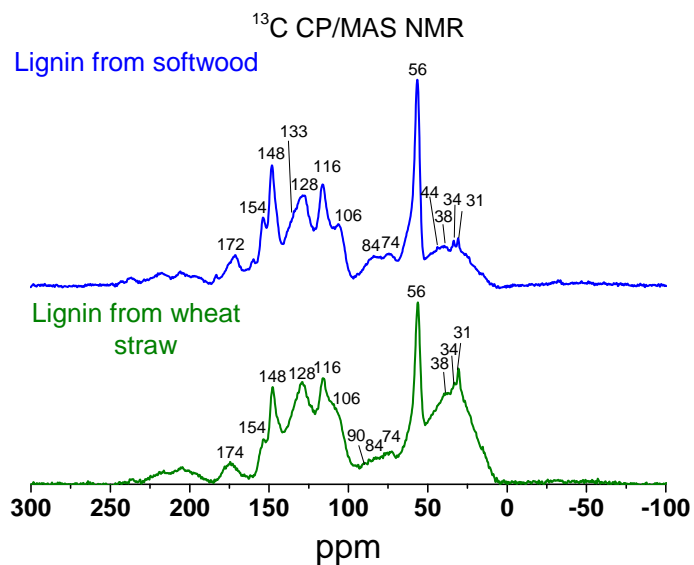
233 **3. Results and discussion**

234 *3.1. Lignin characterization*

235 The effect of lignin type on the organic matter transformation, which
236 affects product yields in high-temperature pyrolysis, was investigated, us-
237 ing ^{13}C CP/MAS and ^{13}C SP/MAS NMR. In the ^{13}C CP/MAS experi-
238 ments the resonances of the carbons in immobile regions of the samples were
239 enhanced by polarization transfer from the highly abundant ^1H nuclei via
240 hetero-nuclear dipolar coupling.



1(a): ^{13}C CP/MAS NMR



1(b): ^{13}C SP/MAS NMR

Figure 1: ^{13}C CP/MAS and ^{13}C SP/MAS spectra of lignin from softwood and lignin from wheat straw.

241 All carbon sites were observed quantitatively by the ^{13}C SP/MAS NMR
 242 measurements. In Figure 1, the ^{13}C CP/MAS and ^{13}C SP/MAS NMR spectra
 243 of lignin from softwood and lignin from wheat straw are displayed, and an
 244 assignment of resonances is shown in Table 2.

Table 2: Resonance assignment of ^{13}C CP/MAS and ^{13}C SP/MAS NMR spectra of lignin from softwood and lignin from wheat straw [42–46].

Chemical shift, ppm	Description
172-174	Carbohydrate; -COO-R, CH ₃ -COO-
153-154	Lignin; S3(e), S5(e)
145-148	Lignin; S3(ne), S5(ne), G1(e), G4(e)
133-138	Lignin; S1(e), S4(e), G1(e)
121-130	Lignin; G6
105-116	Carbohydrates; C1, Lignin; S2, S6, G5, G6
89-92	C4 in cellulose (cr)
84-85	C4 in cellulose (am)
72-75	C2, C3 in carbohydrates; C5 in cellulose
56-57	Lignin, OCH ₃
30-38	CH ₂ in aliphatics

Abbreviations: S, syringyl; G, guaiacyl; ne, in non-etherified arylglycerol β -aryl ethers; e, in etherified arylglycerol β -aryl ethers.

245 The ^{13}C CP/MAS and ^{13}C SP/MAS NMR spectra of both lignin types
 246 are almost identical. The primary difference in the structure of both lignins
 247 was in the signal at 31 ppm (methyl group in aliphatic chains) and 106 ppm.
 248 According to the specifications of the producer, both organosolv lignin sam-
 249 ples contained < 6% residual carbohydrates. The resonances in a range of
 250 72-92 ppm and 106 ppm are most likely due to overlapping resonances from
 251 C1-C4 in cellulose and aromatic carbons in lignin. The signal at 106 ppm was
 252 more recognizable in NMR spectra of lignin from softwood probably due to

253 the greater fraction of carbohydrates remaining in the lignin after organosolv
254 treatment. The more visible signal at 31 ppm was associated with the pres-
255 ence of p-hydroxyphenol unit in lignin from wheat straw [47]. Comparison
256 of the peak intensities in the region 84-90 ppm of organosolv lignin indicates
257 that the cellulose content in organosolv lignin is low compared to Protobind
258 lignins [48]. Figure 2 shows the IR spectra of lignin from softwood and lignin
259 from wheat straw.

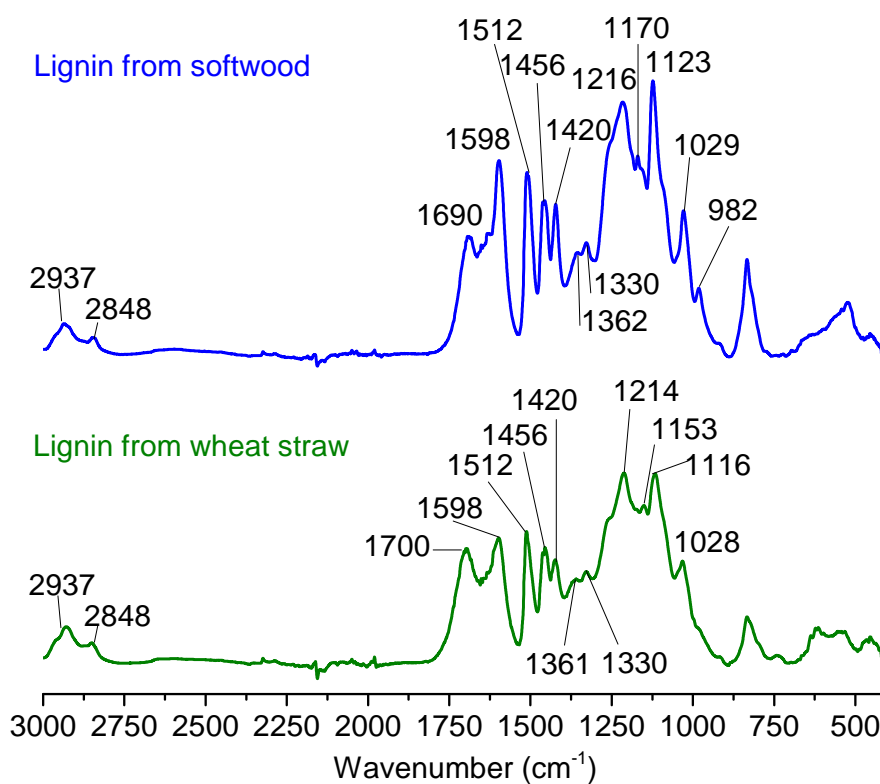


Figure 2: Experimental IR spectra of lignin from softwood and lignin from wheat straw.

260 The assignment of species to each IR band is shown in supplemental ma-

261 terial (Table S-2) using literature data [49–54]. A strong signal was observed
262 for the lignin from softwood at 1598 and 1512 cm^{-1} which were assigned to the
263 C=C group stretching and suggesting the higher content of aromatic groups
264 than in the lignin from wheat straw. The organosolv process results in cleav-
265 age of the β -O-4-linkages which generate both phenolic hydroxyl (1365 cm^{-1})
266 and carbonyl groups (1700 cm^{-1}) [55]. Both lignin samples were character-
267 ized as guaiacyl-syringyl GS type, confirming previous results of Labidi et
268 al. [56]. Moreover, the whole C=O range between 1800-1633 cm^{-1} is intense.
269 Because the C=O vibrations also cause a band around 1270 cm^{-1} , the ab-
270 sorbance here is higher for the lignin from wheat straw than in the case of GS
271 spectra of lignin from softwood [50]. A small band around 1170 cm^{-1} (C=O
272 vibration of esters) is additionally present in the lignin from softwood that
273 is typically to find in the GS lignin type. The IR results indicated that the
274 organosolv process has no significant influence on the lignin structure.

275 3.2. Identification of tar compounds

276 The identification of individual tar compounds from GC-MS analysis was
277 confirmed further in comparison with the reference chromatograms of exter-
278 nal standards and literature results [10, 57–61]. The identified tar compounds
279 with the relevant information were listed in Table 3. Forty five compounds in
280 the pyrolysis tar have been quantified and grouped for the further modeling
281 using SIMCA.

Table 3: List of identified tar compounds with the empirical formula, molecular weight, retention time and compound category.

No	Compounds	Formula	MW	RT	Cat. name
				min	
1	Acetonitrile	C ₂ H ₃ N	41.05	6.18	AH
2	Benzene	C ₆ H ₆	78.11	6.30	A
3	Toluene	C ₇ H ₈	92.14	9.05	A
4	2-Methylthiophene	C ₅ H ₆ S	98.17	9.89	T
5	3-Methylthiophene	C ₅ H ₆ S	98.17	10.29	T
6	Pyrrole	C ₄ H ₄ NH	67.09	10.74	AN
7	Ethylbenzene	C ₈ H ₁₀	106.17	15.90	SA
8	Styrene	C ₈ H ₈	104.15	17.35	SA
9	1,2,4-Trimethylbenzene	C ₉ H ₁₂	120.19	23.00	SA
10	α -Methylstyrene	C ₉ H ₁₀	118.18	23.70	SA
11	Dimethyl Malonate	C ₅ H ₈ O ₄	132.12	24.27	SA
12	1,2,3-Trimethylbenzene	C ₉ H ₁₂	120.19	26.38	SA
13	Phenol	C ₆ H ₅ OH	94.11	26.46	P
14	Benzaldehyde	C ₇ H ₆ O	106.12	26.86	O
15	2-Methylphenol	C ₇ H ₈ O	108.14	32.87	P
16	1-Phenyl-1-propyne	C ₉ H ₈	116.16	33.56	SA
17	3-Methylphenol	C ₇ H ₈ O	108.14	34.81	P
18	Phenyl acetate	C ₈ H ₈ O ₂	136.1	35.13	O
19	3-Methylbenzofuran	C ₉ H ₈ O	132.16	37.35	F
20	2-Methylbenzofuran	C ₉ H ₈ O	132.16	37.60	F
21	2,6-Dimethylphenol	C ₈ H ₁₀ O	122.17	38.12	P
22	Naphthalene	C ₁₀ H ₈	128.17	46.85	N
23	4-Vinylphenol	C ₈ H ₈ O	120.15	49.84	P
24	Quinoline	C ₉ H ₇ N	129.16	55.08	AN
25	1-Methylnaphthalene	C ₁₁ H ₁₀	142.2	58.44	SN
26	1-Indanone	C ₉ H ₈ O	132.16	60.61	SA
27	Indole	C ₈ H ₇ N	117.15	62.11	AN
28	Biphenyl	C ₁₂ H ₁₀	154.21	64.78	PAH
29	2,6-Dimethylnaphthalene	C ₁₂ H ₁₂	156.22	64.98	SN
30	1,3-Dimethylnaphthalene	C ₁₂ H ₁₂	156.22	67.01	SN
31	Biphenylene	C ₁₂ H ₈	152.19	71.55	PAH
32	Acenaphthene	C ₁₂ H ₁₀	154.2	75.02	PAH

No	Compounds	Formula	MW	RT	Cat. name
33	Dibenzofuran	C ₁₂ H ₈ O	168.19	77.01	F
34	Fluorene	C ₁₃ H ₁₀	166.22	80.70	PAH
35	2-Methylfluorene	C ₁₄ H ₁₂	180.25	84.62	PAH
36	4-Methylfluorene	C ₁₄ H ₁₂	180.25	85.48	PAH
37	Phenanthrene	C ₁₄ H ₁₀	178.23	88.05	PAH
38	Anthracene	C ₁₄ H ₁₀	178.23	88.22	PAH
40	1-Methylphenanthrene	C ₁₅ H ₁₁	192.26	90.98	PAH
41	4H-Cyclopenta[def]phenanthrene	C ₁₅ H ₁₀	190.25	91.08	PAH
42	2-Phenylnaphthalene	C ₁₆ H ₁₂	204.27	91.64	PAH
43	Fluoranthene	C ₁₆ H ₁₀	202.26	92.09	PAH
44	Pyrene	C ₁₆ H ₁₀	202.25	92.99	PAH
45	2-Methylpyrene	C ₁₇ H ₁₂	216.28	97.59	PAH

A: Aromatic compounds

AH: Aliphatic hydrocarbons

AN: Aromatic nitrogen-containing compounds

F: Furans

N: Naphthalenes

O: Oxygenates

PAH: Polyaromatic compounds

P: Phenols

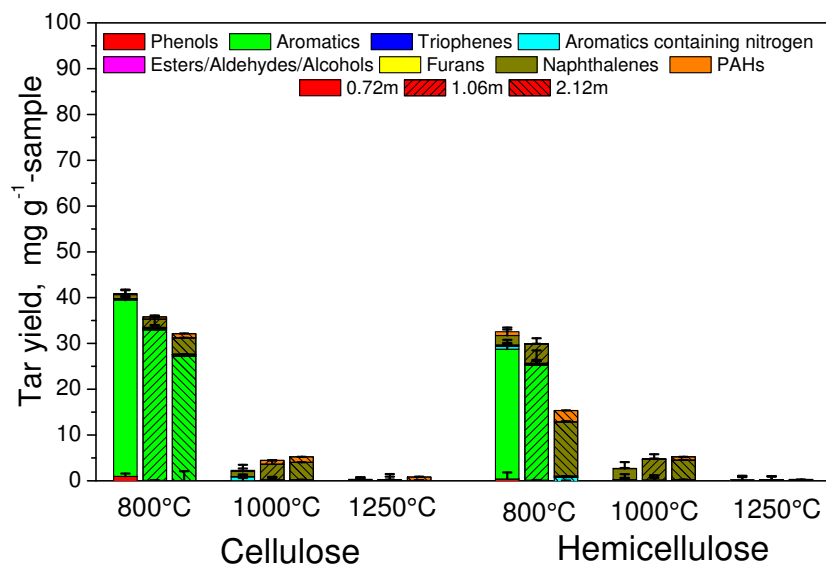
SA: Substituted aromatic compounds

SN: Substituted naphthalenes

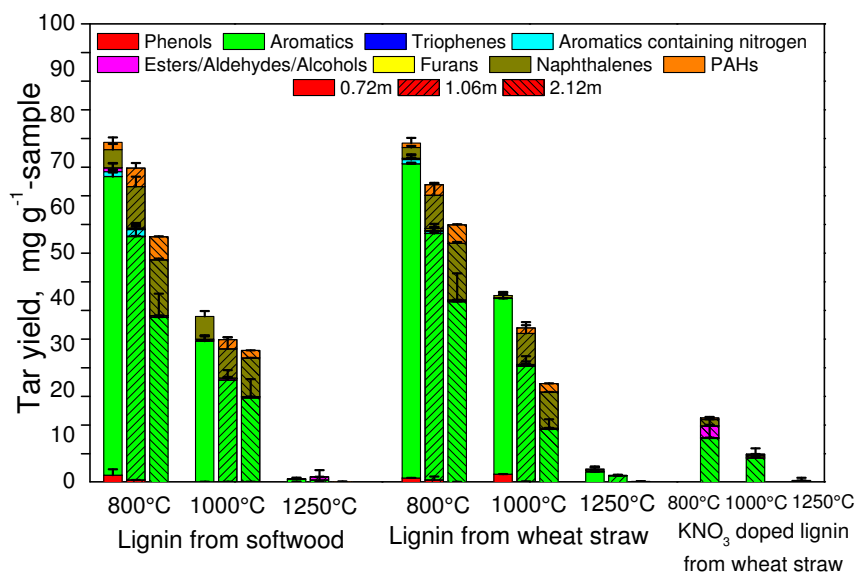
T: Triophenes

282 3.3. Tar and soot yields

283 Figure 3 shows the change in the yields of tar from pyrolysis of cellulose,
284 hemicellulose, lignin from softwood, lignin from wheat straw, and KNO₃
285 doped lignin from wheat straw in the range from 800 to 1250°C at three
286 particle residence times. The pyrolysis of lignin gave a greater tar yield
287 compared to pyrolysis of cellulose and hemicellulose. The differences in the
288 tar yield from pyrolysis of lignin from softwood and lignin from wheat straw
289 were small.



3(a): Cellulose and Hemicellulose



3(b): Lignin

Figure 3: Tar yields (mg g⁻¹ on dry basis) of cellulose, hemicellulose, lignin from softwood, lignin from wheat straw, and KNO₃ doped lignin from wheat straw.

290 In general, the yield of identified tar decreased with the increased heat
291 treatment temperature and with the increased residence time. The addition
292 of alkali nitrates to the lignin from wheat straw led to the decrease in tar
293 yields. The high level of alkali metals in biocarbon catalyzes the conversion
294 of bridges into char, promoting faster devolatilization rates and suppressing
295 tar formation, leading to greater char yields [62, 63]. The yields of phenols
296 and aromatic compounds decreased and the yields of naphthalene and PAHs
297 increased with an increase in heat treatment temperature from 800 to 1000°C
298 in pyrolysis of all lignocellulosic compounds. More esters, aldehydes and al-
299 cohols were formed with the addition of alkali nitrates to lignin from wheat
300 straw at 800°C due to the function of alkali metals to break the aromatic
301 rings [64]. Figure 4 illustrates that the soot yields from pyrolysis of lignocel-
302 lulosic compounds was the greatest at 1250°C, whereas the formation of tars
303 was the lowest under similar operating conditions. The greatest soot yield
304 (daf) was obtained from pyrolysis of lignin due to the stronger formation
305 of PAH precursors. The conversion of tars to the light aromatic compounds
306 could contribute to the greater soot yields with the longer residence time and
307 increased heat treatment temperature. The results showed that significantly
308 less soot was formed during pyrolysis of cellulose and hemicellulose due to
309 the lower fraction of phenolic groups. The formation of soot was not affected
310 by the differences in lignin composition. During pyrolysis of KNO₃ doped
311 lignin from wheat straw, the soot and tar yields decreased with the increased
312 heat treatment temperature.

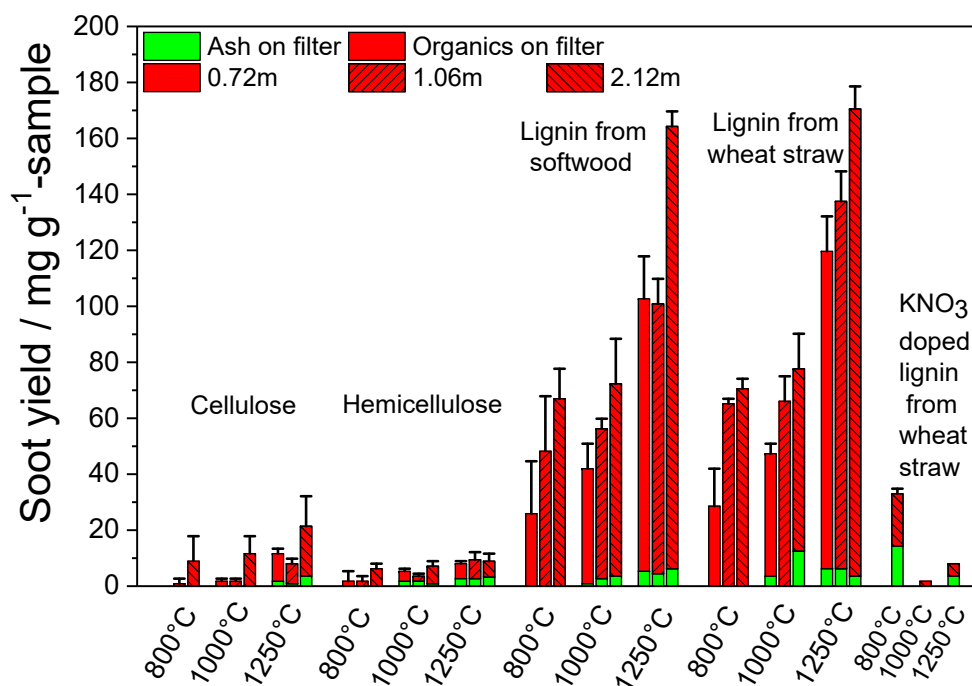


Figure 4: Soot yields (mg g^{-1} on dry basis) of cellulose, hemicellulose, lignin from softwood, lignin from wheat straw, and KNO_3 doped lignin from wheat straw.

313 *3.4. Modeling of tar yields and composition*

314 The OPLS-DA score scatter plot in Figure 5 shows that tar samples
 315 from pyrolysis of cellulose, hemicellulose, softwood lignin and wheat straw
 316 lignin using 0.72 m- and 1.06 m-long reactor tube were well separated into two
 317 clusters. The majority of variables related to tar samples generated in 0.72 m-
 318 long reactor tube appeared in two left quadrants, whereas the majority of
 319 tar samples from pyrolysis in 1.06 m-long tube reactor was plotted in two
 320 right quadrants in the score scatter plot. This indicated a good separation of
 321 model compounds and emphasized the importance of residence time on the

322 tar composition.

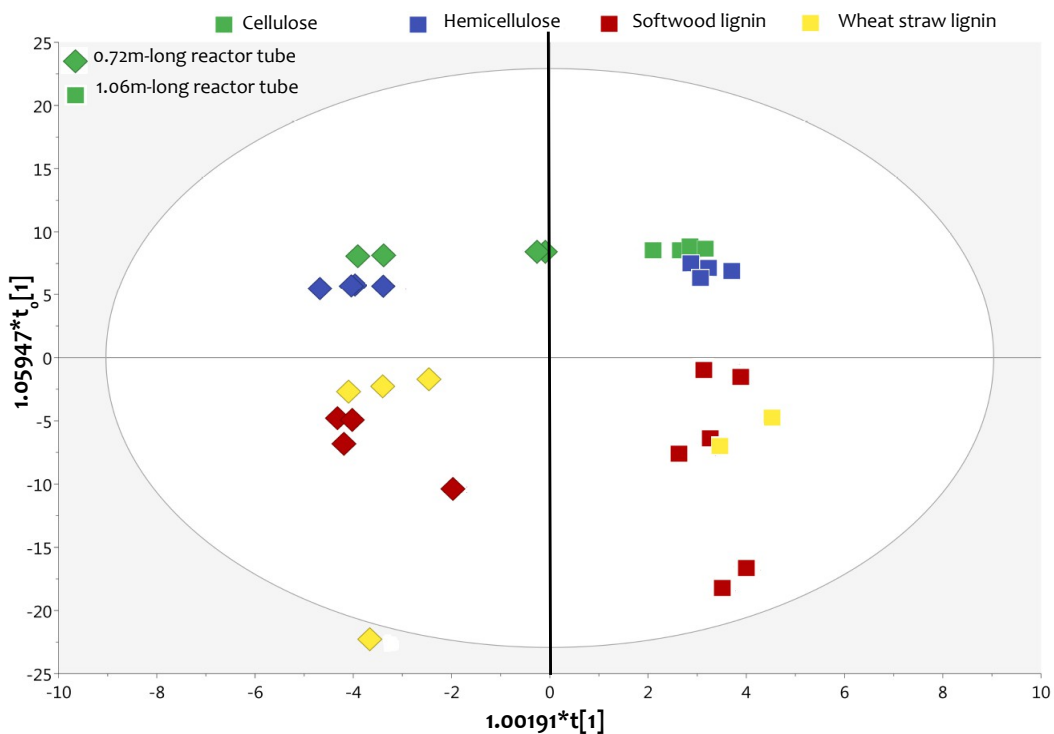


Figure 5: The OPLS-DA score scatter plot ($R^2Y = 0.91$, $Q^2 = 0.85$) displays the clustering of tar samples from pyrolysis of cellulose, hemicellulose, softwood lignin and wheat straw lignin using 0.72 m- and 1.06 m-long reactor tube. Tar samples produced at the same residence time appear to group together. The tar composition of both lignin samples is significantly different from the tar composition of holocelluloses at both residence times.

323 The OPLS-DA column loading plot in Figure 6 confirms that the resi-
324 dence time has a strong influence on the tar composition. Single ring com-
325 pounds such as aliphatic hydrocarbons, phenols, aromatics, oxygenates were
326 more abundant in pyrolysis using 0.72 m-long reactor tube. The 2-4 ring

327 polyaromatic hydrocarbons (PAHs) and nitrogen-containing aromatics were
328 mostly formed in pyrolysis using 1.06 m-long reactor tube, confirming pre-
329 vious results of Onwudili et al. [65]. The low concentrations of large PAH
330 with aliphatic chains might be related to the formation of more stable prod-
331 ucts or soot with increasing residence time [66]. The aromatics with attached
332 aliphatic chains show significantly higher nucleation rates than the polycyclic
333 aromatic hydrocarbons of similar mass without any chain [67]. The concen-
334 trations of benzene and toluene which belong to dominating compounds in tar
335 samples from all lignocellulosic compounds remained unchanged, while the
336 amount of triophenes was similarly small with increasing residence time. The
337 yield of phenanthrene exhibited an order of magnitude higher concentration
338 than anthracene, while the concentrations of phenanthrene and anthracene
339 were almost twice as low as that of naphthalene and its derivatives, confirm-
340 ing the previous results of Kislov et al. [68]. The OPLS-DA column loading
341 plot 6 shows that a small fraction of naphthalene molecules which were not
342 converted to larger PAHs, formed substituted structures with one or two
343 ethyl or methyl groups with increasing residence time. Numerous nitrogen-
344 containing aromatic compounds were formed at long residence time. The
345 OPLS-DA score scatter plot in Figure 5 shows that tar samples from cellulose
346 and hemicellulose pyrolysis appeared to group tightly together in the upper
347 quadrants, whereas both lignin tar samples were clustered in the lower quad-
348 rants. This demonstrated significant differences in tar composition among
349 holocelluloses and lignin. The dominating products derived from pyrolysis of
350 softwood lignin were phenols, aromatic compounds, naphthalenes and PAHs,
351 whereas the oxygenates were mostly present in tar samples from hemicellu-

352 lose pyrolysis (supplementary Figures S-6 and S-7).

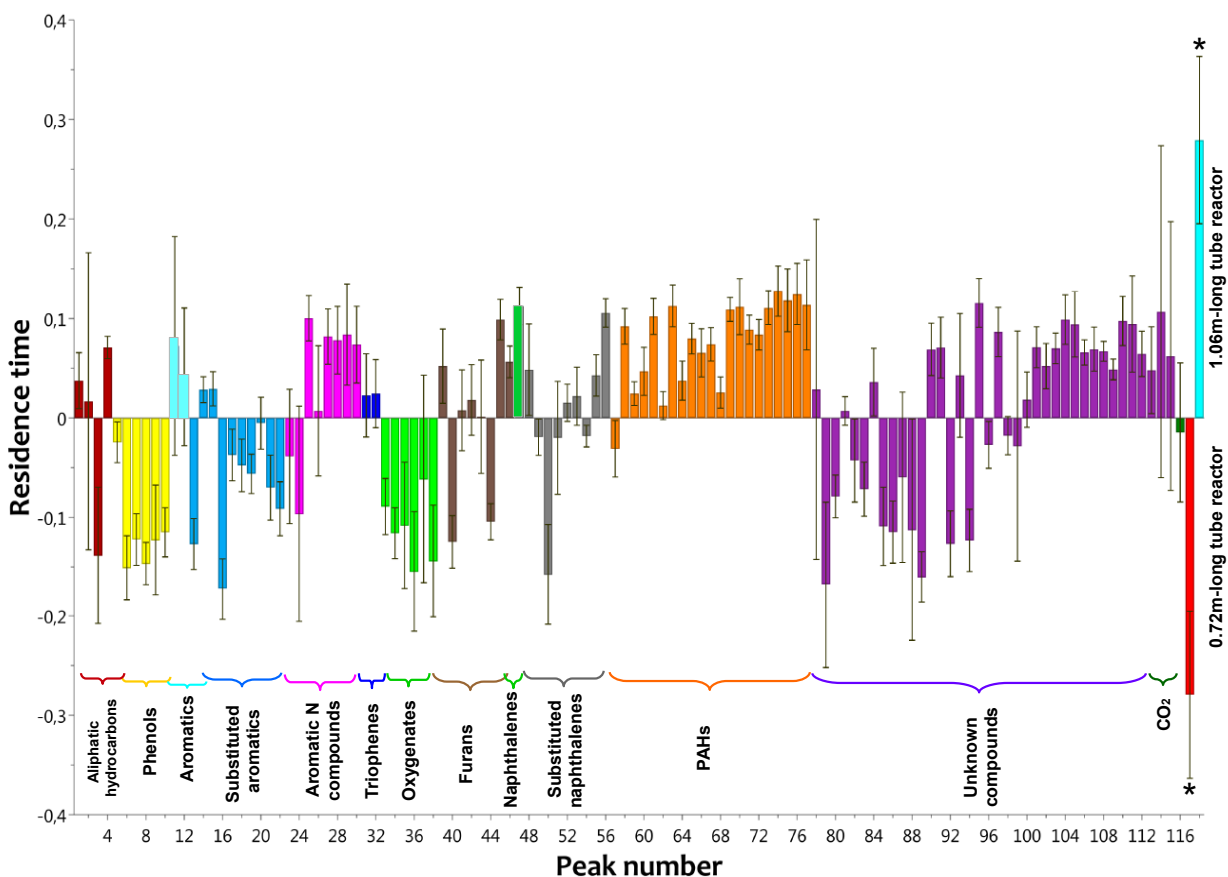


Figure 6: The OPLS-DA column loading plot ($R^2Y = 0.91$, $Q^2 = 0.85$) visualizes differences in the composition of tar samples generated in pyrolysis using 0.72 m- and 1.06 m-long reactor tube. The value of variables pointing in the same direction as columns of tar compounds from pyrolysis using 0.72 m-long reactor tube is higher in that group than the value of variables pointing in the opposite direction. The higher the column and smaller the error bar, the greater the contribution of the variable to the model [69]. Error bars denote the 95 % confidence level.

353 The tar composition within the lignocellulosic compound class (holo-

354 celluloses or lignin) remained nearly similar, as shown in the supplemental
355 material (Figures S-3, S-5, S-6 and S-7). More phenols, aromatic compounds,
356 triophenes, furans, naphthalenes, substituted naphthalenes, substituted aro-
357 matics and PAHs were observed in hemicellulose tars than in products from
358 cellulose pyrolysis due to the lignin-associated impurities in hemicelluloses
359 (supplementary Figure S-3). Interestingly, the cellulose tars contained the
360 high concentration of acetic acids which were probably formed from the de-
361 composition of 5-hydroxymethylfurfural [70].

362 The present OPLS-DA model ($R^2Y = 0.91$, $Q^2 = 0.85$) can explain
363 most of the variations within the data set and can predict well the sedi-
364 ment age from the GC-MS data set. The OPLS-DA model for tar samples
365 from softwood lignin and wheat straw lignin explained 61 % of variation in
366 the tar composition, but was less reliable ($Q^2 = 0.21$) than the OPLS-DA
367 models for cellulose/hemicellulose ($R^2Y = 0.97$, $Q^2 = 0.99$) and hemicellu-
368 lose/softwood lignin tar compounds ($R^2Y = 0.89$, $Q^2 = 0.87$). The prediction
369 of the model for tar samples from softwood lignin and wheat straw lignin was
370 mostly affected by the systematic errors associated with the tar collection and
371 storage [71]. Further progress in the prediction of tar composition can be ad-
372 vanced by improvement in the accuracy of experimental measurements, as
373 well as prediction of tar composition using other advanced techniques. The
374 results showed that the precision of the lignin tar model can not be improved
375 by the variation of predictive and orthogonal components.

376 **4. Conclusion**

377 This work presents yields and composition of tars for lignocellulosic com-
378 pounds reacted in a drop-tube furnace operating in the temperature range
379 from 800 to 1250°C. The present results indicate that the dominating prod-
380 ucts in tars from pyrolysis of all lignocellulosic compounds were aromatics.
381 The minor differences in a tar composition were in a greater fractions of oxy-
382 genates in holocellulose pyrolysis, whereas more phenols, PAHs and naph-
383 thalene were observed in the lignin pyrolysis. No significant differences were
384 observed in the yield and composition of lignin from softwood and lignin from
385 wheat straw despite greater aromatic content in lignin from softwood. Longer
386 residence time and higher heat treatment temperatures led to the decrease
387 in tar yields and greater soot formation. Simultaneous reduction of tar and
388 soot was achieved by impregnation of lignin from wheat straw with aque-
389 ous KNO₃ solution, leading to reduction of light hydrocarbons. The present
390 OPLS-DA model can explain most of the variations within the experimental
391 GC-MS data set by showing that the moisture, oxygen and carbon content in
392 the original feedstock has more influence on the tar compositional differences
393 among the lignocellulosic compounds than the inorganic content with less
394 than 4.3% in the original feedstock.

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